DOI: 10.32113/cellr4 20219 3248 CellR4 2021; 9: e3248

Cord blood technology: a new paradigm

P. Hollands

Consultant Clinical Scientist, Cambridge, UK

Corresponding Author: Peter Hollands, PhD (Cantab), FRSB; e-mail: peterh63@hotmail.com

Keywords: Cord blood, Cord blood collection, Cord blood processing, Cord blood storage, Cord blood transplantation, Evo3medica Sterile Closed Cryo-System (ESCCS), Regenerative medicine, Stem cells.

Abstract

Objective: This opinion paper examines the current status of cord blood stem cell technology using volume reduction and describes a novel cord blood collection, processing and storage system. This technology could represent a new paradigm in using cord blood in regenerative medicine in the future. The decline in the use of cord blood to treat blood disorders, largely driven by haploidentical transplantation, may in the future be replaced by using cord blood in regenerative medicine procedures. The Evo-3medica Sterile Closed Cryo-System (ESCCS) provides a closed cord blood collection, processing and storage system which will considerably reduce the cost of cord blood collection, processing and storage. The ESCCS enables easy collection, processing and storage of whole cord blood, with all of its constituent cells available for future regenerative medicine procedures. This will enable current cord blood banks to easily and cost-effectively switch their focus to regenerative medicine procedures. This is instead of focusing on the ever-decreasing use of cord blood in haemopoietic stem cell transplantation for blood disorders.

Introduction

Cord blood was first used as a unique source of haemopoietic stem cells (HSCs) for transplantation in 1988. It was used to treat a young patient who was suffering from Fanconi anaemia¹. This patient was treated using whole cord blood (without volume reduction). The cord blood came from his sibling and the patient is alive and well today. There is currently an unrelated donor registry of cryopreserved cord blood which has approximately 645,646 cord blood units available globally for transplantion², with 300,000 cord blood units available in the USA alone³. There have even been suggestions that allogeneic cord blood transplantation may become a first line treatment for paediatric leukaemia and related blood disorders⁴. Cord blood may also be transplanted with a less than perfect Human Leucocyte Antigen (HLA) match making it more flexible than bone marrow as a transplant product^{5,6}. Cord blood transplantation in adults is not so attractive since cord blood contains a limited number of HSCs and transplantation to adults either requires stem cell expansion⁷ or the co-transplantation of multiple matched cord blood units (typically 2-3) to reach the required dose of HSCs⁸.

CORD BLOOD COLLECTION, PROCESSING AND CLINICAL USE

Cord blood collection is currently carried out either IN UTERO or EX UTERO with a preference towards EX UTERO in most countries. This is because EX UTERO collection seems to provide lower levels of bacterial contamination and less incidence of low volume and clotting9. Cord blood is usually collected by a specifically trained phlebotomist into a 250 mL bag containing citrate-phosphate dextrose solution (CPD)¹⁰ as an anticoagulant; the collected cord blood is stable for at least 48 hours before processing and storage¹¹.

On arrival at the processing laboratory almost



○ ① S ② This work is licensed under a Creative Commons Attribution-NonCommercial-ShareAlike 4.0 International License

all cord blood undergoes "volume reduction" to obtain the buffy coat, containing CD34+ HSCs, and a total volume of 25 mL is cryopreserved. This volume reduction is achieved by the use of automated systems such as SEPAX by Cytiva (Little Chalfont, Buckinghamshire, UK) or AXP by ThermoGenesis (Rancho Cordova, CA, USA)¹². The rationale for storing volume-reduced cord blood is to reduce the required storage space in the cord blood bank. A 25 mL frozen cord blood unit takes up much less space in the storage tank than a 150 mL frozen cord blood unit¹³. The first cord blood transplant, and many subsequent transplants, used whole blood (without volume reduction) for transplantation with no adverse events^{14,15}.

The volume reduction processing of cord blood must take place in an EU GMP Grade B (ISO Class 5) clean room and open procedures such as the addition of the cryoprotectant (10% v/v Dimethyl sulfoxide, DMSO) are carried out in a EU GMP Grade A Class II flow hood (ISO Class 5)16. Cord blood is usually stored in liquid nitrogen at -196°C (sometimes it is stored in the vapour phase at -180°C) and thawed rapidly in a 37°C water-bath in the transplant center when needed for transplant. These facilities in cord blood banks are expensive to create, run and staff but they are a regulatory requirement in most countries¹⁷. The cost of this advanced technology is handed on to the patient in a private cord blood bank or the charity or hospital in a public cord blood bank.

Cord blood has been transplanted over 35,000 times worldwide to treat a wide range of blood disorders which are often referred to as '80 different diseases' (they are in fact all in one group i.e., blood disorders)¹⁸. This use of cord blood to treat blood disorders has been on the decline in recent years because the option of haploidentical bone marrow transplantation has proved a great success^{19,20}. If the demand for cord blood stem cells to transplant for blood diseases continues to decline, and as regenerative medicine procedures become more common, then preparations must begin to enable cord blood to be used in regenerative medicine.

CEREBRAL PALSY AND AUTISM

The most prominent 'alternative' current use for cord blood is the use of autologous volume reduced cord blood buffy coat in the treatment of cerebral palsy (CP)²¹ and autism spectrum disorder²². The results for CP have shown some signs of benefit

but much more work is needed²¹. The results for autism spectrum disorder are currently less encouraging²². It is also uncertain as to which type of cell, if any, is contributing to the possible benefits which have been seen. Cord blood contains HSCs²³, mesenchymal stem cells (MSCs)²⁴, endothelial progenitor cells (EPCs)^{25,26}, regulatory T cells (Treg)²⁷, myeloid-derived suppressor cells (MDSCs)²⁸, unrestricted somatic stem cells (USSCs)²⁹ and pluripotent very small embryonic-like (VSEL)³⁰ stem cells. Is it not known which of these cells, if any, are contributing to the possible benefit in CP. Only those stem cells which can cross the blood-brain barrier can provide a cellular-based benefit within the central nervous system. One possible explanatory hypothesis is that stem cells from the cord blood, when administered intravenously, may become trapped in capillary beds (e.g., in the lungs)³¹ and from there the trapped cells release growth factors³² and/or exosomes³³ which can cross the bloodbrain barrier.

A type of stem cells present in cord blood, which are currently ignored by many workers, is represented by the pluripotent VSEL stem cells^{34,35}. It is possible that pluripotent VSEL stem cells in cord blood can cross the blood brain barrier and carry out repair within the central nervous system. Therefore, these cells could well be the active component in cord blood buffy coat resulting in the benefits seen so far in the treatment of CP.

PUBLIC AND PRIVATE CORD BLOOD BANKS

There are two basic models for the collection, storage and use of cord blood. The public sector has altruistic cord blood donors providing cord blood, which is processed, stored and tissue typed for use by anyone in need³⁶. The HLA databases of these cord blood banks are available to be searched by transplant physicians around the world thus optimising the cord blood availability. The public cord blood banks generate income from the fee charged to release a cord blood unit for transplant. The private cord blood bank sector collects, processes and stores cord blood for use by individual families only³⁷. Private cord blood banks generate their income from fees for initial collection, processing and storage, annual storage fees and fees for release when a cord blood unit is needed for transplantation in the family. Both public and private cord blood banks currently use a standard 250 mL blood bag containing CPD anticoagulant to collect cord

blood and use volume reduction to 25 mL of buffy coat prior to cryopreservation and storage.

The Use of Cord Blood in Regenerative Medicine

Cord blood is a potential source of stem cells to use in regenerative medicine procedures especially in the areas of cardiovascular, ophthalmic, neurological and endocrine diseases³⁸. The range of stem cell types found in cord blood provides this wide field of potential applications and the presence of MSCs makes the cord blood a potentially important source of these cells which are already showing considerable potential in musculoskeletal diseases³⁹.

In addition to the cellular components of cord blood, the plasma of cord blood has been shown to have important properties which may be the basis of, or play a supporting role in, future regenerative medicine therapies⁴⁰. Emerging technologies such as *EX VIVO* gold treatment of blood using GOLDIC® (Gmund am Tegernsee, Germany) may also enhance the regenerative potential of cord blood plasma^{41,42}. GOLDIC® technology seems to be capable of increasing levels of gelsolin in the blood which has potential diagnostic and therapeutic applications⁴³.

EVO3MEDICA STERILE CLOSED CRYO-SYSTEM (ESCCS)

The current declining use of cord blood as a source of HSCs for the treatment of blood disorders and the increasing potential future use in regenerative medicine procedures raises questions about the current 'gold-standard' of cord blood volume reduction¹². The present system reduces the cord blood volume to 25 mL, and plasma and cells are as a result discarded as medical waste. The concern is that if cord blood becomes an important source of stem cells for regenerative medicine in the future, then volume reduction may have removed some of the critical cells needed. It therefore seems prudent to store whole cord blood for future regenerative medicine procedures to avoid the loss of critical cord blood components and cells. In addition, the volume reduction process is expensive because it requires high levels of technology, clean room facilities and highly trained staff. Storage of whole cord blood would reduce this cost, making cord blood technology more affordable both for patients and institutions across the globe.

The Evo3medica Sterile Closed Cryo-System (ESCCS; Leipzig, Sachsen, Germany) is a new closed cord blood collection, processing and storage system for whole cord blood, which does not require either a clean room facility or expensive processing equipment (Figure 1). It also does not require highly trained processing staff. The cord blood is collected in the same way as when a single 250 mL blood bag is used and sent to the laboratory for processing. When using the ESCCS, the DMSO is added through a sterile DMSO filter directly to

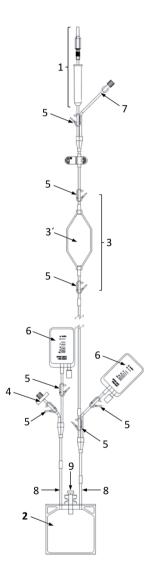


Figure 1. The Evo3medica Sterile Closed Cryo-System (ESCCS). 1. Collection needle; 2. Combined mixing and freezing bag; 3. Intermediate citrate-phosphate dextrose solution (CPD) reservoir; 4. Dimethyl sulfoxide (DMSO) Inlet/sterile filter; 5. Tube clips; 6. Satellite bags; 7. Injection port; 8. Main interconnector; 9. Storage Bag Opening Port (to use after thawing).

the whole blood. There are two satellite bags in the ESCCS system. The first bag is used to take the blood sample for infectious disease serology and flow cytometry. The second bag is used to take a small blood sample plus DMSO (analogous to the small compartment of a 25 mL pall bag) which can be stored with the main sample and used for testing at the time of transplant. All of these procedures can be carried out in a processing room without air filtration because the system is totally closed. The processing of cord blood is much quicker and cheaper using this technology and it creates less medical waste than volume reduction. The collection, freezing and storage of cord blood using the ESCCS all take place in a single Ethylene-Vinyl Acetate (EVA) bag. The ESCCS also has a novel CPD anticoagulant handling system which coats the tubing and bag before blood collection, thus minimising the risk of clotting during collection and transport.

The use of the ESCCS reduces the cost and time taken to process the cord blood and it retains all of the blood cells present, so that they can be used in future regenerative medicine procedures. This is a new paradigm which may change and improve the processing and storage of cord blood for future regenerative medicine procedures. It will reduce the cost of processing and provides a critical undepleted cord blood unit for future use.

It is theoretically possible to use technology such as the Terumo sterile tube welder to connect all of the separate processing components to the collection bag in the laboratory. This is less attractive than a pre-manufactured processing system such as the ESCCS because the tube welds may possibly introduce contamination, it requires more steps in the processing laboratory, it takes more time to carry out processing and the cost is increased because each component has to be sourced, purchased and validated separately. At the time of writing there were no publications on this system for routine cord blood processing.

There is a similar system to the ESCCS developed by Vita34 and called the Decentralised System (DeSy), which is protected by a patent⁴⁴. This follows a traditional multi-bag system

in contrast to the advanced single-bag system used by the ESCCS. The DeSy technology, however, does not seem to play a substantial role in the overall strategy of Vita34.

In contrast, Evo3medica follows the world-wide

vision that every family should be able to participate in medical progress, in general, and in the huge opportunities of regenerative medicine, in particular. Therefore, Evo3medica aims at a significant market penetration behind a global network of local laboratories which provide direct access and reasonable pricing to the customers. The ESCCS is patented in Europe (EP2889047A1)⁴⁵.

CONCLUSIONS

The cord blood industry has come to an inflexion point. The clinical use of cord blood to treat blood disorders is on the decline and the future lies in regenerative medicine. The present volume reduction of cord blood is expensive and may be resulting in the loss of essential cells and plasma which could be very important in future regenerative medicine procedures. The ESCCS provides closed whole cord blood processing and storage (without the need for a clean room) thus removing the need for expensive volume reduction. In addition, the ESCCS retains every cell contained in cord blood to optimise future use in regenerative medicine procedures. Storage facilities may have to increase capacity to accommodate larger stored volumes, but the future benefits of this whole cord blood storage using the ESCCS will far outweigh this additional storage requirement.

FUNDING:

No funding is declared for this article.

ORCID:

Peter Hollands: https://orcid.org/0000-0003-4116-1954

CONFLICT OF INTEREST:

Peter Hollands declares that he has no conflict of interest to disclose.

REFERENCES

- 1. Ballen KK, Gluckman E, Broxmeyer HE. Umbilical cord blood transplantation: the first 25 years and beyond. Blood 2013; 122: 491-498.
- Gratwohl A, Pasquini MC, Aljurf M, Atsuta Y, Baldomero H, Foeken L, Gratwohl M, Bouzas LF, Confer D, Frauendorfer K, Gluckman E, Greinix H, Horowitz M, Iida M, Lipton J, Madrigal A, Mohty M, Noel L, No-

- vitzky N, Nunez J, Oudshoorn M, Passweg J, van Rood J, Szer J, Blume K, Appelbaum FR, Kodera Y, Niederwieser D. Worldwide Network for Blood and Marrow Transplantation (WBMT). One million haemopoietic stem-cell transplants: a retrospective observational study. Lancet Haematol 2015; 2: e91-100. doi: 10.1016/S2352-3026(15)00028-9. Erratum in: Lancet Haematol. 2015; 2: e184. PMID: 26687803.
- Weiss ML, Troyer DL. Stem cells in the umbilical cord. Stem Cell Rev 2006; 2: 155-162.
- Tse W, Bunting KD, Laughlin MJ. New insights into cord blood stem cell transplantation. Curr Opin Hematol 2008; 15: 279-284.
- Kurtzberg J, Lyerly AD, Sugarman J. Untying the Gordian knot: policies, practices, and ethical issues related to banking of umbilical cord blood. J Clin Invest 2005; 115: 2592-2597.
- Laughlin MJ, Barker J, Bambach B, Koc ON, Rizzieri DA, Wagner JE, Gerson SL, Lazarus HM, Cairo M, Stevens CE, Rubinstein P, Kurtzberg J. Hematopoietic engraftment and survival in adult recipients of umbilical-cord blood from unrelated donors. N Engl J Med 2001; 344: 1815-1822.
- Baron F, Ruggeri A, Nagler A. Methods of ex vivo expansion of human cord blood cells: challenges, successes and clinical implications. Expert Rev Hematol 2016; 9: 297-314.
- Stanevsky A, Shimoni A, Yerushalmi R, Nagler A. Cord blood stem cells for hematopoietic transplantation. Stem Cell Rev Rep 2011; 7: 425-433.
- Lasky LC, Lane TA, Miller JP, Lindgren B, Patterson HA, Haley NR, Ballen K. In utero or ex utero cord blood collection: which is better? Transfusion 2002; 42: 1261-1267.
- McCullough J, Weiblen BJ. Citrate phosphate dextrose (CPD) anticoagulant in blood transfusion. Minn Med 1973; 56: 980-982.
- 11. Tron de Bouchony E, Pelletier D, Alcalay D, Bruneau J, Predeau M, Brizard A, Magnin M. Hematopoietic progenitor content of fetal cord blood collected using citrate-phosphate-dextrose: influence of holding temperature and delays. J Hematother 1993; 2: 271-273.
- Solves P, Mirabet V, Roig R. Volume reduction in routine cord blood banking. Curr Stem Cell Res Ther. 2010; 5: 362-366.
- Fasouliotis SJ, Schenker JG. Human umbilical cord blood banking and transplantation: a state of the art. Eur J Obstet Gynecol Reprod Biol 2000; 90: 13-25.
- 14. Munoz J, Shah N, Rezvani K, Hosing C, Bollard CM, Oran B, Olson A, Popat U, Molldrem J, McNiece IK, Shpall EJ. Concise review: umbilical cord blood transplantation: past, present, and future. Stem Cells Transl Med 2014; 3: 1435-1443.
- 15. Kögler G, Somville T, Göbel U, Hakenberg P, Knipper A, Fischer J, Adams O, Krempe C, McKenzie C, Rüttgers H, Meier W, Bellmann O, Streng H, Ring A, Rosseck U, Rocha V, Wernet P. Haematopoietic transplant potential of unrelated and related cord blood: the first six years of the EUROCORD/NETCORD Bank Germany. Klin Padiatr 1999; 211: 224-232.

- 16. Eichler H, Meckies J, Schmut N, Kern S, Klüter H, Zieger W. Präparative und arzneimittelrechtliche Aspekte bei der Sammlung von Stammzellpräparaten aus Plazentarestblut 1 [Aspects of donation and processing of stem cell transplants from umbilical cord blood]. Z Geburtshilfe Neonatol 2001; 205: 218-223.
- 17. Querol S, Gomez S, Pagliuca A, Torradella M, Madrigal JA. Quality rather than quantity: the cord blood bank dilemma. Bone Marrow Transplant 2010; 45: 970-978.
- Huang X, Guo B, Capitano M, Broxmeyer HE. Past, present, and future efforts to enhance the efficacy of cord blood hematopoietic cell transplantation. F1000Res 2019; 8: F1000.
- Oevermann L, Schulte JH, Hundsdörfer P, Hakimeh D, Kogel F, Lang P, Corbacioglu S, Eggert A, Sodani P. HLA-haploidentical hematopoietic stem cell transplantation in pediatric patients with hemoglobinopathies: current practice and new approaches. Bone Marrow Transplant 2019; 54: 743-748.
- Sugita J. HLA-haploidentical stem cell transplantation using posttransplant cyclophosphamide. Int J Hematol 2019; 110: 30-38.
- 21. McDonald CA, Fahey MC, Jenkin G, Miller SL. Umbilical cord blood cells for treatment of cerebral palsy; timing and treatment options. Pediatr Res 2018; 83: 333-344. doi: 10.1038/pr.2017.236 PMID: 28937975.
- 22. Dawson G, Sun JM, Baker J, Carpenter K, Compton S, Deaver M, Franz L, Heilbron N, Herold B, Horrigan J, Howard J, Kosinski A, Major S, Murias M, Page K, Prasad VK, Sabatos-DeVito M, Sanfilippo F, Sikich L, Simmons R, Song A, Vermeer S, Waters-Pick B, Troy J, Kurtzberg J. A Phase II Randomized Clinical Trial of the Safety and Efficacy of Intravenous Umbilical Cord Blood Infusion for Treatment of Children with Autism Spectrum Disorder. J Pediatr 2020; 222: 164-173.e5. doi: 10.1016/j.jpeds.2020.03.011. PMID: 32444220.
- 23. Hows J, Nicol A, Denning-Kendall P, Donaldson C, Nieda M, Bradley B. Cord blood as an alternative source of haemopoietic stem cells. Ann Oncol 1996; 7: 47-51.
- 24. Ding DC, Shyu WC, Lin SZ. Mesenchymal stem cells. Cell Transplant 2011; 20: 5-14.
- 25. Ingram DA, Mead LE, Tanaka H, Meade V, Fenoglio A, Mortell K, Pollok K, Ferkowicz MJ, Gilley D, Yoder MC. Identification of a novel hierarchy of endothelial progenitor cells using human peripheral and umbilical cord blood. Blood 2004; 104: 2752-2760.
- 26. Ravishankar P, Zeballos MA, Balachandran K. Isolation of Endothelial Progenitor Cells from Human Umbilical Cord Blood. J Vis Exp 2017; 127: 56021.
- 27. Motwani K, Peters LD, Vliegen WH, El-Sayed AG, Seay HR, Lopez MC, Baker HV, Posgai AL, Brusko MA, Perry DJ, Bacher R, Larkin J, Haller MJ, Brusko TM. Human Regulatory T Cells From Umbilical Cord Blood Display Increased Repertoire Diversity and Lineage Stability Relative to Adult Peripheral Blood. Front Immunol 2020; 11: 611.
- 28. Dietz S, Schwarz J, Vogelmann M, Spring B, Molnár K, Orlikowsky TW, Wiese F, Holzer U, Poets CF, Gille C, Köstlin-Gille N. Cord blood granulocytic myeloid-derived suppressor cells impair monocyte T cell stimulato-

- ry capacity and response to bacterial stimulation. Pediatr Res 2019; 86: 608-615.
- 29. Schira J, Falkenberg H, Hendricks M, Waldera-Lupa DM, Kögler G, Meyer HE, Müller HW, Stühler K. Characterization of Regenerative Phenotype of Unrestricted Somatic Stem Cells (USSC) from Human Umbilical Cord Blood (hUCB) by Functional Secretome Analysis. Mol Cell Proteomics 2015;14: 2630-2643.
- 30. Chang YJ, Tien KE, Wen CH, Hsieh TB, Hwang SM. Recovery of CD45(-)/Lin(-)/SSEA-4(+) very small embryonic-like stem cells by cord blood bank standard operating procedures. Cytotherapy 2014; 16: 560-565.
- 31. Zhang C, Tan X, Tan L, Liu T, Liu D, Zhang L, Fan S, Su Y, Cheng T, Zhou Y, Shi C. Labeling stem cells with a near-infrared fluorescent heptamethine dye for noninvasive optical tracking. Cell Transplant 2011; 20: 741-751.
- 32. Lee H, Lee SI, Ko Y, Park JB. Evaluation of the secretion and release of vascular endothelial growth factor from two-dimensional culture and three-dimensional cell spheroids formed with stem cells and osteoprecursor cells. Adv Clin Exp Med 2018; 27: 971-977.
- 33. Ma J, Zhao Y, Sun L, Sun X, Zhao X, Sun X, Qian H, Xu W, Zhu W. Exosomes derived from akt-modified human umbilical cord mesenchymal stem cells improve cardiac regeneration and promote angiogenesis via activating platelet-derived growth factor D. Stem Cells Transl Med 2017; 6: 51-59.
- Kuruca SE, Çelik DD, Özerkan D, Erdemir G. Characterization and Isolation of Very Small Embryonic-like (VSEL) Stem Cells Obtained from Various Human Hematopoietic Cell Sources. Stem Cell Rev Rep 2019; 15: 730-742.
- Ratajczak MZ, Suszynska M, Pedziwiatr D, Mierzejewska K, Greco NJ. Umbilical cord blood-derived very small embryonic like stem cells (VSELs) as a source of pluripotent stem cells for regenerative medicine. Pediatr Endocrinol Rev 2012; 9: 639-643.

- 36. Querol S, Rocha V. Procurement and Management of Cord Blood. In: Carreras E, Dufour C, Mohty M, Kröger N, editors. The EBMT Handbook: Hematopoietic Stem Cell Transplantation and Cellular Therapies. 7th ed. Cham (CH): Springer; 2019. Chapter 18. PMID: 32091821.
- Hollands P, McCauley C. Private cord blood banking: current use and clinical future. Stem Cell Rev Rep. 2009; 5: 195-203.
- 38. Harris DT, Badowski M, Ahmad N, Gaballa MA. The potential of cord blood stem cells for use in regenerative medicine. Expert Opin Biol Ther 2007; 7: 1311-1322.
- 39. Fuggle NR, Cooper C, Oreffo ROC, Price AJ, Kaux JF, Maheu E, Cutolo M, Honvo G, Conaghan PG, Berenbaum F, Branco J, Brandi ML, Cortet B, Veronese N, Kurth AA, Matijevic R, Roth R, Pelletier JP, Martel-Pelletier J, Vlaskovska M, Thomas T, Lems WF, Al-Daghri N, Bruyère O, Rizzoli R, Kanis JA, Reginster JY. Alternative and complementary therapies in osteoarthritis and cartilage repair. Aging Clin Exp Res 2020; 32: 547-560.
- 40. Romanov YA, Vtorushina VV, Dugina TN, Romanov AY, Petrova NV. Human umbilical cord blood serum/plasma: cytokine profile and prospective application in regenerative medicine. Bull Exp Biol Med 2019; 168: 173-177.
- 41. Schneider U, Lotzof K, Murrell WD, Goetz von Wachter E, Hollands P. Safety and efficacy of systemically administered autologous Gold-Induced Cytokines (GOLDIC®) CellR4 2021; 9: e3132. DOI: 10.32113/cellr4 20214 3132
- 42. Schneider U, Murrell WD, Hollands P. The regeneration of damaged connective tissue: wishful thinking or reality? CellR4 2021; 9: e3171. DOI: 10.32113/cellr4 20215 3171
- 43. Bucki R, Levental I, Kulakowska A, Janmey PA. Plasma gelsolin: function, prognostic value, and potential therapeutic use. Curr Protein Pept Sci 2008; 9: 541-551.
- 44. https://patents.google.com/patent/DE10151343A1/en (accessed June 24, 2021)
- 45. https://patents.google.com/patent/EP2889047A1/en (accessed June 24, 2021)