Evolving technologies in regenerative medicine

P. Hollands¹, T. Tkebuchava²

¹Freelance Consultant Clinic Scientist, Cambridge, UK

Corresponding Author: Peter Hollands, PhD (Cantab); e-mail: peterh63@hotmail.com

Keywords: Regenerative Medicine, Stem Cell Technology, Evolving Technology, RegMedGeorgia.

Abstract:

This short review explores the constantly evolving technology, both cellular and equipment or technology based, which is currently being experienced in the field of regenerative medicine. There is considerable ongoing research into stem cell technologies but there is currently a translational block in bringing some of these technologies to the clinic. We propose new and evolving technologies which may resolve this cell-based block and quickly bring regenerative medicine technologies safely and effectively to patients in need.

Introduction

Regenerative medicine (excluding routine bone marrow transplantation) has developed over the past 20-35 years based on concepts around stem cells¹. This has been on the premise that stem cells are self-replicating and can therefore potentially repair and restore damaged or diseased tissue, for example during wound healing². Many different stem cells, potentially useful in regenerative medicine, have been identified in almost every tissue in the body. These include Haemopoietic Stem Cells (HSC)³⁻⁶, Mesenchymal Stem Cells (MSC) from various sources^{7,8}, Embryonic Stem Cells^{9,10} and induced Pluripotent Stem Cells (iPSC)^{11,12}. Despite this extensive amount of literature on stem cell technology (some of it in the most highly regarded journals) there are currently no stem cell therapies (apart from HSC transplantation for haematological malignancy) in current routine clinical practice. There seems to be a block at the translational stage¹³ perhaps related to stringent regulatory requirements¹⁴. In this short review, we will present some emerging and evolving technologies which may help to bring much needed help to patients needing regenerative medicine procedures.

EXTRACORPOREAL BLOOD OXYGENATION AND OZONATION (EBOO)

EBOO therapy is an emerging technology which has already been used by some clinicians and appears to be a safe and effective option for some patients¹⁵. There has been considerable progress with the use of Ozone therapy in the treatment of Covid19 both in the acute stage and in 'long Covid'16-19. The use of ozone therapy has recently been described in the safe and effective treatment of musculoskeletal diseases such as rheumatoid arthritis^{20,21}, osteoarthritis²², herniated disc²³ and temporomandibular joint disorder²⁴. Other workers have described considerable the benefit of EBOO in the treatment of peripheral arterial disease, coronary disease, severe dyslipidaemia, coronary disease, Madelung disease and deafness of vascular origin²⁵. The current data on the use of EBOO in peripheral artery disease (PAD) seem especially encouraging with no side effects or complications being reported²⁶. These authors conclude that EBOO is an effective treatment for skin lesions in PAD along with additional benefits to general health and wellbeing. There was, however, no observed change in arterial circulation in these patients suggesting that the



² Medical Director and CEO, RegMedGeorgia, Tbilisi, Georgia

mechanism of action of EBOO is complex and still to be fully understood. There has also been considerable progress in the use of EBOO in the potentially deadly disease of necrotizing fasciitis²⁷. This treatment approach using EBOO is now used in many hospitals as a routine treatment of necrotizing fasciitis. EBOO clearly has a wide range of very beneficial and safe therapeutic use, this makes EBBO an important contributor to the overall holistic patient treatment in parallel to regenerative medicine procedures²⁸. Our own experience with EBOO at RegMedGoergia is equally promising (with publications in preparation) and there is no doubt that EBOO will make a very important contribution to the overall practice of regenerative medicine in the future.

PHOTOMODULATION

Photomodulation (sometimes known as photobiomodulation) has been shown the be a useful process in the optimisation of regenerative medicine procedures involving MSC²⁹. Low energy laser light has also been shown to be effective in the activation of adipose derived stem cells³⁰. It is clear that photomodulation is a useful tool for the spatiotemporal modulation of a wide range of biological systems where photoresponsive components are present³¹. Photomodulation can be used in two main ways³²:

- 1. Photomodulation of a therapeutic vehicle or medication or molecules, e.g., cell surface antigens. These may be responsive to UV, visible and IR light (modulated or un-modulated).
- Photomodulation as a light-generated mediator signal, e.g., production of heat, hypoxia, reactive oxygen species (ROS), and other gas molecules.

Low-level light therapy (LLLT) is an area of intense research at present with a particular focus on using LLLT as a treatment for acne vulgaris, facial dyschromia and rhytids, androgenic alopecia, and wound healing³³. Nevertheless, further clinical trials are needed to fully understand the safety and efficacy of this technology. There has been equally promising work in the use of photobiomodulation to treat oral mucositis in patients undergoing treatment for cancer^{34,35}. There are further recent reports of the benefits of photobiomodulation in wound care but with the cautionary note that clinical protocols need to be optimized by the use of rigorous clinical research studies³⁶. Our own work

in the field of photobiomodulation has shown that a modulated low-power red laser can activate pluripotent human Very Small Embryonic Like (hVSEL) stem cells in autologous Platelet Rich Plasma (PRP) as discussed below. We proposed a mechanism of action being interactions of modulated laser light with the hVSEL stem cells at the quantum level³⁷. We are just at the beginning of understanding and applying photomodulation in regenerative medicine but the future is very bright.

BIOACTIVE AGENTS IN REGENERATIVE MEDICINE

There are many bioactive agents which may be complementary to regenerative medicine. PRP has for example been important and clinically useful not only in aesthetic (cosmetic) procedures but also in regenerative medicine³⁸. Some workers claim that PRP is the 'elixir of youth' when applied for skin rejuvenation and restoration of hair growth³⁹. This may be wishful thinking on the part of the authors, but there is no doubt that PRP is a potent tool which has greatly enhanced regenerative medicine. PRP contains high numbers of platelets which produce various cytokines and activation factors, a high concentration of growth factors and cytokines in the plasma itself, and high numbers of pluripotent hVSEL stem cells which will be discussed further below⁴⁰.

Another bioactive agent which shows great promise to be complementary to regenerative medicine is Hyaluronic Acid (HA)⁴¹. HA has been used in various clinical studies, especially in relation to orthopedic disease such as tendinopathies and osteochondral lesions. The combination of HA with stem cell technology may enhance the overall efficacy of such treatments. HA seems to be very effective in orthopaedic disease when given by the intra-articular route⁴² and the associated stem cells (e.g., MSC) could be delivered by the same route⁴³.

Various types of collagens, (sometimes known as the collagen superfamily), have important structural and physiological roles in regenerative medicine⁴⁴. Collagen can be used to create 3D structures which are exceptionally useful in such procedures as tissue grafts. The collagen enables viable stem cell populations to be maintained in grafted tissue, especially in diseases such as corneal scarring⁴⁵. All of these components, and no doubt many more in the future, will have a significant positive benefit on the overall efficacy of regenerative medicine.

Exosomes in Regenerative Medicine

Mammalian exosomes are small acellular vesicles (30-140 nm in diameter) which are released by most mammalian cells. Exosomes are thought to be involved in cell-to-cell communication and cell-tocell regulation⁴⁶. The current obstacles to bringing exosomes into routine clinical practice include a lack of understanding between exosome structure and function and early-stage development of scalable production⁴⁷. In order to be routinely useful in regenerative medicine then exosomes need to be produced under Good Manufacturing Practice (GMP) conditions in order to ensure safe, consistent, and effective batches of exosomes for clinical use⁴⁸. Despite these hindrances to the overall development of exosome technology, there are reports of clinical-grade exosome production which have been used to treat pancreatic cancer⁴⁹, in general cancer therapeutic procedures⁵⁰, in wound healing⁵¹ and for drug delivery systems⁵². Our own work using stem cells and exosomes to treat cerebrovascular accidents (CVA) is extremely promising (publication in preparation). It is clear that exosomes have an important role in future therapeutic procedures, and it is inevitable that they will become an important component of regenerative medicine in the future

PLANT-DERIVED EXOSOMES (PDE)

Plant-derived exosomes (PDE) are a type of extracellular vesicle and are from 50-100 nm in diameter⁵³. These PDE have been shown to contain lipids, micro RNAs, proteins and many other metabolites and it has been observed that some PDE have anti-inflammatory properties⁵⁴. In comparison to mammalian derived exosomes PDE appear to have a higher bioavailability, less immunogenicity and increased innocuity⁵⁵.

The key advantages of PDE are:

- They are biomimetic to mammalian derived exosomes making them an easy, reliable reproducible and safe alternative to mammalian derived exosomes.
- They are lipophilic and will therefore pass through the skin to the lower dermis and the dermal capillary bed very easily. This means that they can be used as a cosmeceutical thus avoiding complex and expensive licensing procedures.

Plant derived exosomes are certainly an evolving technology in regenerative medicine especially in the treatment of inflammation, chronic pain, burns, anti-aging and the regrowth of hair where they have already shown great potential.

EXTRACORPOREAL SHOCK WAVE THERAPY (ESWT)

ESWT is a non-invasive therapy which has developed from Extracorporeal Shock Wave Lithotripsy (ESWL)⁵⁶. ESWT is proving increasingly important and effective especially in the field of orthopaedics⁵⁷ and sports medicine⁵⁸. The physics of ESWT is relatively complex and beyond the scope if this review. nevertheless there are some excellent reviews on the subject⁵⁹. ESWT relies on a shock or pressure wave transforming into a biological response, this is the so called 'mechnano-transduction' 60-62. The mechanism of action of ESWT is currently being understood at the molecular level which involves cell surface antigen activation. These activated cell surface antigens are in turn stimulated by the appropriate cytokines and growth factors resulting in the beneficial clinical effects seen^{58,59}. A much more detailed understanding will be needed in the future, perhaps even at the quantum level, but at present this work has yet to be completed. It is possible that ESWT activates the Wnt5a/Ca²⁺ signalling in bone marrow mesenchymal stem cells (MSC) which could in turn promote repair and regeneration by these stem cells especially in musculoskeletal disease⁶³. Our own experience with ESWT in RegmedGeorgia has so far been extremely encouraging suggesting that ESWT in another technique which has a great deal to offer in the overall application of regenerative medicine.

PLATELET RICH PLASMA (PRP) IN REGENERATIVE MEDICINE

Platelet-rich plasma (PRP) has been shown to be a valuable therapeutic in regenerative medicine related procedures including the treatment of musculoskeletal disease^{64,65} and in the treatment of sports-related injuries^{66,67}. PRP was traditionally considered to be a blood product which had a platelet count higher than that found in peripheral blood, making it a possible treatment for thrombocytopenia⁶⁸.

Platelets are acellular and they contain four types of granules:

- Alpha granules⁶⁹ containing the adhesive proteins fibrinogen, vitronectin, thrombospondin, and von Willebrand Factor (VWF). In addition, alpha granules contain growth factors and cytokines which may mediate wound repair, inflammation, and angiogenesis⁷⁰.
- Dense (or delta) granules containing ADP, ATP, calcium, serotonin, polyphosphate, and pyrophosphate⁷¹.
- Lysosomes containing hexosaminidase, arylsulphatase or arylsulfatase, β-glucuronidase, β-galactosidase, acid phosphatase, and cathepsins⁷².
- T (or tubular) granules containing TLR9, PDI, and VAMP-8 which are thought to be an alpha granule subtype⁷³

The platelets in PRP clearly have a potential role in the regenerative process⁷⁴. The plasma in PRP contains high concentrations of growth factors and cytokines including interleukins, RANTES, PDGF, VEGF, GM-CSF, MIP 1b, and CXCL chemokine (IP-10)⁷⁵. These cytokines and growth factors, in concentrated form in PRP, enable differentiation, proliferation, tissue morphogenesis, and chemotaxis in tissue healing⁷⁶. The mechanism of action of these cytokines and growth factors in PRP is proposed to be by the autocrine and paracrine route⁷⁷. PRP is clearly of present and future importance in regenerative medicine. It will, no doubt, be used alone and in combination with cell therapies in the future.

HUMAN VERY SMALL EMBRYONIC-LIKE (HVSEL) STEM CELLS IN REGENERATIVE MEDICINE

The third component of PRP, which is often ignored or dismissed as being present in PRP, is hVSEL stem cells. Research has clearly shown the presence of CXCR4+, SSEA4+, Oct 3/4+, CD45-, Lin- hVSEL pluripotent stem cells in PRP derived from human peripheral blood⁷⁸. The biological and therapeutic action of PRP does not currently include discussions on the importance of hVSEL stem cells in PRP in the overall efficacy of PRP treatments.

The persistence of hVSEL stem cells throughout life has been reported⁷⁹ suggesting a potential homeostatic mechanism which maintains the hVSEL stem cell pool throughout life. The bone marrow is the likely source of the hVSEL stem cells. This is supported by the observation of mobilization of hVSEL stem cells

into the peripheral blood following acute myocardial infarction⁸⁰. In addition, hVSEL stem cells have been shown to be present in the human bone marrow and also in human leukapheresis products⁸¹. This supports the hypothesis of migration of hVSEL stem cells from the bone marrow to the peripheral blood during physiological homeostasis and during pathological stimuli. Similar studies *in vitro* have suggested that hVSEL stem cells are the 'original embryonic stem cell' highlighting the critical importance of hVSEL stem cells in normal embryonic development and subsequent physiological homeostasis⁸².

All stem cell types are subject to both intrinsic and extrinsic stress during normal physiology and in pathological states. Such stress can have detrimental effects, especially on rapidly dividing stem cells. There is a population of quiescent VSEL stem cells in murine bone marrow which may be resistant to extrinsic heat stress in the same way as the quiescent population of MSC derived from desquamated endometrium of menstrual blood (eMSC)⁸³. This raises the possibility that quiescent hVSEL stem cells residing in the bone marrow may be available for collection and QiLaser activation following high-dose chemotherapy. If this can be proven, then quiescent hVSEL stem cells may offer an alternative therapeutic route to patients who have undergone chemotherapy with the resultant damage to normal somatic cells. Such an approach may have the ability to enhance somatic cell and tissue repair following high dose chemotherapy⁸⁴.

QILASER ACTIVATION OF PRP

Standard autologous PRP treatments usually involve the collection of peripheral blood into citrate dextrose anticoagulant, centrifugation at room temperature, and then simple reinfusion of the room temperature PRP back into the patient. Such preparation of inactivated PRP may result in premature platelet activation which can be modulated by introducing thrombin into the PRP82. Research has clearly shown that QiLaser (previously known as the SONG modulated laser) modulated laser light interacts with hVSEL stem cells in PRP to upregulate the expression of CXCR4, Oct 3/4, and SSEA4⁷⁸. The use of light as a method of activation stem cells has also been confirmed by other workers⁸⁶. The proposed mode of action for OiLaser modulated laser light on hVSEL stem cell activation has been made using concepts taken from quantum physics⁸⁷. The QiLaser can also be applied to the patient as part of the treatment, to the areas where the stem cell repair is needed e.g. to the head in the case of neurodegenerative disease. This is supported by other authors who propose that enhanced homing of stem cells may improve the efficacy of regenerative medicine procedures⁸⁸. The benefits of QiLaser activated hVSEL stem cells are that they are autologous, they are very easily collected from the peripheral blood and PRP prepared, and they can be used to treat a wide range of diseases. This is because hVSEL stem cells are pluripotent. There is currently no other readily available source of pluripotent human stem cells for clinical use.

CONCLUSIONS

Regenerative medicine is developing at a rapid rate. Cellular-based therapies are slowly being introduced and acellular therapies, such as those described above, have entered routine clinical use. RegMedGeorgia will lead the way in the use of innovative regenerative medicine technologies along with colleagues around the World.

FUNDING:

No funding is declared for this article.

ORCID ID:

P. Hollands: 0000-0003-4116-1954 T. Tkebuchava: 0000-0001-8372-2452

AUTHOR CONTRIBUTIONS:

Peter Hollands: Composing and revising manuscript and joint final approval of manuscript for publication. Tengiz Tkebuchava: Composing and revising manuscript and joint final approval of manuscript for publication.

CONFLICT OF INTEREST:

Professor Peter Hollands is a Freelance Consultant Clinical Scientist. Dr Tengiz Tkebuchava is Medical Director and CEO of RegMedGeorgia

REFERENCES

- Dulak J, Szade K, Szade A, Nowak W, Józkowicz A. Adult stem cells: hopes and hypes of regenerative medicine. Acta Biochim Pol 2015; 62: 329-337.
- 2. Dekoninck S, Blanpain C. Stem cell dynamics, migration and plasticity during wound healing. Nat Cell Biol. 2019; 21: 18-24.

- 3. Hollands P. Differentiation and grafting of haemopoietic stem cells from early postimplantation mouse embryos. Development 1987; 99: 69-76.
- 4. Hollands P. Embryonic haemopoietic stem cell grafts in the treatment of murine genetic anaemia. Br J Haematol 1988; 70: 157-163.
- Hollands P. Transplantation of embryonic haemopoietic stem cells without prior recipient X-irradiation. Br J Haematol 1988; 69: 437-440.
- Hollands P. Differentiation of embryonic haemopoietic stem cells from mouse blastocysts grown in vitro. Development 1988; 102: 135-141.
- 7. Hollands P, Aboyeji D, Orcharton M. Dental pulp stem cells in regenerative medicine. Br Dent J 2018; 10; 35-42.
- 8. Han Y, Li X, Zhang Y, Han Y, Chang F, Ding J. Mesenchymal Stem Cells for Regenerative Medicine. Cells 2019; 8: 886-890.
- 9. Doğan A. Embryonic Stem Cells in Development and Regenerative Medicine. Adv Exp Med Biol 2018; 1079: 1-15.
- Dupont G, Yilmaz E, Loukas M, Macchi V, De Caro R, Tubbs RS. Human embryonic stem cells: Distinct molecular personalities and applications in regenerative medicine. Clin Anat 2019; 32: 354-360.
- 11. Ohnuki M, Takahashi K. Present and future challenges of induced pluripotent stem cells. Philos Trans R Soc Lond B Biol Sci 2015; 370: 20140367.
- Robinton DA, Daley GQ. The promise of induced pluripotent stem cells in research and therapy. Nature 2012; 481: 295-305.
- Banerjee C. Stem cells therapies in basic science and translational medicine: current status and treatment monitoring strategies. Curr Pharm Biotechnol 2011; 12: 469-487.
- Zakrzewski W, Dobrzyński M, Szymonowicz M, Rybak Z. Stem cells: past, present, and future. Stem Cell Res Ther 2019; 10: 68-72.
- Re L, Noci JB, Gadelha Serra ME, Mollica P, Bonetti M, Travagli V. Safety, pitfalls, and misunderstandings about the use of ozone therapy as a regenerative medicine tool. A narrative review. J Biol Regul Homeost Agents 2020; 34: 1-13.
- Marini S, Maggiorotti M, Dardes N, Bonetti M, Martinelli M, Re L, Carinci F, Tavera C. Oxygen-ozone therapy as adjuvant in the current emergency in SARS-COV-2 infection: a clinical study. J Biol Regul Homeost Agents 2020; 34: 757-766.
- Valdenassi L, Franzini M, Ricevuti G, Rinaldi L, Galoforo AC, Tirelli U. Potential mechanisms by which the oxygen-ozone (O2-O3) therapy could contribute to the treatment against the coronavirus COVID-19. Eur Rev Med Pharmacol Sci 2020; 24: 4059-4061.
- 18. Ranaldi GT, Villani ER, Franza L. Rationale for ozonetherapy as an adjuvant therapy in COVID-19: a narrative review. Med Gas Res 2020; 10: 134-138.
- 19. Hernández A, Viñals M, Isidoro T, Vilás F. Potential Role of Oxygen-Ozone Therapy in Treatment of COV-ID-19 Pneumonia. Am J Case Rep 2020; 21: e925849.
- Tartari APS, Moreira FF, Pereira MCDS, Carraro E, Cidral-Filho FJ, Salgado AI, Kerppers II. Anti-inflammatory Effect of Ozone Therapy in an Experimental Model of Rheumatoid Arthritis. Inflammation 2020; 43: 985-993.

- Seyam O, Smith NL, Reid I, Gandhi J, Jiang W, Khan SA. Clinical utility of ozone therapy for musculoskeletal disorders. Med Gas Res 2018; 8: 103-110.
- Costa T, Rodrigues-Manica S, Lopes C, Gomes J, Marona J, Falcão S, Branco J. Ozone Therapy in Knee Osteoarthritis: A Systematic Review. Acta Med Port 2018; 31: 576-580. Portuguese.
- Magalhaes FN, Dotta L, Sasse A, Teixera MJ, Fonoff ET. Ozone therapy as a treatment for low back pain secondary to herniated disc: a systematic review and metaanalysis of randomized controlled trials. Pain Physician 2012;15: E115-29.
- Haghighat S, Oshaghi S. Effectiveness of Ozone Injection Therapy in Temporomandibular Disorders. Adv Biomed Res. 2020; 9: 73.
- Di Paolo N, Gaggiotti E, Galli F. Extracorporeal blood oxygenation and ozonation: clinical and biological implications of ozone therapy. Redox Rep 2005; 10: 121-130.
- Di Paolo N, Bocci V, Salvo DP, Palasciano G, Biagioli M, Meini S, Galli F, Ciari I, Maccari F, Cappelletti F, Di Paolo M, Gaggiotti E. Extracorporeal blood oxygenation and ozonation (EBOO): a controlled trial in patients with peripheral artery disease. Int J Artif Organs 2005; 28: 1039-1050.
- Di Paolo N, Bocci V, Cappelletti F, Petrini G, Gaggiotti E. Necrotizing fasciitis successfully treated with extra-corporeal blood oxygenation and ozonization (EBOO). Int J Artif Organs 2002; 25: 1194-1198.
- 28. Di Paolo N, Bocci V, Garosi G, Borrelli E, Bravi A, Bruci A, Aldinucci C, Capotondo L. Extracorporeal blood oxygenation and ozonation (EBOO) in man preliminary report. Int J Artif Organs 2000; 23: 131-141.
- 29. Pinto H, Goñi Oliver P, Sánchez-Vizcaíno Mengual E. The Effect of Photobiomodulation on Human Mesenchymal Cells: A Literature Review. Aesthetic Plast Surg 2021; 45: 1826-1842.
- Min KH, Byun JH, Heo CY, Kim EH, Choi HY, Pak CS. Effect of Low-Level Laser Therapy on Human Adipose-Derived Stem Cells: In Vitro and In Vivo Studies. Aesthetic Plast Surg 2015; 39: 778-82.
- Zhu C, Kou T, Kadi AA, Li J, Zhang Y. Molecular platforms based on biocompatible photoreactions for photomodulation of biological targets. Org Biomol Chem 2021; 19: 9358-9368.
- Zhou Y, Ye H, Chen Y, Zhu R, Yin L. Photoresponsive Drug/Gene Delivery Systems. Biomacromolecules 2018; 19: 1840-1857.
- Glass GE. Photobiomodulation: The Clinical Applications of Low-Level Light Therapy. Aesthet Surg J 2021; 41: 723-738.
- 34. Elad S, Arany P, Bensadoun RJ, Epstein JB, Barasch A, Raber-Durlacher J. Photobiomodulation therapy in the management of oral mucositis: search for the optimal clinical treatment parameters. Support Care Cancer 2018: 26: 3319-3321.
- 35. Zadik Y, Arany PR, Fregnani ER, Bossi P, Antunes HS, Bensadoun RJ, Gueiros LA, Majorana A, Nair RG, Ranna V, Tissing WJE, Vaddi A, Lubart R, Migliorati CA, Lalla RV, Cheng KKF, Elad S; Mucositis Study Group of the Multinational Association of Supportive Care in

- Cancer/International Society of Oral Oncology (MAS-CC/ISOO). Systematic review of photobiomodulation for the management of oral mucositis in cancer patients and clinical practice guidelines. Support Care Cancer 2019: 27: 3969-3983.
- Mosca RC, Ong AA, Albasha O, Bass K, Arany P. Photobiomodulation Therapy for Wound Care: A Potent, Noninvasive, Photoceutical Approach. Adv Skin Wound Care. 2019; 32: 157-167.
- 37. Brindley J, Hollands P, Ovokaitys T. A Theoretical Mechanism for the Action of SONG-Modulated Laser Light on Human Very Small Embryonic-Like (hVSEL) Stem Cells in Platelet Rich Plasma (PRP). CellR4 2021; 9: e3201.
- 38. Samadi P, Sheykhhasan M, Khoshinani HM. The Use of Platelet-Rich Plasma in Aesthetic and Regenerative Medicine: A Comprehensive Review. Aesthetic Plast Surg 2019; 43: 803-814.
- Elghblawi E. Platelet-rich plasma, the ultimate secret for youthful skin elixir and hair growth triggering. J Cosmet Dermatol. 2018; 17: 423-430.
- Collins T, Alexander D, Barkatali B. Platelet-rich plasma: a narrative review. Open Rev 2021; 6: 225-235.
- 41. Pereira H, Sousa DA, Cunha A, Andrade R, Espregueira-Mendes J, Oliveira JM, Reis RL. Hyaluronic Acid. Adv Exp Med Biol 2018; 1059: 137-153.
- 42. Iannitti T, Lodi D, Palmieri B. Intra-articular injections for the treatment of osteoarthritis: focus on the clinical use of hyaluronic acid. Drugs R D 2011; 11: 13-27.
- Lee WS, Kim HJ, Kim KI, Kim GB, Jin W. Intra-Articular Injection of Autologous Adipose Tissue-Derived Mesenchymal Stem Cells for the Treatment of Knee Osteoarthritis: A Phase IIb, Randomized, Placebo-Controlled Clinical Trial. Stem Cells Transl Med 2019; 8: 504-511.
- 44. Sorushanova A, Delgado LM, Wu Z, Shologu N, Kshirsagar A, Raghunath R, Mullen AM, Bayon Y, Pandit A, Raghunath M, Zeugolis DI. The Collagen Suprafamily: From Biosynthesis to Advanced Biomaterial Development. Adv Mater 2019; 31: e1801651.
- 45. Shojaati G, Khandaker I, Sylakowski K, Funderburgh ML, Du Y, Funderburgh JL. Compressed Collagen Enhances Stem Cell Therapy for Corneal Scarring. Stem Cells Transl Med 2018; 7: 487-494.
- Elahi FM, Farwell DG, Nolta JA, Anderson JD. Preclinical translation of exosomes derived from mesenchymal stem/stromal cells. Stem Cells 2020; 38: 15-21.
- 47. Whitford W, Guterstam P. Exosome manufacturing status. Future Med Chem 2019; 11: 1225-1236.
- 48. Chen YS, Lin EY, Chiou TW, Harn HJ. Exosomes in clinical trial and their production in compliance with good manufacturing practice. Ci Ji Yi Xue Za Zhi 2019; 32: 113-120.
- Mendt M, Kamerkar S, Sugimoto H, McAndrews KM, Wu CC, Gagea M, Yang S, Blanko EVR, Peng Q, Ma X, Marszalek JR, Maitra A, Yee C, Rezvani K, Shpall E, LeBleu VS, Kalluri R. Generation and testing of clinical-grade exosomes for pancreatic cancer. JCI Insight 2018; 3: e99263.
- 50. Dai J, Su Y, Zhong S, Cong L, Liu B, Yang J, Tao Y, He Z, Chen C, Jiang Y. Exosomes: key players in cancer and potential therapeutic strategy. Signal Transduct Target Ther. 2020; 5: 145.

- Sjöqvist S, Ishikawa T, Shimura D, Kasai Y, Imafuku A, Bou-Ghannam S, Iwata T, Kanai N. Exosomes derived from clinical-grade oral mucosal epithelial cell sheets promote wound healing. J Extracell Vesicles 2019; 8: 1565264.
- 52. Meng W, He C, Hao Y, Wang L, Li L, Zhu G. Prospects and challenges of extracellular vesicle-based drug delivery system: considering cell source. Drug Deliv 2020; 27: 585-598.
- 53. Sarvarian P, Samadi P, Gholipour E, Shams Asenjan K, Hojjat-Farsangi M, Motavalli R, Motavalli Khiavi F, Yousefi M. Application of Emerging Plant-Derived Nanoparticles as a Novel Approach for Nano-Drug Delivery Systems. Immunol Invest 2022; 51: 1039-1059.
- 54. Xiao J, Feng S, Wang X, Long K, Luo Y, Wang Y, Ma J, Tang Q, Jin L, Li X, Li M. Identification of exosome-like nanoparticle-derived microRNAs from 11 edible fruits and vegetables. PeerJ 2018; 6: e5186.
- Dad HA, Gu TW, Zhu AQ, Huang LQ, Peng LH. Plant Exosome-like Nanovesicles: Emerging Therapeutics and Drug Delivery Nanoplatforms. Mol Ther 2021; 29: 13-31.
- 56. Pereira-Arias JG, Gamarra-Quintanilla M, Urdaneta-Salegui LF, Mora-Christian JA, Sánchez-Vazquez A, Astobieta-Odriozola A, Ibarluzea-González G. Current status of extracorporeal shock wave lithotripsy in urinary lithiasis. Arch Esp Urol 2017; 70: 263-287.
- Yao G, Chen J, Duan Y, Chen X. Efficacy of Extracorporeal Shock Wave Therapy for Lateral Epicondylitis: A
 Systematic Review and Meta-Analysis. Biomed Res Int
 2020; 2020: 2064781.
- Schroeder AN, Tenforde AS, Jelsing EJ. Extracorporeal Shockwave Therapy in the Management of Sports Medicine Injuries. Curr Sports Med Rep 2021; 20: 298-305.
- 59. Auersperg V, Trieb K. Extracorporeal shock wave therapy: an update. EFORT Open Rev 2020; 5: 584-592.
- Wang FS, Yang KD, Kuo YR, Wang CJ, Sheen-Chen SM, Huang HC, Chen YJ. Temporal and spatial expression of bone morphogenetic proteins in extracorporeal shock wave-promoted healing of segmental defect. Bone 2003; 32: 387-396.
- 61. Ogden JA, Tóth-Kischkat A, Schultheiss R. Principles of shock wave therapy. Clin Orthop Relat Res 2001; 387: 8-17.
- 62. Cheng JH, Wang CJ. Biological mechanism of shockwave in bone. Int J Surg 2015; 24: 143-146.
- 63. Yu L, Liu S, Zhao Z, Xia L, Zhang H, Lou J, Yang J, Xing G, Xing G. Extracorporeal Shock Wave Rebuilt Subchondral Bone In Vivo and Activated Wnt5a/Ca2+ Signaling In Vitro. Biomed Res Int 2017; 2017: 1404650.
- O'Connell B, Wragg NM, Wilson SL. The use of PRP injections in the management of knee osteoarthritis. Cell Tissue Res 2019; 376: 143-152.
- 65. Wu PI, Diaz R, Borg-Stein J. Platelet-Rich Plasma. Phys Med Rehabil Clin N Am 2016; 27: 825-853.
- Grassi A, Napoli F, Romandini I, Samuelsson K, Zaffagnini S, Candrian C, Filardo G. Is Platelet-Rich Plasma (PRP) Effective in the Treatment of Acute Muscle Injuries? A Systematic Review and Meta-Analysis. Sports Med 2018; 48: 971-989.
- 67. Mlynarek RA, Kuhn AW, Bedi A. Platelet-Rich Plasma (PRP) in Orthopedic Sports Medicine. Am J Orthop (Belle Mead NJ) 2016; 45: 290-326.

- 68. Andia I. Platelet-rich plasma biology; In Alves R, Grimalt R (eds): Clinical Indications and Treatment Protocols with Platelet-Rich Plasma in Dermatology. Barcelona, Ediciones Mayo, 2016; pp. 3-15.
- Blair P, Flaumenhaft R. Platelet alpha-granules: basic biology and clinical correlates. Blood Rev 2009; 23: 177-189
- Maynard DM, Heijnen HF, Horne MK, White JG, Gahl WA. Proteomic analysis of platelet alpha-granules using mass spectrometry. J Thromb Haemost 2007; 5: 1945-1955
- McNicol A, Israels SJ. Platelet dense granules: structure, function and implications for haemostasis. Thromb Res 1999; 95: 1-18.
- Thon JN, Italiano JE. Platelets: production, morphology and ultrastructure. Handb Exp Pharmacol 2012; 210: 3-22.
- 73. van Nispen tot Pannerden H, de Haas F, Geerts W, Posthuma G, van Dijk S, Heijnen HF. The platelet interior revisited: electron tomography reveals tubular alphagranule subtypes. Blood 2010; 116: 1147-1156.
- 74. Andia I, Maffulli N. Blood-Derived Products for Tissue Repair/Regeneration. Int J Mol Sci 2019; 20: 4581.
- 75. Amable PR, Carias RB, Teixeira MV, da Cruz Pacheco I, Corrêa do Amaral RJ, Granjeiro JM, Borojevic R. Platelet-rich plasma preparation for regenerative medicine: optimization and quantification of cytokines and growth factors. Stem Cell Res Ther 2013; 4: 67.
- Lubkowska A, Dolegowska B, Banfi G. Growth factor content in PRP and their applicability in medicine. J Biol Regul Homeost Agents 2012; 26: 3S-22S.
- Mishra A, Woodall J Jr, Vieira A. Treatment of tendon and muscle using platelet-rich plasma. Clin Sports Med 2009; 28: 113-125.
- Hollands P, Aboyeji DR, Ovokaitys T. The action of modulated laser light on Human Very Small Embryonic-Like (hVSEL) stem cells in Platelet Rich Plasma (PRP). CellR4 2020; 8: e2990.
- Sovalat H, Scrofani M, Eidenschenk A, Hénon P. Human Very Small Embryonic-Like Stem Cells Are Present in Normal Peripheral Blood of Young, Middle-Aged, and Aged Subjects. Stem Cells Int 2016; 2016: 7651645.
- 80. Zuba-Surma EK, Kucia M, Dawn B, Guo Y, Ratajczak MZ, Bolli R. Bone marrow-derived pluripotent very small embryonic-like stem cells (VSELs) are mobilized after acute myocardial infarction. J Mol Cell Cardiol 2008; 44: 865-873.
- 81. Sovalat H, Scrofani M, Eidenschenk A, Pasquet S, Rimelen V, Hénon P. Identification and isolation from either adult human bone marrow or G-CSF-mobilized peripheral blood of CD34(+)/CD133(+)/CXCR4(+)/Lin(-)CD45(-) cells, featuring morphological, molecular, and phenotypic characteristics of very small embryonic-like (VSEL) stem cells. Exp Hematol 2011; 39: 495-505.
- 82. Augustine TN, van der Spuy WJ, Kaberry LL, Shayi M. Thrombin-Mediated Platelet Activation of Lysed Whole Blood and Platelet-Rich Plasma: A Comparison Between Platelet Activation Markers and Ultrastructural Alterations. Microsc Microanal 2016; 22: 630-639.

- 83. Zemel'ko VI, Grinchuk TM, Domnina AP, Artsybasheva IV, Zenin VV, Kirsanov AA, Bichevaia NK, Korsak VS, Nikol'skiĭ NN. Multipotent mesenchymal stem cells of desquamated endometrium: isolation, characterization and use as feeder layer for maintenance of human embryonic stem cell lines. Tsitologiia 2011; 53: 919-929.
- 84. Liao W, Huang N, Yu J, Jares A, Yang J, Zieve G, Avila C, Jiang X, Zhang XB, Ma Y. Direct Conversion of Cord Blood CD34+ Cells Into Neural Stem Cells by OCT4. Stem Cells Transl Med 2015; 4: 755-763.
- 85. Shimizu M, Natori T, Tsuda K, Yoshida M, Kamada A, Oi K, Ishigaku Y, Oura K, Narumi S, Yamamoto M, Terayama Y. Thrombin-induced platelet aggregation -effect of dabigatran using automated platelet aggregometry. Platelets 2020; 31: 360-364.
- 86. Pedroni ACF, Diniz IMA, Abe GL, Moreira MS, Sipert CR, Marques MM. Photobiomodulation therapy and vitamin C on longevity of cell sheets of human dental pulp stem cells. J Cell Physiol 2018; 233: 7026-7035.
- 87. Brindley J, Hollands P, Ovokaitys T. A Theoretical Mechanism for the Action of SONG-Modulated Laser Light on Human Very Small Embryonic-Like (hVSEL) Stem Cells in Platelet Rich Plasma (PRP) CellR4 2021; 9: e3201.
- 88. Chen FM, Wu LA, Zhang M, Zhang R, Sun HH. Homing of endogenous stem/progenitor cells for in situ tissue regeneration: Promises, strategies, and translational perspectives. Biomaterials 2011; 32: 3189-209.