

# Safety study of cultured human Wharton's Jelly mesenchymal stem cell therapy for multiple indications – a retrospective descriptive study

B.M. Mehling<sup>1</sup>, D.-C. Wu<sup>2</sup>, D. Santora<sup>1</sup>, E. O'Gorman<sup>1</sup>, K. Novakova<sup>1</sup>, G. Spencer<sup>3</sup>, C. Thomas<sup>3</sup>, R. Grant<sup>3</sup>, R. Rutherford<sup>4</sup>, R. Mihályová<sup>5</sup>

<sup>1</sup>BHI Therapeutic Sciences, Hackensack, NJ, USA

<sup>2</sup>Department of Biochemistry and Molecular Biology, Wuhan University School of Basic Medical Sciences, Wuhan, China

<sup>3</sup>Baywest Wellness Hospital and Clinics, Halfmoon Shopping Village, Rose Hall, Montego Bay, St. James, Jamaica

<sup>4</sup>The Ohio State University, Columbus, OH, USA

<sup>5</sup>Blue Horizon International, Bratislava, Slovakia

Corresponding Author: Brian M. Mehling, MD; e-mail: bmehling@bluehorizoninternational.com

**Keywords:** Cell therapy, Mesenchymal stem cells (MSCs), Regenerative Medicine, Wharton's jelly mesenchymal stem cells.

## Abstract

**Objective:** Human umbilical cord Wharton's Jelly derived MSCs (WJ-MSCs) are reported as the most potent cell source of mesenchymal stem cells, however they remain understudied in comparison to autologous sources. This study aimed to evaluate the safety of WJ-MSC therapy for a range of conditions and administration routines, including intravenous, intrathecal, and intraarticular delivery.

**Patients and Methods:** 22 subjects were retrospectively evaluated for adverse events following treatment with  $1 \times 10^8$  WJ-MSCs for a range of conditions including neurological and osteoarthritic indications. Subjects were treated at Blue Horizon International (BHI) clinical sites; 13 subjects at BHI Slovakia (Nemocnica Malacky hospital, Slovakia) and 9 subjects were treated at BHI Jamaica (Montego Bay St. James and Ochos Rios, Jamaica) between 2018 to 2021. Subjects received 1 – 3 dosages of WJ-MSC via intravenous, intrathecal, or intraarticular administration depending on the indication.

**Results:** Subjects were followed on for 6-months and the incidence of adverse events recorded. In total this study reported AEs in 3 subjects from the 32 doses administered in this study, resulting

in an AE rate of 9.3%. Reported AEs consisted of chills and headaches both transient and mild, and resolved without concern. Blood profiling of 75 markers for health and disease in a single subject reported that WJ-MSC treatment poses no hematological safety concern, with only one dietary related marker showing marked change during the follow up period. In contrast, several out of reference hematological markers were corrected 8-months after WJ-MSC therapy.

**Conclusions:** We highlight the minimal occurrence of adverse reactions, most commonly chills and headache associated with intrathecal administration, following WJ-MSC therapy. Overall, this study supports the use of WJ-MSC therapy for various indications and provides safety data for future prospective studies.

## INTRODUCTION

Mesenchymal stem/stromal cells (MSCs) are the leading candidate for regenerative medicine applications due to their established immunomodulatory ability, hypo-immunogenicity, regenerative capabilities and proven clinical safety. MSCs are multipotent, non-hematopoietic and have the capability for self-renewal and differentiation. MSCs can be isolated from many different tissues, most commonly bone-marrow, adipose, or umbilical cord (cord blood or Wharton's Jelly) origins<sup>1-4</sup>. The therapeutic



effect of MSC therapy arises largely through immunomodulatory repair mechanisms. MSCs detect and home to sites of inflammation and damage<sup>5,6</sup>, where they exhibit a dose-dependent antiproliferative effect on T and B lymphocytes, dendritic cells and natural killer cells<sup>7,8</sup>. The reduction in local inflammation at the injury site provides a suitable environment for repair, which is furthered by secretion of trophic factors from MSCs that promote cell survival<sup>9</sup>, angiogenesis<sup>10</sup> and accelerate tissue regeneration<sup>11</sup>. The pharmacokinetic characteristics and *in vivo* biodistribution of intravenous MSC delivery is well defined and non-toxic in many species, including dogs<sup>12</sup>, rat<sup>13</sup>, mice<sup>14</sup>, sheep<sup>15</sup>, and non-human primates<sup>5,16,17</sup>. These studies have characterized how MSCs distribute immediately under normal physiological conditions to the lungs after intravenous infusion, and later are found in the liver, spleen, bone marrow and kidneys<sup>13,14,17</sup>. If injury or inflammation is present, MSCs will subsequently migrate to the leading areas of damage, as identified in the injured muscle, skin, bone marrow, thymus and gut following total body irradiation in non-human primate<sup>5,16,17</sup>.

Safety is the paramount concern during the development of any new therapeutic intervention. Human MSC (hMSC) therapy presents an impeccable safety profile. A meta-analysis of 36 clinical trials of human MSC therapy reported no serious adverse reactions, including no association with acute toxicity, thromboembolism, abnormal cell growths, neurological deterioration, or death<sup>18</sup>. This study reported the only adverse reaction as transient fever occurring in a subset of patients to a particular preparation of MSCs, with no long-term health concern. Further studies exemplify the safety of hMSC therapy which is routinely used with no severe adverse reactions reported<sup>19-24</sup>. Further, hMSC therapy presents no health concerns over long term follow ups. A 5-year study post hMSC treatment in stroke patients has reported no significant adverse reactions observed in the treatment group versus the control<sup>25</sup>.

Blue Horizon International (BHI) has to date treated thousands of patients with hMSCs without any serious adverse events. Published research from BHI treatments continues to contribute to the expanding safety data supporting the use of hMSCs for a range of human conditions. We recently published a retrospective cohort study to evaluate the safety and efficacy of adipose tissue derived MSC therapy for osteoarthritis<sup>26</sup>. Evaluation of 350 patients receiving the therapy showed a significant

improvement in pain levels and mobility, and critically, reported no severe adverse events or complications. Prior, we published data on the safety of MSC-containing cord blood therapy in 30 patients with spinal cord injury<sup>27</sup>. No subjects developed adverse reactions, further demonstrating the safety of hMSC therapy. Our clinical trial data for autologous MSC therapy as a spinal cord injury treatment in 20 patients reported no severe adverse effects<sup>28</sup>. The most common adverse event, fever, and headache disappeared without treatment within 24 to 48 hours. To provide further safety and immunogenicity data, we characterized the blood profile and immune response in 29 patients receiving human umbilical cord blood derived MSC treatment for chronic inflammation<sup>29,30</sup>. Our data demonstrated that no essential changes in blood markers (including general health blood test panel and inflammatory markers) occurs following stem cell treatment and when followed up for 3 months.

The use of allogenic Wharton’s Jelly derived MSCs (WJ-MSCs) has many advantages over autologous MSCs in which a patient’s own cells are harvested from bodily stores. This includes circumventing the pain and healing process of invasive stem cell harvesting from a patient. Furthermore, MSCs of umbilical cord Wharton’s Jelly origin offer the highest level of potency for therapeutic benefit as they exist in a more naive state than adult MSCs, and thus exhibit increased proliferation ability and anti-inflammatory effects<sup>31</sup>. Specifically, WJ-MSC administration is safe and effective for many indications, including in COVID-19<sup>32</sup>, acute graft versus host disease<sup>33</sup>, and type 2 diabetes<sup>34</sup>, with long term safety confirmed over a 3 year follow on. Further, WJ-MSCs illicit no infusion-related toxicity, no development of treatment related adverse events, nor ectopic tissue formation even at high dosages<sup>33,35</sup>. In this study, we confirm the safety of human allogeneic WJ-MSCs delivered at a high dose (administration of  $1 \times 10^8$  cells total), via multiple delivery routes (intravenous (IV), intrathecal (IT) or intra-articular (IA)), and in conjunction with Mannitol in some cases, for the treatment of various indications. Mannitol is a blood-brain barrier permeabilizer that can facilitate intravenously delivered stem cells to exert therapeutic benefits on the central nervous system, significantly improving recovery in pre-clinical stroke and traumatic brain injury<sup>36-41</sup>. Therefore, Mannitol provides a useful adjunct for subjects unable to tolerate intrathecal administration.

## PATIENTS AND METHODS

### STUDY SUBJECTS AND PROCEDURES

This retrospective study of 22 subjects that were treated with allogeneic human WJ-MSCs at BHI clinical sites between 2017 to 2021. All patients that were treated with human WJ-MSCs for any indication were included in the study. Participants were to be excluded if they had an active systemic infection or an active malignancy during the treatment phase. 13 subjects were treated at BHI Slovakia (Nemocnica Malacky Hospital, Slovakia) and 9 subjects were treated at BHI Jamaica (Montego Bay St. James and Ochos Rios, Jamaica) who were included in the study. Subjects were followed up to a minimum of 6 months post treatment.

Grouped subject characteristics are detailed in Table 1. Of the 13 subjects treated at BHI Slovakia, 10 were male (77%) with mean age of 42 (range 24 – 78), and 3 were female (23%) with a mean age of 62 (range 59 – 64). Of the 9 subjects treated at BHI Jamaica, 7 were male (77.8%) with mean age of 40 (range 25 – 59), and 2 were female (22.2%) with a mean age of 50 (aged 48, 51). Subject specific characteristics, indications and treatment modalities are listed in Table 2.

The study was approved by the Institutional Review Board of the Institute of Regenerative and Cellular Medicine (IRCM-2022-313, IRCM-2022-330, IRCM-2022-331, approved April 28, 2022) and by the Ministry of Health of the Slovak Republic. The BHI Jamaica clinic and internal research studies are licensed with approval from the Minister of Health of Jamaica. All procedures followed were in accordance with the Ethical Standards of the Responsible Committee on Human Experimentation (institutional and national) and with the Helsinki Declaration of 1975, as revised in 2013<sup>42</sup>. All subjects provided written informed consent for inclusion in the study.

### CELL ISOLATION AND EXPANSION

WJ-MSCs were isolated from the Wharton's Jelly fraction of umbilical cord tissue donated with consent of the mother following live birth. The umbilical tissue was cleaned and placed in cultivation medium (79% DMEM (low glucose), 20% FBS, 1% Penicillin-Streptomycin) and incubated at 37°C, 5% CO<sub>2</sub>, 90% humidity for 21 days. Cultivation medium was changed on day 7, 10 and 15. After 21 days, the cell culture was quality checked for blood cell cultivation and sterility. The WJ-MSCs were

**Table 1.** Grouped indication characteristics and AEs.

Indication	N	Sex	Mean Age (range)	#Doses	Delivery Route	Adverse Reaction
<b>BHI Slovakia</b>						
Osteoarthritis (total)	6	4M, 2F	47 (24 – 78)	1	IA	No
Knee	4	M	39 (34 - 43)	1	IA	No
Hip	1	F	43	1	IA	No
Knee & Hip	1	F	59	1	IA	No
Stroke	2	1M, 1F	62.5 (61, 64)	2	IV + IT	Headache (F)
Stroke with MI	1	M	58	2	IV + IT	No
Spinal cord injury	2	M	25 (24, 26)	2	IV + IT	No
Hypoxic brain injury	1	M	3.5	2	IV + IT	No
Vascular insufficiency with diabetic wounds	1	M	78	2	Locally + IV	No
<b>BHI Jamaica</b>						
Osteoarthritis	1	M	54	2	IV	Chills
Muscular weakness	1	M	41	1	IV + Mannitol	Chills, Headache
Neurological spasms	1	F	51	1	IV + Mannitol	No
Age-related decline	2	M	35 (29, 40)	1	IV, IV + Mannitol	No
Multiple Sclerosis	1	F	48	3	IV + IT	No
Stroke	1	M	59	3	IV + IT	No
Hip pain	1	M	30	1	IV	No
Knee pain	1	M	25	2	IV	No

M = Male, F = Female, IA = Intraarticular, IV = Intravenous, IT = Intrathecal, MI = Myocardial Infarction

**Table 2.** Grouped indication characteristics and AEs.

Subject ID	Age	Sex	Diagnosis	Doses	Delivery	Adverse events
SK1	78	M	Vascular insufficiency, diabetic wounds	2 (6 months apart)	Locally + IV	No
SK2	24	M	Spinal cord injury	2 (3 months apart)	IV + IT	No
SK3	64	F	Stroke	1	IV + IT	Headache
SK4	61	M	Stroke	1	IV + IT	No
SK5	58	M	Stroke, Myocardial infarction	2 (6 months apart)	IV + IT	No
SK6	3.5	M	Hypoxic brain injury	2 (3 months apart)	IV + IT	No
SK7	26	M	Spinal cord injury	2 (7 months apart)	IV + IT	No
SK8	63	F	Knee Osteoarthritis	1	IA	No
SK9	59	F	Knee & Hip Osteoarthritis	1	IA	No
SK10	56	M	Knee Osteoarthritis	1	IA	No
SK11	43	M	Hip Osteoarthritis	1	IA	No
SK12	34	M	Knee Osteoarthritis	1	IA	No
SK13	41	M	Knee Osteoarthritis	1	IA	No
JM1	41	M	Muscular Weakness	1	IV + Mannitol	Chills, headache
JM2	54	M	Osteoarthritis	1	IV	Chills
JM3	51	F	Neurological Spasms	1	IV + Mannitol	Chills, slight headache
JM4	40	M	Age-related decline	1	IV + Mannitol	No
JM5	48	F	Multiple Sclerosis	3 (3 days between doses 1 and 2, then 4 months)	IV + IT	No
JM6	59	M	Stroke, left arm and left leg weakness	3 (5 days apart)	IV + IT	No
JM7	25	M	Knee pain	2 (3 months apart)	IV	No
JM8	30	M	Hip pain	1	IV	No
JM9	29	M	Age-related decline	1	IV	No

M = Male, F = Female, IA = Intraarticular, IV = Intravenous, IT = Intrathecal, MI = Myocardial Infarction

dissociated using trypsin and cryopreserved at passage 4 in 10% DMSO. Cells were characterized for positive expression of CD29, CD44, CD73, CD90, CD105, CD166 markers and negative for non-MSC markers CD11b, CD14, CD19, CD31, CD34, CD45, CD79 and HLA-DR<sup>43</sup>. WJ-MSCs were collected and processed under current good manufacturing practice (cGMP) and current good tissue practice (cGTP) specifications and follows a routine protocol across study sites. WJ-MSCS underwent initial pathogen testing for a range of infectious agents prior to cryopreservation or administration. Cell viability, identity and safety was confirmed following low-passage expansion.

#### CELL INFUSION

Cells were thawed in a water bath, centrifuged, and resuspended for infusion. At the Jamaica site, cells were resuspended in 50 mL of normal saline solution. In the Slovakian site, the patient’s own platelet rich plasma (PRP) was combined with saline to make a

20% PRP solution. For intravenous delivery, the cell suspension was filtered through a 170 to 260-micron filter to remove clots and infusion was maintained at a rate of 100 milliliters per hour. For IV delivery only,  $1 \times 10^8$  cell dosage was administered. For intrathecal administration, 2.5 mL of WJ-MSC infusion solution was administered consisting of 1.0 mL patient’s own PRP (Slovakia) or saline (Jamaica) and 1.5 mL WJ-MSC suspension. Subjects were placed in the lateral position and a board-certified anesthesiologist sterilized the lumbar area prior to injection of 2% lidocaine for anesthesia. A spinal needle was inserted under sterile technique into the spinal canal and 2.5 mL of cerebral spinal fluid was aspirated, and the volume replaced with the corresponding infusion of WJ-MSCs. For IV and intrathecal delivery,  $7.5 \times 10^7$  cells are administered intravenously, and  $2.5 \times 10^7$  cells were delivered by intrathecal injection (total cell dosage delivered across both delivery routes amounts to  $1 \times 10^8$  cells). Where intrathecal administration was not tolerated by the subject,  $1 \times$

**Table 3.** Hematological safety evaluation of subject JM4 pre and post WJ-MSC treatment.

Marker	Pre-Treatment	Flag	Post-Treatment	Flag (+7 months)	Reference Interval/Units
Hematocrit	52.5	High	51.3	High	37.5 - 51 %
Creatinine	1.58	High	1.3	High	0.76 - 1.27 mg/dL
A/G ratio	2.3	High	2.4	High	1.2 - 2.2
Hemoglobin	18	High	17	In reference	13 - 17.7 g/dL
BUN	27	High	24	In reference	06 to 24 mg/dL
AST	62	High	36	In reference	0 - 40 IU/L
Triglycerides	184	High	51	In reference	0 -149 mg/dL
eGFR	54	Low	68	In reference	>59 mL/min/1.73
Protein	1+	Abnormal	trace	In reference	Negative - trace
Ketones	Trace	Abnormal	Negative	In reference	Negative
Occult Blood	Trace	Abnormal	Negative	In reference	Negative
Specific Gravity	>1.030	Abnormal	>1.030	Abnormal	1.005 - 1.030
Hep B Core Ab	Positive	Abnormal	Positive	Abnormal	Negative
T Pallidum Ab	Reactive	Abnormal	Reactive	Abnormal	Non-reactive
LDL Cholesterol	82	In reference	114	High	0 - 99 mg/dL

A/G = albumin/globulin, BUN = Blood urea nitrogen, AST = aspartate aminotransferase, eGFR = Estimated glomerular filtration rate.

$10^8$  cells was administered by IV infusion following pretreatment with Mannitol. At minute zero 25% mannitol was introduced into 100 cc of saline solution. The Mannitol solution was infused by intravenous injection over a 10- to 15-minute timeframe, followed immediately by IV infusion of WJ-MSCs as described above. For intraarticular injection,  $2.5 \times 10^7$  cells are injected directly into the affected joint in combination with  $7.5 \times 10^7$  cells administered intravenously as previously described (total cell dosage by both delivery routes amounts to  $1 \times 10^8$  cells).

#### STATISTICAL ANALYSIS

No statistical analysis was performed in this study as it is a descriptive reporting of cases. Age was reported as mean with range for grouped subjects.

## RESULTS

#### ADVERSE EVENTS AND SAFETY EVALUATION

In total, 22 subjects treated for a range of conditions were assessed for serious (SAE) or mild adverse events (AE) (Table 2). In the Slovakian cohort of 13 subjects, a single AE was reported as a transient headache in one stroke patient (ID: SK3, female, 64 years old) that received IV and IT administration of WJ-MSCs. This AE was deemed related to the administration procedure, as headache following IT administration has been reported in 2% of sub-

jects elsewhere<sup>22</sup>. The AE was transient and self-resolved without clinical concern.

In the Jamaican cohort of 9 subjects, 2 subjects reported AEs. One subject (ID: JM2, male, 54 years of age) who received a single IV infusion of WJ-MSCs for osteoarthritis experienced chills persisting for 45 minutes post administration. Chills is a relatively common reaction to cell administration (see discussion) and this AE resolved without clinical concern. Another subject (ID: JM3, female, 51 years of age) who received a single IV infusion of WJ-MSCs for neurological spasms experienced transient chills and a slight headache immediately post administration. Both AEs self-resolved without clinical concern.

In total this study reported AEs in 3 subjects from the 32 doses administered in this study, resulting in an AE rate of 9.3%. Notably, most of these AEs were both transient and mild, and resolved without concern.

#### SINGLE SUBJECT CASE STUDY

Complete blood profiling was conducted pre- and post-treatment to assess changes in blood markers that may be associated with the cell treatment. Here, we report interim results composed of a single subject case study (subject ID: JM4, male, 40 years old). This case study evaluated changes in the blood profile of a subject that received one administration of WJ-MSCs by IV infusion in conjunc-

tion with Mannitol, for age-related decline. The subject reported no AEs during or post treatment. Blood markers flagged as outside of their respective reference intervals pre- or post-treatment were noted and compared to the alternative time point. Fourteen blood markers were flagged as high, low, or abnormal to the reference range in the subject prior to WJ-MSC administration (Table 3). When evaluated at 7-months post cell therapy, 8 of these out-of-reference blood markers (hemoglobin, BUN, eGFR, AST, protein, ketones, occult blood and triglycerides) were corrected to within normal reference intervals. Of the remaining 6 out-of-reference markers; 2 were improved towards the normal reference interval (hematocrit, creatinine), 2 were infectious disease antibody tests (HepB, T Pallidum), and 1 marker was elevated slightly (A/G ratio). Only 1 blood marker associated with diet, LDL cholesterol, was elevated to outside of the normal reference interval in the post-treatment assessment vs. pre-treatment.

Therefore, blood profiling of 75 markers for health and disease in these subject reports that WJ-MSC treatment poses no hematological safety concern, with only one dietary related marker showing marked change during the follow up period which is likely unrelated to the cell administration.

## DISCUSSION

### *EVALUATION OF REPORTED ADVERSE EVENTS*

Two incidents of headaches following cell administration were reported by 2 subjects in this study. These AEs occurred following administration via the intrathecal or intravenous in combination with Mannitol routes. Headaches are a common side effect of intrathecal catheter insertion with “headache attributed to intrathecal injection” included in the International Classification of Headache disorders since its second edition<sup>44</sup>. Their incidence is well documented in literature concerning chemotherapy, pain management and lumbar punctures. It is widely accepted that risk for headache appears to be related to the size of the needle, age and sex of the patient, number of dural punctures (attempts), previous spine surgery, and direction of the bevel<sup>45</sup>. It is hypothesized that a headache occurs due to leak of CSF from the subarachnoid space, causing a drop in intracranial pressure<sup>46,47</sup>. Research estimates that up to a third of chemotherapy patients and lumbar

puncture patients will experience these headaches post-catheter insertion and although bothersome, this effect is typically short-lived and easily managed by analgesia and fluid intake<sup>48,49</sup>. Previous literature does not indicate any significant correlation between dosage of cells and incidence of headache. Headache has been reported as an AE post intrathecal administration of a range of lesser doses that are utilized in this study. Doses of  $4.2 \times 10^6$  cells,  $3 \times 10^7$  cells,  $5.3 \times 10^6$  cells and  $1 \times 10^6$  cells per kg have each been associated with incidence of headache post-administration of stem cells via intrathecal catheter<sup>21,22,50-52</sup>. Therefore, it can be deemed that the occurrence of headaches is not associated with the increased dosage of  $1 \times 10^8$  cells delivered in the current study. Additionally, headache is a commonly accepted side effect of intravenous Mannitol treatment<sup>53</sup>. Considering the prevalence and acceptance of this reaction across other intrathecal procedures, dosages and Mannitol use, the incidence of headache in this study is non-remarkable. Further, the occurrence of headache may be a result of the intrathecal administration route or Mannitol use rather than the WJ-MSC therapy.

Two incidents of subjects experiencing chills post treatment were observed in this study. Current cell practice uses dimethyl sulfoxide (DMSO) as a protective agent to maintain cell viability in the cryopreservation process. However, studies have demonstrated a link between DMSO toxicity and incidence of chills in patients who receive cryopreserved cells as opposed to cells that have been freshly harvested prior to transplantation or transfusion<sup>54-56</sup>. In a study directly comparing the incidence of AE’s in patients receiving cryopreserved cells or freshly harvested cells, chills were reported in 31.1% of patients transfused with cryopreserved marrow, compared to 1.4% of those who received fresh allogeneic marrow<sup>57</sup>. In the current study, chills were only experienced by a small percentage of subjects in the Jamaica cohort who received cells that were not washed prior to infusion. Without placebo control it is difficult to determine which element caused the reaction, such as the cells or the procedure. However, previously reported evidence of DMSO toxicity and chills associated with cryopreservation suggests that this could be a defining factor. At present, further investigation is being conducted into the cause of reported chills.

### CASE STUDY EVALUATION

No safety concerns were highlighted upon review of the single case study described above. Of the 14 abnormal blood markers identified, eight were improved and corrected inside reference intervals when assessed 7 months post treatment. The most dramatic change that occurred was that of the triglycerides. Although some markers such as hemoglobin, hematocrit and protein were only fractionally elevated at pre-treatment, levels of triglycerides were significantly raised. This measure was comfortably within normal range 7 months post-treatment. Derived from glycerol and three fatty acids, triglycerides are a type of lipid found in the blood, with elevated levels of this component contributing to cardiovascular risk<sup>58</sup>. As hypertriglyceridemia is typically associated with poor diet, lifestyle factors or poorly controlled diabetes, it is difficult to evaluate or ascertain the impact of the MSC treatment versus potential lifestyle changes employed by the subject post-administration. However, research has indicated that MSC therapy has the potential to reduce lipid imbalance in the blood with promising results emerging from clinical trials. Cell therapy has been found to be beneficial in altering a dyslipidemic profile, with documentation of the use of bone-marrow MSCs and adipose-derived MSCs for this purpose<sup>59-61</sup>.

With regard to the single case study, there were some abnormalities specific to liver enzymes pre-treatment. Aspartate aminotransferase (AST) was elevated outside normal parameters during the pre-treatment blood analysis, indicating a mild increase in the amount of liver enzymes being excreted into the bloodstream. Liver damage is typically investigated and characterized through comparison of the ratio of liver enzymes, naming AST and alanine aminotransferase (ALT). In this patient, ALT was measured within normal limits, suggesting an element of liver damage or mild cirrhosis. It is important to note that this patient additionally tested positive for Hepatitis B core antibodies, an infection which can cause a degree of liver damage. Irrespective of the determinant for the elevated AST marker, post-treatment the patient experienced corrected regulation of this enzyme level. Although some aspects of the MSCs ability to repair liver cirrhosis remain unclear, clinical trials utilized MSCs have demonstrated 25-37% reduction in fibrosis and improvement in serum total bilirubin and model for end-stage liver disease scores<sup>62,63</sup>. Further trials are

warranted to better understand the correlation between WJ-MSC administration and improvement of these measured hematological markers.

### CONCLUSIONS

This study reports the safety (up to 6 months post treatment) of hWJ-MSCs when delivered via intravenous (including in combination with mannitol), intrathecal or intraarticular administration for a range of indications. We highlight the minimal occurrence of mild adverse reactions, most commonly chills and headache, following high dosage of  $1 \times 10^8$  hWJ-MSCs. Notably, these adverse events were transient and resolved without concern. Hematological profiling of a single subject pre- and post-treatment reports no blood abnormalities and, in the contrary, correction of several biomarkers to within the normal range. We aim to further characterize the long-term safety and efficacy of hWJ-MSC in future studies.

### ACKNOWLEDGEMENTS:

The authors are grateful to all the individuals participating in this study. The study would not have been possible without the cooperation of the patients and their families, the donors and the assistance of the doctors, nurses, and physical therapists at BHI locations.

### ETHICS APPROVAL:

The study was approved by the Institutional Review Board of the Institute of Regenerative and Cellular Medicine (IRCM-2022-313, IRCM-2022-330, IRCM-2022-331, approved April 28, 2022). All procedures followed were in accordance with the ethical standards of the responsible committee on human experimentation (institutional and national) and with the Helsinki Declaration of 1975, as revised in 2013<sup>42</sup>.

### INFORMED CONSENT:

All subjects provided written informed consent for inclusion in the study.

### AVAILABILITY OF DATA AND MATERIAL:

The datasets generated and analyzed during the current study are available from the corresponding author on reasonable request.

### CONFLICT OF INTEREST:

BM, DS, RM and KN are employed by Blue Horizon Therapeutic Sciences. EOG, GSM CT and RG are paid contractors of Blue Horizon Therapeutic Sciences.

### FUNDING:

This retrospective study was funded by BHI Therapeutic Sciences including patient fees for treatment.

**AUTHOR CONTRIBUTIONS:**

BM conceived, designed the study, and treated patients. DS, KN and RM set up the study, obtained the ethical approval, managed the patients, and collected data. DCW prepared the cell product. GS, CT and RG administered the cell therapies and monitored patients. EOG analyzed the data, interpreted results, and wrote the manuscript. RR processed patients. All authors read and approved the final version of the manuscript.

**ORCID:**

Brian Mehling – 0000-0003-4602-8263  
 Dongcheng Wu – 0000-0001-8372-2217  
 Ellen O’Gorman – 0000-0003-2535-1096  
 Doreen Santora – 0000-0001-5285-0371  
 Katarina Novakova – 0000-0001-5841-9988  
 Germain Spencer – 0000-0002-5760-6746  
 Charmaine Thomas – 0000-0001-8667-6789  
 Rohan Grant – 0000-003-4323-0861  
 Ryan Rutherford – 0000-0002-4485-6922  
 Renata Mihályová – 0000-0003-1708-9550

**REFERENCES**

- Erices A, Conget P, Minguell JJ. Mesenchymal progenitor cells in human umbilical cord blood. *Br J Haematol* 2000; 109: 235-242. doi:10.1046/j.1365-2141.2000.01986.x
- Lee RH, Kim B, Choi I, Kim H, Choi HS, Suh, K, Bae YC, Jung JS. Characterization and Expression Analysis of Mesenchymal Stem Cells from Human Bone Marrow and Adipose Tissue. *Cell Physiol Biochem* 2004; 14: 311-324. doi:10.1159/000080341
- Mareschi K, Ferrero I, Rustichelli D, Aschero S, Gammaitoni L, Aglietta M, Madon E, Fagioli F. Expansion of mesenchymal stem cells isolated from pediatric and adult donor bone marrow. *J Cell Biochem* 2006; 97: 744-754. doi:10.1002/jcb.20681
- Wang HS, Hung SC, Peng ST, Huang CC, Wei HM, Guo YJ, Fu YS, Lai MC, Chen CC. Mesenchymal stem cells in the Wharton’s jelly of the human umbilical cord. *Stem Cells Dayt Ohio* 2004; 22: 1330-1337. doi:10.1634/stemcells.2004-0013
- Chapel A, Bertho JM, Bensidhoum M, Fouillard L, Young RG, Frick J, Demarquay C, Mathieu E, Trompier F, Dudoignon N, Germain C, Mazurier C, Aigueperse J, Borneman J, Gorin NC, Gourmelon P, Thierry D. Mesenchymal stem cells home to injured tissues when co-infused with hematopoietic cells to treat a radiation-induced multi-organ failure syndrome. *J Gene Med* 2003; 5: 1028-1038. doi:10.1002/jgm.452
- Rustad KC, Gurtner GC. Mesenchymal Stem Cells Home to Sites of Injury and Inflammation. *Adv Wound Care* 2012; 4: 147-152. doi:10.1089/wound.2011.0314
- Bartholomew A, Sturgeon C, Siatskas M, Ferrer K, McIntosh K, Patil S, Hardy W, Devine S, Ucker D, Deans R, Moseley A, Hoffman R. Mesenchymal stem cells suppress lymphocyte proliferation in vitro and prolong skin graft survival in vivo. *Exp Hematol* 2002; 30: 42-48. doi:10.1016/s0301-472x(01)00769-x
- Tyndall A, Walker UA, Cope A, Dazzi F, De Bari C, Fibbe W, Guiducci S, Jones S, Jorgenson C, Le Blanc K, Luyten F, McGonagle D, Martin I, Bocelli-Tyndall C, Pennesi G, Pistoia V, Pitzalis C, Ucceli A, Wulffraat N, Feldman M. Immunomodulatory properties of mesenchymal stem cells: a review based on an interdisciplinary meeting held at the Kennedy Institute of Rheumatology Division, London, UK, 31 October 2005. *Arthritis Res Ther* 2007; 9: 301. doi:10.1186/ar2103
- Mukai T, Tojo A, Nagamura-Inoue T. Umbilical Cord-Derived Mesenchymal Stromal Cells Contribute to Neuroprotection in Neonatal Cortical Neurons Damaged by Oxygen-Glucose Deprivation. *Front Neurol* 2018; 9: 466. doi:10.3389/fneur.2018.00466
- Kinnaird T, Stabile E, Burnett M, Lee C, Barr S, Fuchs S, Epstein S. Marrow-Derived Stromal Cells Express Genes Encoding a Broad Spectrum of Arteriogenic Cytokines and Promote In Vitro and In Vivo Arteriogenesis Through Paracrine Mechanisms. *Circ Res* 2004; 94: 678-685. doi:10.1161/01.RES.0000118601.37875.AC
- Sémont A, François S, Mouiseddine M, Francois A, Sache A, Frick J, Thierry D, Chapel A, Fisher JP. Mesenchymal Stem Cells Increase Self-Renewal of Small Intestinal Epithelium and Accelerate Structural Recovery after Radiation Injury. In: Fisher JP, ed. *Tissue Engineering. Advances in Experimental Medicine and Biology*. Springer US; 2007: 19-30. doi:10.1007/978-0-387-34133-0\_2
- Mosca JD, Hendricks JK, Buyaner D, Davis-Sproul J, Chuang LC, Majumdar MK, Chopra R, Barry F, Marphy M, Thiede MA, Junker U, Rigg RJ, Forestell SP, Bohnlein E, Storb R, Sandmaier BM. Mesenchymal stem cells as vehicles for gene delivery. *Clin Orthop* 2000; (379 Suppl): S71-90. doi:10.1097/00003086-200010001-00011
- Gao J, Dennis JE, Muzic RF, Lundberg M, Caplan AI. The dynamic in vivo distribution of bone marrow-derived mesenchymal stem cells after infusion. *Cells Tissues Organs* 2001; 169: 12-20. doi:10.1159/000047856
- Allers C, Sierralta WD, Neubauer S, Rivera F, Minguell JJ, Conget PA. Dynamic of distribution of human bone marrow-derived mesenchymal stem cells after transplantation into adult unconditioned mice. *Transplantation* 2004; 78: 503-508. doi:10.1097/01.tp.0000128334.93343.b3
- Liechty KW, MacKenzie TC, Shaaban AF, Radu A, Moseley AB, Deans R, Marshak DR, Flake AW. Human mesenchymal stem cells engraft and demonstrate site-specific differentiation after in utero transplantation in sheep. *Nat Med* 2000; 6: 1282-1286. doi:10.1038/81395
- Devine SM, Cobbs C, Jennings M, Bartholomew A, Hoffman R. Mesenchymal stem cells distribute to a wide range of tissues following systemic infusion into nonhuman primates. *Blood* 2003; 101: 2999-3001. doi:10.1182/blood-2002-06-1830
- Devine SM, Bartholomew AM, Mahmud N, Nelson M, Patil S, Hardy W, Sturgeon C, Hewett T, Chung T, Stock W, Sher D, Weissman S, Ferrer K, Mosca J, Deans R, Moseley A, Hoffman R. Mesenchymal stem cells are capable of homing to the bone marrow of non-human primates following systemic infusion. *Exp Hematol* 2001; 29: 244-255. doi:10.1016/s0301-472x(00)00635-4



18. Lalu MM, McIntyre L, Pugliese C, Fergusson D, Winston BW, Marshall JC, Granton J, Stewart DJ. Safety of Cell Therapy with Mesenchymal Stromal Cells (Safe-Cell): A Systematic Review and Meta-Analysis of Clinical Trials. *PLoS One* 2012; 7: e47559. doi:10.1371/journal.pone.0047559
19. Bang OY, Lee JS, Lee PH, Lee G. Autologous mesenchymal stem cell transplantation in stroke patients. *Ann Neurol* 2005; 57: 874-882. doi:10.1002/ana.20501
20. Suárez-Monteagudo C, Hernández-Ramírez P, Álvarez-González L, Garvia-Maeso I, de la Cuétara-Bernal K, Castillo-Díaz L, Bringas-Vega M, Martínez-Aching G, Morales-Chacón LM, Báez-Martín M, Sánchez-Catasús C, Carballo-Barreda M, Rodríguez-Rojas R, Gómez-Fernández L, Alberti-Amador E, Macías-Abraham C, Balea E, Rosales LC, del Valle Pérez L, Ferrer BB, Gonzalez RM, Bergado JA. Autologous bone marrow stem cell neurotransplantation in stroke patients. An open study. *Restor Neurol Neurosci* 2009; 27: 151-161. doi:10.3233/RNN-2009-0483
21. Barczewska M, Grudniak M, Maksymowicz S, Siwek T, Oldak T, Jezierska-Woźniak K, Gładysz D, Maksymowicz W. Safety of intrathecal injection of Wharton's jelly-derived mesenchymal stem cells in amyotrophic lateral sclerosis therapy. *Neural Regen Res* 2019; 14: 313-318. doi:10.4103/1673-5374.243723
22. Pan K, Deng L, Chen P, Peng Q, Pan J, Wu Y, Wang Y. Safety and Feasibility of Repeated Intrathecal Allogeneic Bone Marrow-Derived Mesenchymal Stromal Cells in Patients with Neurological Diseases. *Stem Cells Int* 2019. doi:10.1155/2019/8421281
23. Jaillard A, Hommel M, Moisan A, Zeffiro TA, Favre-Wiki IM, Barbieux-Guillot M, Vadot W, Marcel S, Lamalle L, Grand S, Detante O. Autologous Mesenchymal Stem Cells Improve Motor Recovery in Subacute Ischemic Stroke: a Randomized Clinical Trial. *Transl Stroke Res* 2020; 11: 910-923. doi:10.1007/s12975-020-00787-z
24. Albu S, Kumru H, Coll R, Vives J, Valles M, Benito-Penalva J, Rodríguez L, Codinach M, Hernández J, Navarro X, Vidal J. Clinical effects of intrathecal administration of expanded Wharton jelly mesenchymal stromal cells in patients with chronic complete spinal cord injury: a randomized controlled study. *Cytotherapy* 2021; 23: 146-156. doi:10.1016/j.jcyt.2020.08.008
25. Lee JS, Hong JM, Moon GJ, Lee PH, Ahn YH, Bang OY. A Long-Term Follow-Up Study of Intravenous Autologous Mesenchymal Stem Cell Transplantation in Patients With Ischemic Stroke. *Stem Cells* 2010; 28: 1099-1106. doi:10.1002/stem.430
26. Mehling B, Hric M, Salatkova A, Vetrak R, Santora D, Ovariova M, Mihalyova R, Manvelyan M. A Retrospective Study of Stromal Vascular Fraction Cell Therapy for Osteoarthritis. *J Clin Med Res* 2020; 12: 747-751. doi:10.14740/jocmr4354
27. Mehling B, Wu DC, Quartararo L, Roman SJ, Pickoff A, Stolfi A, Szelestey B, Rutherford R, Manvelyan M. Allogeneic umbilical cord blood mononuclear cell therapy for spinal cord injury – a retrospective cohort study. *CellR4* 2018; 6: e2521
28. Jiang PC, Xiong WP, Wang G, Ma C, Yao WQ, Kendell SF, Mehling BM, Yuan XH, Wu DC. A clinical trial report of autologous bone marrow-derived mesenchymal stem cell transplantation in patients with spinal cord injury. *Exp Ther Med* 2013; 6: 140-146. doi:10.3892/etm.2013.1083
29. Mehling BM. Evaluation of Immune response to Intravenously Administered Human Cord Blood Stem Cells in the Treatment of Symptoms Related to Chronic Inflammation. *J Stem Cell Res Ther* 2015; 5. doi:10.4172/2157-7633.1000297
30. Mehling BM, Quartararo L, Manvelyan M, Wang P, Wu DC. Safety Study of Intravenously Administered Human Cord Blood Stem Cells in the Treatment of Symptoms Related to Chronic Inflammation. *Int J Med Health Sci* 2015; 9: 573-576.
31. Jin HJ, Bae YK, Kim M, Kwon SJ, Jeon HB, Choi SJ, Kim SW, Yang YS, Oh W, Chang JW. Comparative analysis of human mesenchymal stem cells from bone marrow, adipose tissue, and umbilical cord blood as sources of cell therapy. *Int J Mol Sci* 2013; 14: 17986-18001. doi:10.3390/ijms140917986
32. Saleh M, Vaezi AA, Aliannejad R, Sohrabpour AA, Kiaei SZF, Shadnough M, Siavashi V, Aghaghazvini L, Abdoli S, Chahardouli B, Seyhoun I, Alijani N, Verdi J. Cell therapy in patients with COVID-19 using Wharton's jelly mesenchymal stem cells: a phase 1 clinical trial. *Stem Cell Res Ther* 2021; 12: 410. doi:10.1186/s13287-021-02483-7
33. Soder RP, Dawn B, Weiss ML, Dunavin N, Weir S, Mitchell J, Li M, Shune L, Singh A, Ganguly S, Morrison M, Abdelhakim H, Godwin A, Abhyankar S, McGuirk J. A Phase I Study to Evaluate Two Doses of Wharton's Jelly-Derived Mesenchymal Stromal Cells for the Treatment of De Novo High-Risk or Steroid-Refractory Acute Graft Versus Host Disease. *Stem Cell Rev Rep* 2020; 16: 979-991. doi:10.1007/s12015-020-10015-8
34. Hu J, Wang Y, Gong H, Yu C, Guo C, Wang F, Yan S, Xu H. Long term effect and safety of Wharton's jelly-derived mesenchymal stem cells on type 2 diabetes. *Exp Ther Med* 2016; 12: 1857-1866. doi:10.3892/etm.2016.3544
35. Wang Y, Han ZB, Ma J, Zuo C, Geng J, Gong W, Sun Y, Li H, Wang B, Zhnag L, He Y, Han ZC. A Toxicity Study of Multiple-Administration Human Umbilical Cord Mesenchymal Stem Cells in Cynomolgus Monkeys. *Stem Cells Develop* 2012; 21: 1401-1408. doi: http://doi.org/10.1089/scd.2011.0441
36. Seyfried DM, Han Y, Yang D, Ding J, Savant-Bhonsale S, Shukairy MS, Chopp M. Mannitol enhances delivery of marrow stromal cells to the brain after experimental intracerebral hemorrhage. *Brain Res* 2008; 1224: 12-19. doi:10.1016/j.brainres.2008.05.080
37. Okuma Y, Wang F, Toyoshima A, Kameda M, Hishikawa T, Tokunaga K, Sugi K, Liu K, Jaruma J, Nishibori M, Yasuhara T, Date I. Mannitol enhances therapeutic effects of intra-arterial transplantation of mesenchymal stem cells into the brain after traumatic brain injury. *Neurosci Lett* 2013; 554: 156-161. doi:10.1016/j.neulet.2013.08.058
38. Gonzales-Portillo GS, Sanberg PR, Franzblau M, Gonzales-Portillo C, Diamandis T, Staples M, Sanberg CD, Borlongan CV. Mannitol-enhanced delivery of stem cells and their growth factors across the blood-brain barrier. *Cell Transplant* 2014; 23: 531-539. doi:10.3727/096368914X678337

39. Borlongan CV, Hadman M, Sanberg CD, Sanberg PR. Central nervous system entry of peripherally injected umbilical cord blood cells is not required for neuroprotection in stroke. *Stroke* 2004; 35: 2385-2389. doi:10.1161/01.STR.0000141680.49960.d7
40. Yasuhara T, Hara K, Maki M, Xu L, Yu G, Ali MM, Masuda T, Yu SJ, Bae EK, Hayashi T, Matsukawa N, Kaneko Y, Kuzmin-Nichils N, Ellovitch S, Cruz EL, Klasko SK, SANberg CD, Sanberg PR, Borlongan CV. Mannitol facilitates neurotrophic factor up-regulation and behavioural recovery in neonatal hypoxic-ischaemic rats with human umbilical cord blood grafts. *J Cell Mol Med* 2010; 14: 914-921. doi:10.1111/j.1582-4934.2008.00671.x
41. Tajiri N, Lee JY, Acosta S, Sanberg PR, Borlongan CV. Breaking the Blood-Brain Barrier with Mannitol to Aid Stem Cell Therapeutics in the Chronic Stroke Brain. *Cell Transplant* 2016; 25: 1453-1460. doi:10.3727/096368916X690971
42. WMA - The World Medical Association-WMA Declaration of Helsinki – Ethical Principles for Medical Research Involving Human Subjects. Accessed May 2, 2022. <https://www.wma.net/policies-post/wma-declaration-of-helsinki-ethical-principles-for-medical-research-involving-human-subjects/>
43. Dominici M, Le Blanc K, Mueller I, Slaper-Cortenbach I, Marini FC, Krause DS, Deans RJ, Keating A, Prockop DJ, Horwitz EM. Minimal criteria for defining multipotent mesenchymal stromal cells. The International Society for Cellular Therapy position statement. *Cytotherapy* 2006; 8: 315-317. doi:10.1080/14653240600855905
44. Headache Classification Subcommittee of the International Headache Society. The International Classification of Headache Disorders: 2nd edition. *Cephalalgia Int J Headache* 2004; 24 Suppl 1: 9-160. doi:10.1111/j.1468-2982.2003.00824.x
45. Staats PS. Complications of Intrathecal Therapy. *Pain Med* 2008; 9: S102-S107. doi:10.1111/j.1526-4637.2008.00445.x
46. Ahmed SV, Jayawarna C, Jude E. Post lumbar puncture headache: diagnosis and management. *Postgrad Med J* 2006; 82: 713-716. doi:10.1136/pgmj.2006.044792
47. Grant R, Condon B, Hart I, Teasdale GM. Changes in intracranial CSF volume after lumbar puncture and their relationship to post-LP headache. *J Neurol Neurosurg Psychiatry* 1991; 54: 440-442. doi:10.1136/jnnp.54.5.440
48. De la Riva P, Andres-Marín N, Gonzalo-Yubero N, Tainta-Cuezva M, Caminos N, Urtasun-Ocariz MA, Martí-Massó JF. Headache and other complications following intrathecal chemotherapy administration. *Cephalalgia* 2017; 37: 1109-1110. doi:10.1177/0333102416668658
49. Gaiser R. Postdural puncture headache. *Curr Opin Anesthesiol* 2006; 19: 249-253. doi:10.1097/01.aco.0000192809.71408.ba
50. Hur JW, Cho TH, Park DH, Lee JB, Park JY, Chung YG. Intrathecal transplantation of autologous adipose-derived mesenchymal stem cells for treating spinal cord injury: A human trial. *J Spinal Cord Med* 2016; 39: 655-664. doi:10.1179/2045772315Y.0000000048
51. Oh KW, Noh MY, Kwon MS, Kim HY, Oh SI, Park J, Kim HJ, Ki CS, Kim SH. Repeated Intrathecal Mesenchymal Stem Cells for Amyotrophic Lateral Sclerosis. *Ann Neurol* 2018; 84: 361-373. doi:10.1002/ana.25302
52. Sharma AK, Sane HM, Paranjape AA, Gokulchandran N, Nagrajan A, D'sa M, Badhe PB. The effect of autologous bone marrow mononuclear cell transplantation on the survival duration in Amyotrophic Lateral Sclerosis - a retrospective controlled study. *Am J Stem Cells* 2015; 4: 50-65.
53. Side Effects of Mannitol IV (Mannitol Injection), Warnings, Uses. RxList. Accessed January 8, 2022. <https://www.rxlist.com/mannitol-iv-side-effects-drug-center.htm>
54. Davis J, Rowley S, Braine H, Piantadosi S, Santos G. Clinical toxicity of cryopreserved bone marrow graft infusion. *Blood* 1990; 75: 781-786. doi:10.1182/blood.V75.3.781.781
55. Awan M, Buriak I, Fleck R, Fuller B, Goltsev A, Kerby J, Lowdell M, Mericka P, Petrenko A, Pentrenko Y, Rogulska O, Stolzing A, Stacey GN. Dimethyl sulfoxide: a central player since the dawn of cryobiology, is efficacy balanced by toxicity? *Regen Med* 2020; 15: 1463-1491. doi:10.2217/rme-2019-0145
56. Truong TH, Moorjani R, Dewey D, Guilcher GMT, Prokopishyn NL, Lewis VA. Adverse reactions during stem cell infusion in children treated with autologous and allogeneic stem cell transplantation. *Bone Marrow Transplant* 2016; 51: 680-686. doi:10.1038/bmt.2015.331
57. Stroncek D, Fautsch S, Lasky L, Hurd D, Ramsay N, McCullough J. Adverse reactions in patients transfused with cryopreserved marrow. *Transfusion (Paris)* 1991; 31: 521-526. doi:10.1046/j.1537-2995.1991.31691306250.x
58. Rosas S, Szapary P, Rader DJ. Management of selected lipid abnormalities: hypertriglyceridemia, isolated low HDL-cholesterol, lipoprotein(a), and lipid abnormalities in renal diseases and following solid organ transplantation. *Cardiol Clin* 2003; 21: 377-392. doi:10.1016/S0733-8651(03)00075-4
59. Hussein EN, Hamed GM, Seif AA, Ahmed MA, Abu Zahra FAE. Effects of Mesenchymal Stem Cells Therapy on Cardiovascular Risk Factors in Experimental Diabetic Kidney Disease. *Can J Kidney Health Dis* 2020; 7: 2054358120957429. doi:10.1177/2054358120957429
60. Li F, Guo X, Chen SY. Function and Therapeutic Potential of Mesenchymal Stem Cells in Atherosclerosis. *Front Cardiovasc Med* 2017; 4: 32. doi:10.3389/fcvm.2017.00032
61. Kirwin T, Gomes A, Amin R, Sufi A, Goswami S, Wang B. Mechanisms underlying the therapeutic potential of mesenchymal stem cells in atherosclerosis. *Regen Med*. Published online June 30, 2021. doi:10.2217/rme-2021-0024
62. Lin BL, Chen JF, Qiu WH, Wang KW, Xie DY, Chen XY, Liu QL, Peng L, Li JG, Mei YY, Weng WZ, Peng YW, Cao HJ, Xie JQ, Xie SB, Xiang AP, Gao ZL. Allogeneic bone marrow-derived mesenchymal stromal cells for hepatitis B virus-related acute-on-chronic liver failure: A randomized controlled trial. *Hepatology* 2017; 66: 209-219. doi:10.1002/hep.29189
63. Suk KT, Yoon JH, Kim MY, Kim CW, Kim Jk, Park H, Hwang SG, Kim DJ, Lee BS, Lee SH, Kim HS, Jang JY, Lee CH, Kim BS, Jang YO, Cho MY, Jung ES, Kim YM, Bae SH, Baik SK. Transplantation with autologous bone marrow-derived mesenchymal stem cells for alcoholic cirrhosis: Phase 2 trial. *Hepatology* 2016; 64: 2185-2197. doi:10.1002/hep.28693