

Treatment of Osteoarthritis with Autologous and Allogeneic Expanded Bone Marrow Mesenchymal Stem Cells

P. Hollands¹, D. Porter²

¹Freelance Consultant Clinical Scientist, Cambridge, UK

²Medical Director, Opus Biological, London, UK

Corresponding Author: Peter Hollands, PhD; e-mail: peterh63@hotmail.com

Keywords: Allogeneic, Autologous, Bone marrow, Expansion, Mesenchymal stem cells, Osteoarthritis.

ABSTRACT

In this review we explore the past, present and future treatment of osteoarthritis using autologous and allogeneic bone marrow mesenchymal stem cells (MSCs). Osteoarthritis is one of the most prevalent joint diseases worldwide. It causes pain, loss of function and may lead to disability. At a cellular level, osteoarthritis causes biochemical changes in the composition of cartilage leading to progressive tissue degeneration. The majority of conventional treatments involve symptom control but offer only modest clinical benefits without any reversal of the cellular degeneration. Cell-based therapies in animal models have shown encouraging results and there are now a number of human case reports, pilot studies and follow-up studies that demonstrate the reversal of lesion formation. Opus Biological has designed a therapy and follow-up algorithm utilizing the feasibility and safety studies conducted in recent years to offer patients MSCs as a valid alternative to other conventional therapies for treatment of chronic osteoarthritis. MSC therapy for osteoarthritis does not require hospitalization, is a minimally invasive and low risk procedure, provides pain relief and significantly improves cartilage quality thereby enhancing joint function.

INTRODUCTION

Osteoarthritis (OA) is the most prevalent chronic joint disease as well as a frequent cause of joint pain, functional loss and disability^{1,2}. The disease most commonly affects the joints of the knees³, hands⁴, feet⁵ and spine⁶. It is also relatively common in shoulder⁷ and hip joints⁸. Even though OA is most commonly related to aging⁹, there are also a number of modifiable and non-modifiable risk factors for the development of this condition. These include obesity¹⁰, lack of exercise¹¹, genetic predisposition¹², bone mineral density status¹³, occupational injury¹⁴, trauma¹⁵ and female sex (due to menopause)¹⁶. Globally there is an estimated 10-15% prevalence of OA, with the numbers higher in the female population¹⁷. OA is an increasing risk to our global community due to the advances in medicine leading to an aging population and due to increases in the prevalence of modifiable risk factors such as obesity¹⁸. According to recent estimates, by 2050 the number of people aged over 60 will account for more than 20% of the world's population. By 2050, 130 million people will suffer from OA worldwide, and 40 million will be severely disabled by the disease. A World Health Organization report estimates that of 20% of those aged over 60, a conservative 15% will have symptomatic OA, and 33% of these people will be severely disabled by OA¹⁹. The cost to society of OA can be measured not only in the cost of adaptive aids and devices but also in the cost of medication, surgery²⁰, nursing care, residential care, and time off work and subsequent social care relating to sick



pay. International institutions have been reluctant to put a figure to the large indirect costs derived from the decrease in productivity caused by OA.

PATHOGENESIS OF OA

Although there are still many unknowns in the pathogenesis of OA, this appears to be primarily linked to biochemical and biomechanical changes in the joint cartilage²¹. The current models suggest that suboptimal nutrient (glucose)²² and oxygen supply²³, coupled with downregulation of extracellular/pericellular matrix components²⁴, increased supply of proteinases²⁵ and a relative increase in apoptosis of chondrocytes²⁶, accounts for a cumulative effect on the inability to withstand normal mechanical stresses. The resultant cartilage debris alongside catabolic mediators in the joint promote the activation of synovial macrophages²⁷, which in turn leads to synovial inflammation²⁸ that limits native cartilage repair²⁹.

CURRENT TREATMENT OF OA

The American Academy of Orthopaedic Surgeons recommends only physical³⁰ and educational therapy³¹, symptomatic drugs such as paracetamol³², non-steroidal anti-inflammatory drugs³³ and in some cases intra-articular corticosteroid injections³⁴. The American College of Rheumatology recommends a similar regimen including physical therapy, viscosupplementation³⁵ with hyaluronate injections³⁶, and possible but conflicting evidence for glucosamine³⁷ and/or chondroitin sulphate³⁸. Arthroscopic surgery³⁹ and acupuncture^{40,41} have only demonstrated modest clinical benefits. Another alternative treatment modality investigated over recent years is Autologous Chondrocyte Implantation (ACI)⁴². Briefly, chondrocytes are taken from patients, culture-expanded *in vitro* and then re-implanted back into the affected joints of patients. The procedure is invasive and has been shown to be less successful than the total knee replacement surgery to date⁴³.

MESENCHYMAL STEM CELLS

Mesenchymal stem cells (MSCs, sometimes also referred to as Mesenchymal Stromal Cells)^{44,45} were identified in the 1970s while examining the clonogenic potential of bone marrow cells. MSCs

were initially termed “colony-forming unit fibroblasts” (CFU-Fs)⁴⁶. Clinical trials investigating the use of MSCs began in 1995, with the results demonstrating no adverse reaction and no safety concerns, and since then hundreds of clinical trials have followed⁴⁷. In 2006, The International Society for Cellular Therapy (ISCT) published the criteria for defining MSCs⁴⁸. The ISCT stated that MSCs must be plastic-adherent under standard *in vitro* culture conditions; express CD105, CD73, and CD90, and lack expression of CD45, CD34, CD14 or CD11b, CD79a, CD19, and HLA-DR (as assessed by flow cytometry). In addition, the ISCT stated that MSCs must be able to differentiate into osteoblasts, adipocytes and chondrocytes *in vitro*⁴⁸. MSCs have been shown to be present in bone marrow⁴⁹, umbilical cord blood (with an associated clinical trial to treat cerebral palsy)⁵⁰, umbilical cord tissue⁵¹, placenta⁵², amniotic membrane⁵³, amniotic fluid⁵⁴, periosteum⁵⁵, trabecular bone⁵⁶, adipose tissue⁵⁷, synovium⁵⁸, skeletal muscle⁵⁹ and deciduous and permanent teeth⁶⁰. Independent of their origin, MSCs are capable of differentiating *in vitro* into different cell types of the connective tissue lineages such as bone, fat, muscle, tendon and ligament as well as cartilage⁶¹. MSCs have been shown to elicit immunosuppressive and immunomodulatory effects⁶² on T lymphocytes⁶³, B cells⁶⁴, dendritic cells (DCs)⁶⁵ and natural killer (NK) cells⁶⁶. These effects occur either via the cell-cell interaction route⁶⁷ or via the secretion of anti-inflammatory molecules such as indoleamine 2,3-dioxygenase (IDO)⁶⁸, prostaglandin E2 (PGE2)⁶⁹, interleukin-4 (IL-4)⁷⁰, interleukin-10 (IL-10)⁷¹ and transforming growth factor beta (TGF- β)⁷². These properties make MSCs the ideal choice for cell-based regenerative medicine procedures⁷³. The ability to differentiate *in vitro* into chondrocytes⁷⁴, combined with their anti-inflammatory and immunomodulatory properties, make MSCs the obvious choice for the treatment of diseases such as OA⁷⁵. Autologous⁷⁶ or allogeneic⁷⁷ bone marrow-derived MSCs are the most widely used MSCs in clinical research and treatment modalities across a plethora of disease indications, and are often considered to be the gold standard MSC type⁷⁸ because of the characterization that has occurred exhaustively over the last 5 decades⁷⁹.

Over the last 25 years the surgical implantation of autologous chondrocytes has been used to treat

local cartilage defects⁸⁰. MSCs have chondrogenic potential which is enhanced by co-culture with chondrocytes⁸¹. Co-cultured MSCs induce chondrocyte proliferation and extracellular matrix protein synthesis, including aggrecan⁸² and type II collagen⁸³. Therefore, MSCs can be used in place of chondrocytes for cartilage regeneration⁸⁴. The replacement of chondrocytes with MSCs is advantageous especially for diffuse chondral lesions as MSCs are more readily obtainable and can be expanded *in vitro*⁸⁵. A publication in 2010 argued on the merits of autologous bone marrow-derived MSCs *vs.* autologous chondrocyte implantation and observed that MSCs are as effective in cartilage repair as implanted chondrocytes. In addition, MSCs were more cost-effective, reduced donor site morbidity and resulted in one less knee surgery⁸⁶.

The philosophy of most current cell-based therapeutic approaches to the treatment of OA is to take the treatment to the site of the injury⁸⁷. Intra-articular injection of cell-based treatments such as Platelet-Rich Plasma (PRP)⁸⁸ or MSCs⁸⁹ allows these cells to interact with recipient cells and with the surface area within the joint space specifically targeting injured or degenerated tissues⁹⁰. MSCs are regulated by the microenvironment which they inhabit and by the interactions with the surrounding cells⁹¹, and they secrete bioactive factors for tissue regeneration in response to that microenvironment⁹². In the future it may also be possible to utilize the MSC-secretome to avoid the use of MSCs, thus potentially simplifying the treatment⁹³. Nevertheless, it is likely that the MSCs themselves may have more actions than simply the secretome alone in the overall tissue repair process⁹⁴.

The immunomodulatory effects of MSCs have been described above, but it is worth reminding that MSCs have a multimodal mechanism of action making them the “cell candidate of choice” in the treatment of OA. This is because MSCs stimulate native chondrogenic progenitor cells and their subsequent differentiation into mature chondrocytes mediated by bone morphogenetic proteins (BMPs)⁹⁵ and TGF- β 1⁹⁶. The differentiation of the MSCs themselves into chondrogenic progenitor cells is mediated by changes in the expression of regulatory genes like *Sox9*, *HoxA*, *HoxD* and *Gli3*⁹⁷. These chondrogenic progenitor cells then differentiate into mature chondrocytes with the ability to synthesize type II collagen, which maintains the structural integrity of hyaline cartilage⁹⁸.

AUTOLOGOUS MESENCHYMAL STEM CELLS

Animal studies have shown beneficial effects on cartilage repair utilizing autologous MSCs in rabbits⁹⁹, rats¹⁰⁰, pigs¹⁰¹ and guinea pigs¹⁰². The analgesic effect of MSC treatment in humans has been reported and this effect can be enhanced even further by the pre-treatment using the CB2 receptor agonist AM1241¹⁰³. The use of human autologous expanded bone marrow MSCs has been demonstrated to be effective in the treatment of chronic patellar tendinopathy¹⁰⁴ and in degenerative disc disease^{105,106}.

We have safely conducted intra-articular infusions of 40×10^6 autologous expanded bone marrow MSCs in patients with grade 2-4 OA of joints including the knee, hip, ankle, shoulder, and wrist. These treatments were carried out in the UK under a Medicines and Healthcare Products Regulatory Agency (MHRA) and a Human Tissue Authority (HTA) license. Expanded bone marrow MSCs are categorized to be Advanced therapy medicinal products (ATMPs)¹⁰⁷. The current research is highlighting the efficacy of expanded MSCs¹⁰⁸ in both patient outcome and duration of effect¹⁰⁹. The process of MSC expansion is, quite rightly, highly regulated and the MSCs generated are subject to rigorous quality and safety testing throughout manufacturing before they are released for treatment¹¹⁰.

The patients underwent follow up at 3-, 6-, 12- and 24-month intervals with outcome scores for pain and function at each stage. Clinical examination was also performed at the same time as utilizing validated outcome tools. Prior to treatment our patients underwent Magnetic resonance imaging (MRI) of the affected joint and at 12 months had a repeat MRI scan to discern radiological improvements. The patients were assessed using quantitative T2 mapping¹¹¹ to evaluate articular cartilage quality as T2 relaxation time is sensitive to changes in cartilage hydration and collagen fibril orientation¹¹². OA is known to cause an increase in T2 relaxation time¹¹³. T2 mapping and whole-organ magnetic resonance imaging score (WORMS)¹¹⁴ are currently the most common qualitative parameters for evaluation of cartilage regeneration, although the optimal imaging study would be able to provide an accurate assessment of cartilage thickness and volume, show morphologic changes to the cartilage surface, demonstrate internal cartilage signal changes and determine signal abnormalities

in subchondral bone. Orozco et al¹¹⁵ showed the safety and efficacy of autologous expanded bone marrow MSCs administered in 12 patients with chronic knee pain unresponsive to conservative treatments and radiologic evidence of OA. Quantification of cartilage quality by T2 relaxation measurements showed a highly significant reduction of poor cartilage areas, with improvement of cartilage quality in 11 of the 12 patients¹¹⁵.

AUTOLOGOUS EXPANDED BONE MARROW-DERIVED MSC TREATMENT: THE PROCEDURE

The procedure in the UK must begin with a Human Tissue Authority (HTA) Human Application License for the procurement and distribution of human cells or tissues. The patient receives conscious sedation, in a hospital operating theatre with local anaesthetic, and 60-100 mL of bone marrow is collected from the posterior iliac crest of the patient. The aspirate is then transferred to the expansion laboratory via an approved medical courier if the laboratory is not on the same site as the procurement operating theatre.

MSC ISOLATION AND EXPANSION

MSC isolation and expansion is typically performed in a laboratory under Good Manufacturing Practice (GMP) conditions and in the UK with license from the MHRA. The MSC expansion laboratory must also be licensed by the HTA UK for processing, storage and distribution of human cells and tissue for Human Application. MSC expansion is then carried out using either a manual technique¹¹⁶ or by the use of a closed system bioreactor¹¹⁷. In either case the phenotype and potency of the resultant MSCs must be retained. The manual expansion technique is slow and labour-intensive, whereas the bioreactor technology enables more cells to be produced in less time with less manual labour.

The expanded MSC product is tested for sterility, Mycoplasma and endotoxin¹¹⁸. The results of the quality control testing, together with the full review of the batch documentation and the results of all environmental monitoring that took place during the cell culture process, allows the intermediate product to be released for use as the final product. The final product is transported, within

temperature-controlled containers, by a dedicated medical courier service to the treatment center for administration to the patient.

ALLOGENEIC MSCs

Initially, the use of allogeneic MSCs was limited due to the concern that they would lose their immunogenicity and immunomodulatory properties on transplantation¹¹⁹. MSCs do not express class II human leukocyte antigen (HLA) molecules¹²⁰ and are therefore considered to be immune privileged¹²⁰ or possibly “immune evasive”¹²¹. The proposed use of allogeneic MSCs for clinical purposes has resulted in recent increases in the number of studies exploring the possibility of using allogeneic MSCs for the treatment of cartilage injuries¹²². Allogeneic MSCs have been combined with isolated autologous chondrocytes in a fibrin glue and applied to focal areas of cartilage damage¹²³. There have been no reported immunological issues from any of these studies. The use of allogeneic MSCs in the treatment of OA has demonstrated significant improvements in pain, leading to an overall improvement in joint function and suggesting cartilage regeneration on imaging¹²⁴. The use of allogeneic MSCs has a number of benefits for the patient. Allogeneic MSC treatment is less expensive and less time-consuming than the autologous version. The utilization of allogeneic MSCs removes certain key steps in the process, such as admission to a hospital facility and then an operating theatre procedure for bone marrow aspiration under conscious sedation.

CONCLUSIONS

We have a team which is a collective of unique and experienced healthcare professionals working together with one unified vision to bring the state of the art of stem cell treatment to patients as a safe and effective option for those seeking an alternative avenue to conventional treatments. Stem cell science is not new, in fact the technology is decades old, but only now are scientists and clinicians harnessing and understanding the potential of this avenue for a broad spectrum of diseases. We have seen the power of MSCs throughout our own practice in the treatment of musculoskeletal injuries and diseases. We work with the academic community and the regulators in the UK and the EU to adhere to the core of scientific

principle and abide by regulations that preserve our patients' safety. Our world leading scientific laboratory can tailor the treatments that we offer to those suffering from musculoskeletal injury and disease. MSC-based therapy provides a complete package of care on the orthobiologics spectrum, from PRP through allogeneic to autologous MSCs. We understand that not all of our patients will require MSC treatment, but we also understand that patients want relief from pain of OA and a betterment in their daily lives. The spectrum of orthobiologics now available can help our patients to experience pain free function alongside a renewed sense of health. In the age of "one size fits all" practice we want to put the power back into the hands of our patients. We deliver truly unique and individualized care for each patient, in partnership with world class professionals. In the future we aim to help patients suffering from an array of conditions across a multitude of specialties ranging from those with Crohn's disease¹²⁵ to patients experiencing the life limiting consequences of multiple sclerosis¹²⁶. Stem cell science can be the forefront of disease modification and, more importantly, disease reversal in such conditions.

FUNDING:

No funding is declared for this article.

AUTHOR CONTRIBUTIONS:

Peter Hollands: Conception and design of the manuscript, drafting the article and making critical revisions related to relevant intellectual content of the manuscript and final approval. David Porter: Conception and design of the manuscript, drafting the article and making critical revisions related to relevant intellectual content of the manuscript and final approval.

ORCID:

Peter Hollands: <https://orcid.org/0000-0003-4116-1954>

David Porter: <https://orcid.org/0000-0001-9659-0196>

CONFLICT OF INTEREST:

Peter Hollands has no conflict of interest to disclose. David Porter is Medical Director of Opus Biological (London, UK).

REFERENCES

- Arden N, Nevitt MC. Osteoarthritis: epidemiology. *Best Pract Res Clin Rheumatol* 2006; 20: 3-25. doi: 10.1016/j.berh.2005.09.007. PMID: 16483904.
- Vina ER, Kwok CK. Epidemiology of osteoarthritis: literature update. *Curr Opin Rheumatol* 2018; 30: 160-167. doi: 10.1097/BOR.0000000000000479. PMID: 29227353; PMCID: PMC5832048.
- Springer B, Bechler U, Waldstein W, Rueckl K, Boettner F. Five Questions to Identify Patients With Osteoarthritis of the Knee. *J Arthroplasty* 2020; 35: 52-56. doi: 10.1016/j.arth.2019.08.054. PMID: 31563394.
- Marshall M, Watt FE, Vincent TL, Dziedzic K. Hand osteoarthritis: clinical phenotypes, molecular mechanisms and disease management. *Nat Rev Rheumatol* 2018; 14: 641-656. doi: 10.1038/s41584-018-0095-4. PMID: 30305701.
- Rodríguez-Sanz D, Tovaruela-Carrión N, López-López D, Palomo-López P, Romero-Morales C, Navarro-Flores E, Calvo-Lobo C. Foot disorders in the elderly: A mini-review. *Dis Mon* 2018; 64: 64-91. doi: 10.1016/j.disamonth.2017.08.001. PMID: 28826743.
- Goode AP, Cleveland RJ, Schwartz TA, Nelson AE, Kraus VB, Hillstrom HJ, Hannan MT, Flowers P, Renner JB, Jordan JM, Golightly YM. Relationship of joint hypermobility with low back pain and lumbar spine osteoarthritis. *BMC Musculoskelet Disord* 2019; 20: 158. doi: 10.1186/s12891-019-2523-2. PMID: 30967130; PMCID: PMC6456963.
- Mehl J, Imhoff AB, Beitzel K. Omarthrose: Pathogenese, Diagnostik und konservative Therapieoptionen [Osteoarthritis of the shoulder: pathogenesis, diagnostics and conservative treatment options]. *Orthopade* 2018; 47: 368-376. doi: 10.1007/s00132-018-3542-7. PMID: 29464283.
- Rees HW. Management of Osteoarthritis of the Hip. *J Am Acad Orthop Surg* 2020; 28: e288-e291. doi: 10.5435/JAAOS-D-19-00416. PMID: 31800436.
- Sacitharan PK. Ageing and Osteoarthritis. *Subcell Biochem* 2019; 91: 123-159. doi: 10.1007/978-981-13-3681-2_6. PMID: 30888652.
- Kulkarni K, Karssiens T, Kumar V, Pandit H. Obesity and osteoarthritis. *Maturitas* 2016; 89: 22-28. doi: 10.1016/j.maturitas.2016.04.006. PMID: 27180156.
- Booth FW, Roberts CK, Laye MJ. Lack of exercise is a major cause of chronic diseases. *Compr Physiol* 2012; 2: 1143-1211. doi: 10.1002/cphy.c110025. PMID: 23798298; PMCID: PMC4241367.
- Yucesoy B, Charles LE, Baker B, Burchfiel CM. Occupational and genetic risk factors for osteoarthritis: a review. *Work* 2015; 50: 261-273. doi: 10.3233/WOR-131739. PMID: 24004806; PMCID: PMC4562436.
- Ishii Y, Noguchi H, Sato J, Ishii H, Todoroki K, Toyabe SI. Association between bone mineral density distribution and various radiographic parameters in patients with advanced medial osteoarthritis of the knee. *J Orthop Sci* 2019; 24: 686-692. doi: 10.1016/j.jos.2018.12.019. PMID: 30630770.
- Dulay GS, Cooper C, Dennison EM. Knee pain, knee injury, knee osteoarthritis & work. *Best Pract Res Clin Rheumatol* 2015; 29: 454-461. doi: 10.1016/j.berh.2015.05.005. PMID: 26612241.
- Lieberthal J, Sambamurthy N, Scanzello CR. Inflammation in joint injury and post-traumatic osteoarthritis. *Osteoarthritis Cartilage* 2015; 23: 1825-1834. doi: 10.1016/j.joca.2015.08.015. PMID: 26521728; PMCID: PMC4630675.
- Mahajan A, Patni R. Menopause and osteoarthritis: any association? *J Midlife Health* 2018; 9: 171-172. doi: 10.4103/jmh.JMH_157_18. PMID: 30692810; PMCID: PMC6332715.

17. Hunter DJ, Bierma-Zeinstra S. Osteoarthritis. *Lancet* 2019; 393: 1745-1759. doi: 10.1016/S0140-6736(19)30417-9. PMID: 31034380.
18. Musumeci G, Aiello FC, Szychlinska MA, Di Rosa M, Castrogiovanni P, Mobasher A. Osteoarthritis in the XXIst century: risk factors and behaviours that influence disease onset and progression. *Int J Mol Sci* 2015; 16: 6093-6112. doi: 10.3390/ijms16036093. PMID: 25785564; PMCID: PMC4394521.
19. de Wit M, Cooper C, Tugwell P, Bere N, Kirwan J, Conaghan PG, Roberts C, Aujoulat I, Al-Daghri N, Araujo de Carvalho I, Barker M, Bedlington N, Brandi ML, Bruyère O, Burt N, Halbout P, Hilgsmann M, Jiwa F, Kanis JA, Laslop A, Lawrence W, Pinto D, Prieto Yerro C, Rabenda V, Rizzoli R, Scholte-Voshaar M, Vlaskovska M, Reginster JY. Practical guidance for engaging patients in health research, treatment guidelines and regulatory processes: results of an expert group meeting organized by the World Health Organization (WHO) and the European Society for Clinical and Economic Aspects of Osteoporosis, Osteoarthritis and Musculoskeletal Diseases (ESCEO). *Aging Clin Exp Res* 2019; 31: 905-915. doi: 10.1007/s40520-019-01193-8. PMID: 30993659; PMCID: PMC6589151.
20. Beard DJ, Davies LJ, Cook JA, MacLennan G, Price A, Kent S, Hudson J, Carr A, Leal J, Campbell H, Fitzpatrick R, Arden N, Murray D, Campbell MK; TOPKAT Study Group. The clinical and cost-effectiveness of total versus partial knee replacement in patients with medial compartment osteoarthritis (TOPKAT): 5-year outcomes of a randomised controlled trial. *Lancet* 2019; 394: 746-756. doi: 10.1016/S0140-6736(19)31281-4. PMID: 31326135; PMCID: PMC6727069.
21. Geyer M, Schönfeld C. Novel Insights into the pathogenesis of osteoarthritis. *Curr Rheumatol Rev* 2018; 14: 98-107. doi: 10.2174/1573397113666170807122312. PMID: 28782470.
22. Cisewski SE, Zhang L, Kuo J, Wright GJ, Wu Y, Kern MJ, Yao H. The effects of oxygen level and glucose concentration on the metabolism of porcine TMJ disc cells. *Osteoarthritis Cartilage* 2015; 23: 1790-1796. doi: 10.1016/j.joca.2015.05.021. PMID: 26033165; PMCID: PMC4577453.
23. Henrotin Y, Kurz B, Aigner T. Oxygen and reactive oxygen species in cartilage degradation: friends or foes? *Osteoarthritis Cartilage* 2005; 13: 643-654. doi: 10.1016/j.joca.2005.04.002. PMID: 15936958.
24. Guilak F, Nims RJ, Dicks A, Wu CL, Meulenbelt I. Osteoarthritis as a disease of the cartilage pericellular matrix. *Matrix Biol* 2018; 71: 40-50. doi: 10.1016/j.matbio.2018.05.008. PMID: 29800616; PMCID: PMC6146061.
25. Plaas A, Osborn B, Yoshihara Y, Bai Y, Bloom T, Nelson F, Mikecz K, Sandy JD. Aggrecanolytic in human osteoarthritis: confocal localization and biochemical characterization of ADAMTS5-hyaluronan complexes in articular cartilages. *Osteoarthritis Cartilage* 2007; 15: 719-734. doi: 10.1016/j.joca.2006.12.008. PMID: 17360199.
26. Hwang HS, Kim HA. Chondrocyte apoptosis in the pathogenesis of osteoarthritis. *Int J Mol Sci* 2015; 16: 26035-26054. doi: 10.3390/ijms161125943. PMID: 26528972; PMCID: PMC4661802.
27. Griffin TM, Scanzello CR. Innate inflammation and synovial macrophages in osteoarthritis pathophysiology. *Clin Exp Rheumatol* 2019; 37: 57-63. PMID: 31621560; PMCID: PMC6842324.
28. de Lange-Brokaar BJ, Ioan-Facsinay A, van Osch GJ, Zuurmond AM, Schoones J, Toes RE, Huizinga TW, Kloppenburg M. Synovial inflammation, immune cells and their cytokines in osteoarthritis: a review. *Osteoarthritis Cartilage* 2012; 20: 1484-1499. doi: 10.1016/j.joca.2012.08.027. PMID: 22960092.
29. Woodell-May JE, Sommerfeld SD. Role of inflammation and the immune system in the progression of osteoarthritis. *J Orthop Res* 2020; 38: 253-257. doi: 10.1002/jor.24457. PMID: 31469192.
30. Wellsandt E, Golightly Y. Exercise in the management of knee and hip osteoarthritis. *Curr Opin Rheumatol*. 2018; 30: 151-159. doi: 10.1097/BOR.0000000000000478. PMID: 29251659.
31. Hussain SM, Neilly DW, Baliga S, Patil S, Meek R. Knee osteoarthritis: a review of management options. *Scott Med J* 2016; 61: 7-16. doi: 10.1177/0036933015619588. Epub 2016 Jun 21. PMID: 27330013.
32. Conaghan PG, Arden N, Avouac B, Migliore A, Rizzoli R. Safety of paracetamol in osteoarthritis: what does the literature say? *Drugs Aging* 2019; 36: 7-14. doi: 10.1007/s40266-019-00658-9. PMID: 31073920; PMCID: PMC6509082.
33. Rannou F, Pelletier JP, Martel-Pelletier J. Efficacy and safety of topical NSAIDs in the management of osteoarthritis: Evidence from real-life setting trials and surveys. *Semin Arthritis Rheum* 2016; 45: S18-21. doi: 10.1016/j.semarthrit.2015.11.007. PMID: 26806189.
34. Jüni P, Hari R, Rutjes AW, Fischer R, Silleta MG, Reichenbach S, da Costa BR. Intra-articular corticosteroid for knee osteoarthritis. *Cochrane Database Syst Rev* 2015; (10): CD005328. doi: 10.1002/14651858.CD005328.pub3. PMID: 26490760.
35. Falkowski K, Skiba G, Czerner M, Szmajda M, Bączkiewicz D. Effects of Viscosupplementation on Knee Joint Arthrokinematics - Pilot Study. *Ortop Traumatol Rehabil* 2018; 20: 409-419. doi: 10.5604/01.3001.0012.8277. PMID: 30648664.
36. Letizia Mauro G, Scaturro D, Sanfilippo A, Benedetti MG. Intra-articular hyaluronic acid injections for hip osteoarthritis. *J Biol Regul Homeost Agents*. 2018; 32: 1303-1309. PMID: 30334430.
37. Harrison-Muñoz S, Rojas-Briones V, Irarrázaval S. Is glucosamine effective for osteoarthritis? *Medwave* 2017; 17: e6867. doi: 10.5867/medwave.2017.6867. PMID: 28306711.
38. Bishnoi M, Jain A, Hurkat P, Jain SK. Chondroitin sulphate: a focus on osteoarthritis. *Glycoconj J* 2016; 33: 693-705. doi: 10.1007/s10719-016-9665-3. PMID: 27194526.
39. Thorlund JB, Juhl CB, Roos EM, Lohmander LS. Arthroscopic surgery for degenerative knee: systematic review and meta-analysis of benefits and harms. *BMJ* 2015; 350: h2747. doi: 10.1136/bmj.h2747. PMID: 26080045; PMCID: PMC4469973.
40. Li J, Li YX, Luo LJ, Ye J, Zhong DL, Xiao QW, Zheng H, Geng CM, Jin RJ, Liang FR. The effectiveness and safety of acupuncture for knee osteoarthritis: an overview of systematic reviews. *Medicine (Baltimore)* 2019; 98: e16301. doi: 10.1097/MD.0000000000016301. PMID: 31305415; PMCID: PMC6641846.

41. Manyanga T, Froese M, Zarychanski R, Abou-Setta A, Friesen C, Tennenhouse M, Shay BL. Pain management with acupuncture in osteoarthritis: a systematic review and meta-analysis. *BMC Complement Altern Med* 2014; 14: 312. doi: 10.1186/1472-6882-14-312. PMID: 25151529; PMCID: PMC4158087.
42. Mistry H, Connock M, Pink J, Shyangdan D, Clar C, Royle P, Court R, Biant LC, Metcalfe A, Waugh N. Autologous chondrocyte implantation in the knee: systematic review and economic evaluation. *Health Technol Assess* 2017; 21: 1-294. doi: 10.3310/hta21060. PMID: 28244303; PMCID: PMC5346885.
43. Zellner J, Krutsch W, Pfeifer C, Koch M, Nerlich M, Angele P. Autologous chondrocyte implantation for cartilage repair: current perspectives. *Orthop Res Rev*. 2015; 7: 149-158 doi: 10.2147/ORR.S65569
44. Ding DC, Shyu WC, Lin SZ. Mesenchymal stem cells. *Cell Transplant* 2011; 20: 5-14. doi: 10.3727/096368910X. PMID: 21396235.
45. Galipeau J, Sensébé L. Mesenchymal Stromal Cells: Clinical Challenges and Therapeutic Opportunities. *Cell Stem Cell* 2018; 22: 824-833. doi: 10.1016/j.stem.2018.05.004. PMID: 29859173; PMCID: PMC6434696.
46. Friedenstein AJ. Precursor cells of mechanocytes. *Int Rev Cytol* 1976; 47: 327-359. doi: 10.1016/s0074-7696(08)60092-3. PMID: 11195.
47. Squillaro T, Peluso G, Galderisi U. Clinical trials with mesenchymal stem cells: an update. *Cell Transplant* 2016; 25: 829-848. doi: 10.3727/096368915X689622. PMID: 26423725.
48. Dominici M, Le Blanc K, Mueller I, Slaper-Cortenbach I, Marini F, Krause D, Deans R, Keating A, Prockop Dj, Horwitz E. Minimal criteria for defining multipotent mesenchymal stromal cells. The International Society for Cellular Therapy position statement. *Cytotherapy* 2006; 8: 315-317. doi: 10.1080/14653240600855905. PMID: 16923606.
49. Charbord P. Bone marrow mesenchymal stem cells: historical overview and concepts. *Hum Gene Ther* 2010; 21: 1045-1056. doi: 10.1089/hum.2010.115. PMID: 20565251; PMCID: PMC4823383.
50. Huang L, Zhang C, Gu J, Wu W, Shen Z, Zhou X, Lu H. A randomized, placebo-controlled trial of human umbilical cord blood mesenchymal stem cell infusion for children with cerebral palsy. *Cell Transplant* 2018; 27: 325-334. doi: 10.1177/0963689717729379. PMID: 29637820; PMCID: PMC5898688.
51. Ding DC, Chang YH, Shyu WC, Lin SZ. Human umbilical cord mesenchymal stem cells: a new era for stem cell therapy. *Cell Transplant* 2015; 24: 339-347. doi: 10.3727/096368915X686841. PMID: 25622293.
52. Antoniadou E, David AL. Placental stem cells. *Best Pract Res Clin Obstet Gynaecol* 2016; 31: 13-29. doi: 10.1016/j.bpobgyn.2015.08.014. PMID: 26547389.
53. Farhadhosseinabadi B, Farahani M, Tayebi T, Jafari A, Biniazan F, Modaresifar K, Moravvej H, Bahrami S, Redl H, Tayebi L, Nikejrad H. Amniotic membrane and its epithelial and mesenchymal stem cells as an appropriate source for skin tissue engineering and regenerative medicine. *Artif Cells Nanomed Biotechnol* 2018; 46: 431-440. doi: 10.1080/21691401.2018.1458730. PMID: 29687742.
54. Harrell CR, Gazdic M, Fellabaum C, Jovicic N, Djonov V, Arsenijevic N, Volarevic V. Therapeutic Potential of Amniotic Fluid Derived Mesenchymal Stem Cells Based on their Differentiation Capacity and Immunomodulatory Properties. *Curr Stem Cell Res Ther* 2019; 14: 327-336. doi: 10.2174/1574888X14666190222201749. PMID: 30806325.
55. Wang YL, Hong A, Yen TH, Hong HH. Isolation of mesenchymal stem cells from human alveolar periosteum and effects of vitamin d on osteogenic activity of periosteum-derived cells. *J Vis Exp* 2018; 135: 57166. doi: 10.3791/57166. PMID: 29782010; PMCID: PMC6101109.
56. Sakaguchi Y, Sekiya I, Yagishita K, Ichinose S, Shinomiya K, Muneta T. Suspended cells from trabecular bone by collagenase digestion become virtually identical to mesenchymal stem cells obtained from marrow aspirates. *Blood* 2004; 104: 2728-2735. doi: 10.1182/blood-2003-12-4452. PMID: 15242873.
57. Minter D, Marra KG, Rubin JP. Adipose-derived mesenchymal stem cells: biology and potential applications. *Adv Biochem Eng Biotechnol* 2013; 129: 59-71. doi: 10.1007/10_2012_146. PMID: 22825719.
58. Zupan J, Drobnič M, Stražar K. Synovium-Derived Mesenchymal Stem/Stromal Cells and their Promise for Cartilage Regeneration. *Adv Exp Med Biol* 2020; 1212: 87-106. doi: 10.1007/5584_2019_381. PMID: 31069722.
59. Čamernik K, Marc J, Zupan J. Human skeletal muscle-derived mesenchymal stem/stromal cell isolation and growth kinetics analysis. *Methods Mol Biol* 2019; 2045: 119-129. doi: 10.1007/7651_2018_201. PMID: 30499023.
60. Hollands P, Aboyeji D, Orcharton M. Dental pulp stem cells in regenerative medicine. *Br Dent J* 2018; doi: 10.1038/sj.bdj.2018.348. PMID: 29725075.
61. Heo JS, Choi Y, Kim HS, Kim HO. Comparison of molecular profiles of human mesenchymal stem cells derived from bone marrow, umbilical cord blood, placenta and adipose tissue. *Int J Mol Med* 2016; 37: 115-125. doi: 10.3892/ijmm.2015.2413. PMID: 26719857; PMCID: PMC4687432.
62. Jiang W, Xu J. Immune modulation by mesenchymal stem cells. *Cell Prolif* 2020; 53: e12712. doi: 10.1111/cpr.12712. PMID: 31730279; PMCID: PMC6985662.
63. Luque-Campos N, Contreras-López RA, Jose Paredes-Martínez M, Torres MJ, Bahraoui S, Wei M, Espinoza F, Djouad F, Elizondo-Vega RJ, Luz-Crawford P. Mesenchymal stem cells improve rheumatoid arthritis progression by controlling memory t cell response. *Front Immunol* 2019; 10: 798. doi: 10.3389/fimmu.2019.00798. PMID: 31040848; PMCID: PMC6477064.
64. Fan L, Hu C, Chen J, Cen P, Wang J, Li L. Interaction between Mesenchymal Stem Cells and B-Cells. *Int J Mol Sci* 2016; 17: 650. doi: 10.3390/ijms17050650. PMID: 27164080; PMCID: PMC4881476.
65. Regmi S, Pathak S, Kim JO, Yong CS, Jeong JH. Mesenchymal stem cell therapy for the treatment of inflammatory diseases: Challenges, opportunities, and future perspectives. *Eur J Cell Biol* 2019; 98: 151041. doi: 10.1016/j.ejcb.2019.04.002. PMID: 31023504.

66. Najar M, Fayyad-Kazan M, Merimi M, Burny A, Bron D, Fayyad-Kazan H, Meuleman N, Lagneaux L. Mesenchymal stromal cells and natural killer cells: a complex story of love and hate. *Curr Stem Cell Res Ther* 2019; 14: 14-21. doi: 10.2174/1574888X13666180912125736. PMID: 30207245.
67. Gneccchi M, Danieli P, Malpasso G, Ciuffreda MC. Paracrine mechanisms of mesenchymal stem cells in tissue repair. *Methods Mol Biol* 2016; 1416: 123-146. doi: 10.1007/978-1-4939-3584-0_7. PMID: 27236669.
68. He JG, Li BB, Zhou L, Yan D, Xie QL, Zhao W. Indoleamine 2,3-dioxygenase-transfected mesenchymal stem cells suppress heart allograft rejection by increasing the production and activity of dendritic cells and regulatory T cells. *J Investig Med* 2020; 68: 728-737. doi: 10.1136/jim-2019-001160. PMID: 31892638.
69. Zhang Z, Huang S, Wu S, Qi J, Li W, Liu S, Cong Y, Chen H, Lu L, Shi S, Wang D, Chen W, Sun L. Clearance of apoptotic cells by mesenchymal stem cells contributes to immunosuppression via PGE2. *EBioMedicine* 2019; 45: 341-350. doi: 10.1016/j.ebiom.2019.06.016. PMID: 31248835; PMCID: PMC6642220.
70. Ueno M, Lo CW, Barati D, Conrad B, Lin T, Kohno Y, Utsunomiya T, Zhang N, Maruyama M, Rhee C, Huang E, Romero-Lopez M, Tong X, Yao Z, Zwingenberger S, Yang F, Goodman SB. Interleukin-4 overexpressing mesenchymal stem cells within gelatin-based microribbon hydrogels enhance bone healing in a murine long bone critical-size defect model. *J Biomed Mater Res A* 2020; 108: 2240-2250. doi: 10.1002/jbm.a.36982. PMID: 32363683; PMCID: PMC7483618.
71. Shao M, Wang D, Zhou Y, Du K, Liu W. Interleukin-10 delivered by mesenchymal stem cells attenuates experimental autoimmune myocarditis. *Int Immunopharmacol* 2020; 81: 106212. doi: 10.1016/j.intimp.2020.106212. PMID: 32062070.
72. Zhao Y, Teng B, Sun X, Dong Y, Wang S, Hu Y, Wang Z, Ma X, Yang Q. Synergistic effects of kartogenin and transforming growth factor- β 3 on chondrogenesis of human umbilical cord mesenchymal stem cells in vitro. *Orthop Surg* 2020; 12: 938-945. doi: 10.1111/os.12691. PMID: 32462800; PMCID: PMC7307229.
73. Han Y, Li X, Zhang Y, Han Y, Chang F, Ding J. Mesenchymal stem cells for regenerative medicine. *Cells* 2019; 8: 886. doi: 10.3390/cells8080886. PMID: 31412678; PMCID: PMC6721852.
74. Nazempour A, Van Wie BJ. Chondrocytes, mesenchymal stem cells, and their combination in articular cartilage regenerative medicine. *Ann Biomed Eng* 2016; 44: 1325-1354. doi: 10.1007/s10439-016-1575-9. PMID: 26987846.
75. Shariatzadeh M, Song J, Wilson SL. The efficacy of different sources of mesenchymal stem cells for the treatment of knee osteoarthritis. *Cell Tissue Res* 2019; 378: 399-410. doi: 10.1007/s00441-019-03069-9. PMID: 31309317.
76. Emadedin M, Labibzadeh N, Liastani MG, Karimi A, Jaroughi N, Bolurieh T, Hosseini SE, Baharvand H, Aghdami N. Intra-articular implantation of autologous bone marrow-derived mesenchymal stromal cells to treat knee osteoarthritis: a randomized, triple-blind, placebo-controlled phase 1/2 clinical trial. *Cytotherapy* 2018; 20: 1238-1246. doi: 10.1016/j.jcyt.2018.08.005. PMID: 30318332.
77. Vega A, Martín-Ferrero MA, Del Canto F, Alberca M, García V, Munar A, Orozco L, Soler R, Fuertes JJ, Huguet M, Sánchez A, García-Sancho J. Treatment of knee osteoarthritis with allogeneic bone marrow mesenchymal stem cells: a randomized controlled trial. *Transplantation* 2015; 99: 1681-1690. doi: 10.1097/TP.0000000000000678. PMID: 25822648.
78. De Bari C, Roelofs AJ. Stem cell-based therapeutic strategies for cartilage defects and osteoarthritis. *Curr Opin Pharmacol* 2018; 40: 74-80. doi: 10.1016/j.coph.2018.03.009. PMID: 29625333.
79. Mushahary D, Spittler A, Kasper C, Weber V, Charwat V. Isolation, cultivation, and characterization of human mesenchymal stem cells. *Cytometry A* 2018; 93: 19-31. doi: 10.1002/cyto.a.23242. PMID: 29072818.
80. Hinckel BB, Gomoll AH. Autologous chondrocytes and next-generation matrix-based autologous chondrocyte implantation. *Clin Sports Med* 2017; 36: 525-548. doi: 10.1016/j.csm.2017.02.008. PMID: 28577711.
81. Chen Y, Ouyang X, Wu Y, Guo S, Xie Y, Wang G. Co-culture and mechanical stimulation on mesenchymal stem cells and chondrocytes for cartilage tissue engineering. *Curr Stem Cell Res Ther* 2020; 15: 54-60. doi: 10.2174/1574888X14666191029104249. PMID: 31660820.
82. Hodax JK, Quintos JB, Gruppuso PA, Chen Q, Desai S, Jayasuriya CT. Aggrecan is required for chondrocyte differentiation in ATDC5 chondroprogenitor cells. *PLoS One* 2019; 14: e0218399. doi: 10.1371/journal.pone.0218399. PMID: 31206541; PMCID: PMC6576788.
83. Yoon HJ, Kim SB, Somaiya D, Noh MJ, Choi KB, Lim CL, Lee HY, Lee YJ, Yi Y, Lee KH. Type II collagen and glycosaminoglycan expression induction in primary human chondrocyte by TGF- β 1. *BMC Musculoskelet Disord* 2015; 16: 141. doi: 10.1186/s12891-015-0599-x. PMID: 26059549; PMCID: PMC4460646.
84. Colombini A, Perucca Orfei C, Kouroupis D, Ragni E, De Luca P, Viganò M, Correa D, de Girolamo L. Mesenchymal stem cells in the treatment of articular cartilage degeneration: New biological insights for an old-timer cell. *Cytotherapy* 2019; 21: 1179-1197. doi: 10.1016/j.jcyt.2019.10.004. PMID: 31784241.
85. Beyer Nardi N, da Silva Meirelles L. Mesenchymal stem cells: isolation, in vitro expansion and characterization. *Handb Exp Pharmacol* 2006; 174: 249-282. PMID: 16370331.
86. Nejadnik H, Hui JH, Feng Choong EP, Tai BC, Lee EH. Autologous bone marrow-derived mesenchymal stem cells versus autologous chondrocyte implantation: an observational cohort study. *Am J Sports Med* 2010; 38: 1110-1116. doi: 10.1177/0363546509359067. PMID: 20392971.
87. Chahal J, Gómez-Aristizábal A, Shestopaloff K, Bhatt S, Chaboureaud A, Fazio A, Chisholm J, Weston A, Chiovitti J, Keating A, Kapoor M, Ogilvie-Harris DJ, Syed KA, Gandhi R, Mahomed NN, Marshall KW, Sussman MS, Naraghi AM, Viswanathan S. Bone marrow mesenchymal stromal cell treatment in patients with osteoarthritis results in overall improvement in pain and symptoms and reduces synovial inflammation. *Stem Cells Transl Med* 2019; 8: 746-757. doi: 10.1002/sctm.18-0183. PMID: 30964245; PMCID: PMC6646697.
88. Chen P, Huang L, Ma Y, Zhang D, Zhang X, Zhou J, Ruan A, Wang Q. Intra-articular platelet-rich plasma injection

- for knee osteoarthritis: a summary of meta-analyses. *J Orthop Surg Res* 2019; 14: 385. doi: 10.1186/s13018-019-1363-y. PMID: 31775816; PMCID: PMC6880602.
89. Kim SH, Ha CW, Park YB, Nam E, Lee JE, Lee HJ. Intra-articular injection of mesenchymal stem cells for clinical outcomes and cartilage repair in osteoarthritis of the knee: a meta-analysis of randomized controlled trials. *Arch Orthop Trauma Surg* 2019; 139: 971-980. doi: 10.1007/s00402-019-03140-8. PMID: 30756165.
90. Naji A, Eitoku M, Favier B, Deschaseaux F, Rouas-Freiss N, Suganuma N. Biological functions of mesenchymal stem cells and clinical implications. *Cell Mol Life Sci* 2019; 76: 3323-3348. doi: 10.1007/s00018-019-03125-1. PMID: 31055643.
91. Birmingham E, Niebur GL, McHugh PE, Shaw G, Barry FP, McNamara LM. Osteogenic differentiation of mesenchymal stem cells is regulated by osteocyte and osteoblast cells in a simplified bone niche. *Eur Cell Mater* 2012; 23: 13-27. doi: 10.22203/ecm.v023a02. PMID: 22241610.
92. Murphy MB, Moncivais K, Caplan AI. Mesenchymal stem cells: environmentally responsive therapeutics for regenerative medicine. *Exp Mol Med* 2013; 45: e54. doi: 10.1038/emm.2013.94. PMID: 24232253; PMCID: PMC3849579.
93. Vizoso FJ, Eiro N, Cid S, Schneider J, Perez-Fernandez R. Mesenchymal stem cell secretome: toward cell-free therapeutic strategies in regenerative medicine. *Int J Mol Sci* 2017; 18: 1852. doi: 10.3390/ijms18091852. PMID: 28841158; PMCID: PMC5618501.
94. Gonzalez-Pujana A, Igartua M, Santos-Vizcaino E, Hernandez RM. Mesenchymal stromal cell based therapies for the treatment of immune disorders: recent milestones and future challenges. *Expert Opin Drug Deliv* 2020; 17: 189-200. doi: 10.1080/17425247.2020.1714587. PMID: 31918562.
95. Um S, Kim HY, Lee JH, Song IS, Seo BM. TSG-6 secreted by mesenchymal stem cells suppresses immune reactions influenced by BMP-2 through p38 and MEK mitogen-activated protein kinase pathway. *Cell Tissue Res* 2017; 368: 551-561. doi: 10.1007/s00441-017-2581-4. PMID: 28247086.
96. Wang R, Xu B, Xu H. TGF- β 1 promoted chondrocyte proliferation by regulating Sp1 through MSC-exosomes derived miR-135b. *Cell Cycle* 2018; 17: 2756-2765. doi: 10.1080/15384101.2018.1556063. PMID: 30526325; PMCID: PMC6343719.
97. Jiang H, Hong T, Wang T, Wang X, Cao L, Xu X, Zheng M. Gene expression profiling of human bone marrow mesenchymal stem cells during osteogenic differentiation. *J Cell Physiol* 2019; 234: 7070-7077. doi: 10.1002/jcp.27461. PMID: 30378112.
98. Gupta PK, Das AK, Chullikana A, Majumdar AS. Mesenchymal stem cells for cartilage repair in osteoarthritis. *Stem Cell Res Ther* 2012; 3: 25. doi: 10.1186/scrt116. PMID: 22776206; PMCID: PMC3580463.
99. Dwivedi G, Chevrier A, Alameh MG, Hoemann CD, Buschmann MD. Quality of cartilage repair from marrow stimulation correlates with cell number, clonogenic, chondrogenic, and matrix production potential of underlying bone marrow stromal cells in a rabbit model. *Cartilage* 2021; 12: 237-250. doi: 10.1177/1947603518812555. PMID: 30569762; PMCID: PMC7970370.
100. Fan L, Chen J, Tao Y, Heng BC, Yu J, Yang Z, Ge Z. Enhancement of the chondrogenic differentiation of mesenchymal stem cells and cartilage repair by ghrelin. *J Orthop Res* 2019; 37: 1387-1397. doi: 10.1002/jor.24224. PMID: 30644571.
101. Gelse K, Mühle C, Knaup K, Swoboda B, Wiesener M, Hennig F, Olk A, Schneider H. Chondrogenic differentiation of growth factor-stimulated precursor cells in cartilage repair tissue is associated with increased HIF-1 α activity. *Osteoarthritis Cartilage* 2008; 16: 1457-1465. doi: 10.1016/j.joca.2008.04.006. PMID: 18524637.
102. Frölich K, Scherzed A, Mlynski R, Technau A, Hagen R, Kleinsasser N, Radeloff A. Multipotent stromal cells for autologous cell therapy approaches in the guinea pig model. *ORL J Otorhinolaryngol Relat Spec* 2011; 73: 9-16. doi: 10.1159/000320598. PMID: 20975314.
103. Xie J, Ren J, Liu N, Wu C, Xiao D, Luo H, Du J. Pretreatment with AM1241 enhances the analgesic effect of intrathecally administered mesenchymal stem cells. *Stem Cells Int* 2019; 2019: 7025473. doi: 10.1155/2019/7025473. PMID: 31611918; PMCID: PMC6755285.
104. Rodas G, Soler-Rich R, Rius-Tarruella J, Alomar X, Balias R, Orozco L, Masci L, Maffulli N. Effect of autologous expanded bone marrow mesenchymal stem cells or leukocyte-poor platelet-rich plasma in chronic patellar tendinopathy (With Gap >3 mm): preliminary outcomes after 6 months of a double-blind, randomized, prospective study. *Am J Sports Med* 2021; 49: 1492-1504. doi: 10.1177/0363546521998725. PMID: 33783227.
105. Richardson SM, Kalamegam G, Pushparaj PN, Matta C, Memic A, Khademhosseini A, Mobasheri R, Poletti FL, Hoyland JA, Mobasheri A. Mesenchymal stem cells in regenerative medicine: focus on articular cartilage and intervertebral disc regeneration. *Methods* 2016; 99: 69-80. doi: 10.1016/j.jymeth.2015.09.015. PMID: 26384579.
106. Centeno C, Markle J, Dodson E, Stemper I, Williams CJ, Hyzy M, Ichim T, Freeman M. Treatment of lumbar degenerative disc disease-associated radicular pain with culture-expanded autologous mesenchymal stem cells: a pilot study on safety and efficacy. *J Transl Med* 2017; 15: 197. doi: 10.1186/s12967-017-1300-y. PMID: 28938891; PMCID: PMC5610473.
107. Elseberg CL, Salzig D, Czermak P. Bioreactor expansion of human mesenchymal stem cells according to GMP requirements. *Methods Mol Biol* 2015; 1283: 199-218. doi: 10.1007/7651_2014_117. PMID: 25540116.
108. Orozco L, Munar A, Soler R, Alberca M, Soler F, Huguet M, Sentís J, Sánchez A, García-Sancho J. Treatment of knee osteoarthritis with autologous mesenchymal stem cells: a pilot study. *Transplantation* 2013; 95: 1535-1541. doi: 10.1097/TP.0b013e318291a2da. PMID: 23680930.
109. Saris TFF, de Windt TS, Kester EC, Vonk LA, Custers RJH, Saris DBF. Five-year outcome of 1-stage cell-based cartilage repair using recycled autologous chondrons and allogeneic mesenchymal stromal cells: a first-in-human clinical trial. *Am J Sports Med* 2021; 49: 941-947. doi: 10.1177/0363546520988069. PMID: 33591794.

110. Veyrat-Masson R, Boiret-Dupré N, Rapatel C, Descamps S, Guillouard L, Guérin JJ, Pigeon P, Boisgard S, Chassagne J, Berger MG. Mesenchymal content of fresh bone marrow: a proposed quality control method for cell therapy. *Br J Haematol* 2007; 139: 312-320. doi: 10.1111/j.1365-2141.2007.06786.x. PMID: 17897309.
111. Wang N, Badar F, Xia Y. Resolution-dependent influences of compressed sensing in quantitative T2 mapping of articular cartilage. *NMR Biomed* 2020; 33: e4260. doi: 10.1002/nbm.4260. PMID: 32040226; PMCID: PMC7415577.
112. Wang X, Wrigley TV, Bennell KL, Wang Y, Fortin K, Cicuttini FM, Lloyd DG, Bryant AL. Cartilage quantitative T2 relaxation time 2-4 years following isolated anterior cruciate ligament reconstruction. *J Orthop Res* 2018; 36: 2022-2029. doi: 10.1002/jor.23846. PMID: 29280504.
113. David-Vaudey E, Ghosh S, Ries M, Majumdar S. T2 relaxation time measurements in osteoarthritis. *Magn Reson Imaging* 2004; 22: 673-682. doi: 10.1016/j.mri.2004.01.071. PMID: 15172061.
114. Peterfy CG, Guermazi A, Zaim S, Tirman PF, Miaux Y, White D, Kothari M, Lu Y, Fye K, Zhao S, Genant HK. Whole-Organ Magnetic Resonance Imaging Score (WORMS) of the knee in osteoarthritis. *Osteoarthritis Cartilage* 2004; 12: 177-190. doi: 10.1016/j.joca.2003.11.003. PMID: 14972335.
115. Orozco L, Munar A, Soler R, Alberca M, Soler F, Huguet M, Sentís J, Sánchez A, García-Sancho J. Treatment of knee osteoarthritis with autologous mesenchymal stem cells. *Trans J* 2013; 95: 1535-1541 doi: 10.1097/TP.0b013e318291a2da
116. Lawrence LM, Cottrill A, Valluri A, Marenzi G, Denning KL, Valluri J, Claudio PP, Day JB. Minimally manipulative method for the expansion of human bone marrow mesenchymal stem cells to treat osseous defects. *Int J Mol Sci* 2019; 20: 612. doi: 10.3390/ijms20030612. PMID: 30708975; PMCID: PMC6387189.
117. Hassan MNFB, Yazid MD, Yunus MHM, Chowdhury SR, Lokanathan Y, Idrus RBH, Ng AMH, Law JX. Large-scale expansion of human mesenchymal stem cells. *Stem Cells Int* 2020; 2020: 9529465. doi: 10.1155/2020/9529465. PMID: 32733574; PMCID: PMC7378617.
118. Cundell T, Drummond S, Ford I, Reber D, Singer D; Members of the Pharmaceutical Microbiology Expert Discussion Group. Risk Assessment Approach to Microbiological Controls of Cell Therapies. *PDA J Pharm Sci Technol* 2020; 74: 229-248. doi: 10.5731/pda-jpst.2019.010546. PMID: 31941793.
119. Ryan AE, Lohan P, O'Flynn L, Treacy O, Chen X, Coleman C, Shaw G, Murphy M, Barry F, Griffin MD, Ritter T. Chondrogenic differentiation increases antidonor immune response to allogeneic mesenchymal stem cell transplantation. *Mol Ther* 2014; 22: 655-667. doi: 10.1038/mt.2013.261. PMID: 24184966; PMCID: PMC3944342.
120. Le Blanc K, Ringdén O. Immunomodulation by mesenchymal stem cells and clinical experience. *J Intern Med* 2007; 262: 509-525. doi: 10.1111/j.1365-2796.2007.01844.x. PMID: 17949362.
121. Ankrum JA, Ong JF, Karp JM. Mesenchymal stem cells: immune evasive, not immune privileged. *Nat Biotechnol* 2014; 32: 252-260. doi: 10.1038/nbt.2816. PMID: 24561556; PMCID: PMC4320647.
122. Trebinjac S, Gharairi M. Mesenchymal stem cells for treatment of tendon and ligament injuries-clinical evidence. *Med Arch* 2020; 74: 387-390. doi: 10.5455/medarh.2020.74.387-390. PMID: 33424096; PMCID: PMC7780758.
123. Bekkers JE, Tsuchida AI, van Rijen MH, Vonk LA, Dhert WJ, Creemers LB, Saris DB. Single-stage cell-based cartilage regeneration using a combination of chondrons and mesenchymal stromal cells: comparison with microfracture. *Am J Sports Med* 2013; 41: 2158-2166. doi: 10.1177/0363546513494181. PMID: 23831891.
124. Vega A, Martín-Ferrero MA, Del Canto F, Alberca M, García V, Munar A, Orozco L, Soler R, Fuertes JJ, Huguet M, Sánchez A, García-Sancho J. Treatment of knee osteoarthritis with allogeneic bone marrow mesenchymal stem cells: a randomized controlled trial. *Transplantation* 2015; 99: 1681-1690. doi: 10.1097/TP.0000000000000678. PMID: 25822648.
125. Carvello M, Lightner A, Yamamoto T, Kotze PG, Spinelli A. Mesenchymal stem cells for perianal crohn's disease. *Cells* 2019; 8: 764. doi: 10.3390/cells8070764. PMID: 31340546; PMCID: PMC6679174.
126. Mansoor SR, Zabihi E, Ghasemi-Kasman M. The potential use of mesenchymal stem cells for the treatment of multiple sclerosis. *Life Sci* 2019; 235: 116830. doi: 10.1016/j.lfs.2019.116830. PMID: 31487529.