GLP-1 receptor agonists can reverse immunosuppressioninduced beta-cell dysfunction

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ABSTRACT

It is well known that some of the agents commonly used during immunosuppressive (IS) therapy after solid organ transplantation can contribute to beta cell dysfunction and result in diabetes mellitus in the transplant recipient. Some of the risks associated with post-transplant diabetes mellitus (PTDM) include cardiovascular disease (CVD), graft failure and mortality. Since this significance was recognized, many studies are ongoing to refine the IS therapy regimen to reduce or discontinue corticosteroids and calcineurin inhibitors (CNI).

A promising addition to the immunosuppression treatment regimen to treat PTDM is glucagon-like peptide-1 receptor (GLP-1R) agonists or incretin mimetics normally used in the treatment of type 2 diabetes (T2D). Studies show that adding GLP-1R agonists to the immunosuppression regimen after solid organ transplant is beneficial not only for the health of the islet beta cells but also positively affects immune function in metabolic disorders by suppressing the activation of CD4+ T lymphocyte cytokine expression. Additional benefits include decreased cardiac graft vasculopathy, improvement of hepatic steatosis, preservation of kidney function, enhanced graft survival and improved all-cause mortality rates for solid organ transplant recipients.

ABBREVIATIONS

AATG = rabbit antithymocyte globulins, Aza = azathioprine, CD26 = cluster of differentiation antigen 26, CNI = calcineurin inhibitors, Cn/NFAT =

calcineurin/Nuclear Factor of Activated T cells, CsA = cyclosporine A, DCG = dense core granule, CV = cardiovascular, CVD = cardiovascular disease, DPP-4 = dipeptidyl peptidase-4, DPP-4i = dipeptidyl peptidase-4 inhibitors, GCs = glucocorticoids, GLP-1R = Glucagon-like peptide-1 receptor, HbA1c = glycated hemoglobin, IS = Immunosuppression, MMF = mycophenolate mofetil, mTOR = mammalian target of rapamycin, PPAR- γ = peroxisome proliferator-activated receptor gamma, PTDM = post-transplant diabetes mellitus, SGLT2 = sodium-glucose cotransporter 2, SOT = solid organ transplant, T2D = type 2 diabetes, Tac = tacrolimus, TGF-β1= transforming growth factor- β 1, TZD = thiazolidinediones.

Introduction

With all the success achieved in solid organ transplants (SOT) there is nevertheless room for improvement. Immunosuppression (IS) is required to keep the immune system of the recipient from rejecting the allogeneic tissue of the donor. However, because of the complexity of the human immune system and our efforts to quell it, the adverse effects of longterm IS are still being discovered and addressed. Historically, immunosuppression was limited to inhibiting T-cells in the recipient first with whole body irradiation then with steroids, neither of which succeeded at improving graft survival. Then with the development of tissue matching of the donor to the recipient in the early 1960s, and a combination of drugs like azathioprine and steroids, graft survival improved, and SOTs became more widespread. When cyclosporine came into use in the 1980s, 1-year graft survival rates improved from 50% to 80%¹. Novel agents, including antibodies and fusion proteins against specific targets of the immune

system have been introduced over time and today, most IS cocktails contain several of the following: rabbit antithymocyte globulins (ATG), basiliximab, alemtuzumab, corticosteroids, tacrolimus (Tac), cyclosporine A (CsA), azathioprine (Aza), mycophenolate mofetil (MMF), sirolimus (rapamycin), everolimus, belatacept, intravenous immunoglobulin and rituximab (Table 1)¹⁻³. However, despite the success

of these drugs in preventing organ rejection, some of them have been linked to adverse events that include post-transplant diabetes mellitus (PTDM).

IMMUNOSUPPRESSION AND POST-TRANSPLANT DIABETES MELLITUS

IS needs to be administered for the life time of the organ recipient. However, long-term treatment with

Table 1. Immunosuppressive agents and their association with PTDM.

IS Agent	Activity of drug	PTDM Risk	Glycemic Effect	Notes	References
Antithymocyte globulins (ATG)	T-cell inhibitor	Low	No impact	Induction IS	Jasiak et al ¹ Black et al ² Shivaswamy et al ³ Munshi et al ⁷
Basiliximab	IL-2 receptor antagonist	Increased	Impaired glucose homeostasis	Induction IS	Jasiak et al ¹ Shivaswamy et al ³ Munshi et al ⁷
Alemtuzumab (Campath-1H)	Humanized anti-CD52 mono- conal antibody for lymphocyte depletion	Low	No impact	Induction IS	Jasiak et al ¹ Black et al ²
Corticosteroids	Steroid	High	 Increased hepatic gluconeogenesis Peripheral insulin resistance Dyslipidemia 	Induction IS, Maintenance IS	Jasiak et al ¹ Black et al ² Munshi et al ⁷ Ducloux et al ¹² Chowdhury et al ¹³ Shapiro et al ¹⁴ McMahon et al ¹⁵ Penfornis et al ¹⁶ Ahmed et al ²⁹
Tacrolimus/ FK506 (Tac)	Calcineurin inhibitor	High	 Beta cell apoptosis Peripheral insulin resistance Decreased insulin secretion 	Maintenance IS	Jasiak et al ¹ Black et al ² Chowdhury et al ¹³ Ducloux et al ¹² Chowdhury et al ¹³ Shapiro et al ¹⁴ Penfornis et al ¹⁶ Ahmed et al ²⁹ Cehic et al ⁵⁷
Cyclosporine A (CsA)	Calcineurin inhibitor	High	HyperglycemiaInsulin resistanceDecreased insulin secretion	Maintenance IS	Jasiak et al ¹ Black et al ² Cehic et al ⁵⁷
Azathioprine (Aza)	Prevents T-cell proliferation by inhibiting purine synthesis	Low	No impact	Maintenance IS	Jasiak et al ¹ Shivaswamy et al ³ Cehic et al ⁵⁷
Mycophenolate mofetil (MMF)	Prevents T-cell proliferation by depleting guanosine nucleotides	Low	No impact	Maintenance IS	Jasiak et al ¹ Shivaswamy et al ³ Munshi et al ⁷ Cehic et al ⁵⁷ Barlow et al ⁷³

Table 1 (continued). Immunosuppressive agents and their association with PTDM.

IS Agent	Activity of drug	PTDM Risk	Glycemic Effect	Notes	References
Sirolimus (rapamycin)	mTOR inhibitor	Increased	 Increased beta cell apoptosis Reduced beta cell proliferation Dyslipidemia Peripheral insulin resistance Dose-dependent hyperglycemia 	Maintenance IS	Jasiak et al ¹ Shivaswamy et al ³ Cehic et al ⁵⁷ Barlow et al ⁷³
Everolimus	mTOR inhibitor	Increased	Insulin resistanceDecreased insulin secretion	Maintenance IS	Jasiak et al ¹ Shivaswamy et al ³ Cehic et al ⁵⁷
Belatacept	Targets CD80/86 blocking T-cell costimulation pathway	Low	Lowered HbA1c	Maintenance IS but rejection events occurred when used without CNIs or steroids	Jasiak et al ¹ Jenssen et al ⁵ Terrec et al ²⁰
Rituximab	Targets CD20 antigen on B cells	Low	Slight chance of hyperglycemia due to a decrease in insulin secretion	Off-label use for kidney transplants but showed increased rate of infections including CMV	Jasiak et al ¹
Intravenous immunoglobulin (IVIG)	Role in immuno- modulation is complex and not clearly understood	None	No impact	Reduces T-cell activation, induces B cell apoptosis and inhibits complement	Jasiak et al ¹
Daclizumab	Humanized mab, binds to CD25, the alpha subunit of the IL-2 receptor on T cells	Low	May cause hyperglycemiaMay cause liverMay cause liver complications	Induction IS, withdrawn from global market global market due to incidents of encephalitis	Jasiak et al ¹ Black et al ² Penfornis et al ¹⁶ Penfornis et al ¹⁶ Bianchi et al ⁷⁴

Abbreviations: CMV = cytomegalovirus, HbA1c = glycated hemoglobin, IL-2 = interleukin 2, IS = immunosuppression, mab = monoclonal antibody, mTOR = mammalian target of rapamycin, PTDM = post-transplant diabetes mellitus.

some of the most common agents included in IS cocktails are associated with unfavorable consequences including PTDM. The incidence of PTDM is difficult to determine, with reports ranging from 2%-74%³⁻⁷ and PTDM cannot be underestimated as a cause for increased risk of cardiovascular morbidity and all-cause mortality in transplant patients⁸⁻¹⁰.

Most significant culprits

As far back as 1964, Starzl et al¹¹ noted new onset diabetes in patients after kidney transplantation. Some immunosuppressive agents are more

prone to cause impaired glucose tolerance and post-transplant diabetes than others. Arguably, at the top of the list are corticosteroids and calcineurin inhibitors such as tacrolimus/FK506 (Tac) and cyclosporine A (CsA)¹²⁻¹⁴. Corticosteroids increase both hepatic glucose production through gluconeogenesis stimulation and peripheral insulin resistance¹⁵. As for calcineurin inhibitors, there is a direct correlation between the effect on insulin secretion and dose administered¹⁶. It has been documented that diabetes caused by drugs like Tac and CsA can be partial-

ly abrogated or completely reversed by lowering the dose or switching to a different immuno-suppressant¹⁷⁻²⁰. Both sirolimus and everolimus function as inhibitors of the mammalian target of rapamycin (mTOR) and improvement of glucose intolerance and dyslipidemia have been reported when the dose of mTOR inhibitors are reduced³. Sirolimus, tacrolimus and everolimus have been shown to reduce beta cell mass through apoptosis and all have an anti-proliferative effect on beta cells³. Other IS agents such as MMF, belatacept and Aza have not been shown to have an impact on glucose metabolism or insulin secretion (Table 1).

HOW IMMUNOSUPPRESSION AFFECTS GLUCOSE METABOLISM AND INSULIN SECRETION

Normal beta cell development from birth encompasses two important functions. The first is responsiveness to glucose. This incorporates the ability to increase insulin production after sensing an increase in blood glucose levels. Mature beta cells must have the ability to generate and store insulin in secretory granules within the cell in preparation for release when needed to metabolize glucose. The second function of central importance is the ability of beta cells to expand to an appropriate mass as the organism grows to adulthood. Glucokinase is a crucial regulator of beta cell maturation and proliferation. It has been shown to ultimately stimulate pathways that lead to insulin production, secretion and proliferation²¹⁻²⁵. The calcineurin/Nuclear Factor of Activated T cells (Cn/NFAT) pathway that manages the proliferation, survival and differentiation of many cell types including lymphocytes and neurons has also been implicated in the regulation of beta cell function²⁶. Based on the observation that 10%-30% of transplant patients receiving calcineurin inhibitors such as tacrolimus developed diabetes mellitus, Goodyer et al²⁷ generated a genetically modified mouse to test the role of Cn/NFAT. Their results indicate that Cn/NFAT signaling is required during beta cell maturation for expression of insulin, Pdx1, Glut2 and glucokinase genes in mice and that Cn/NFAT signaling regulates expression of beta cell dense core granule (DCG) formation. Experiments exposing juvenile human islets to tacrolimus showed a similar reduction of gene expression encoding DCG components, and, therefore, Cn/NFAT signaling²⁷.

The means by which glucocorticoids (GCs) affect glucose homeostasis are complex and not well understood; however, GCs exert the majority of their effects on glucose metabolism through activation of glucocorticoid receptors. GCs are naturally produced by the adrenal cortex in response to stress. Long-term excess of endogenous or exogenously administered GCs impacts all aspects of glucose metabolism including hepatic gluconeogenesis and impaired insulin sensitivity at the level of skeletal muscle and adipose tissue, resulting in hyperglycemia and dyslipidemia. Clinically, chronic excessive GC signaling is associated with Cushing syndrome and possibly metabolic syndrome²⁸. In most cases, glucose regulation is restored once the corticosteroids are discontinued²⁹. The incidence of hyperglycemia in post-transplant patients receiving corticosteroids is reportedly a substantial 17%-32%³⁰.

THE ROLE OF GLUCAGON-LIKE PEPTIDE-1 IN METABOLISM

Glucagon-like peptide-1 (GLP-1) is a hormone classified as an incretin that responds to an increase in blood glucose levels by promoting insulin secretion. It is produced mainly by enteroendocrine L cells in the distal ileum and colon during the post-translational processing of the proglucagon peptide and is upregulated rapidly by food intake. In the periphery, GLP-1 affects gut motility as well as inhibits glucagon secretion and gastric acid secretion³¹. There is also an effect on appetite and weight control via the central nervous system^{32,33}. One of the most significant effects of GLP-1 is regulating beta-cell function. It is believed to enhance insulin secretion through the control of ion channels involved in K_{ATP}-dependent insulin secretion³¹. GLP-1 stimulates beta cell Ca²⁺ transients through the GLP-1R and these are known to activate the calcium-dependent Cn/NFAT signaling pathway that regulates beta cell proliferation and function in juvenile and adult islets^{34,35}.

The GLP-1 peptide hormone is rapidly degraded by dipeptidyl peptidase-4 (DPP-4) also known as cluster of differentiation antigen 26 (CD26) and neutral endopeptidase 24.11 (NEP 24.11) with a half-life of about 2 minutes³¹. GLP-1 functions by coupling to its specific G-protein receptor, the GLP-1 receptor (GLP-1R) that is expressed mainly in pancreatic beta cells. Because of the rapid de-

struction of the molecule, T2D therapies have focused on treatments such as GLP-1R agonists or DPP-4 inhibitors (DPP-4i) to increase GLP-1 activity. In T2D patients, GLP-1R agonists are associated with improved glycemic control, weight loss, cardiovascular protection and, unlike other secretagogues such as sulfonylureas, lower risk of hypoglycemia^{33,36,37}.

INCRETIN-BASED THERAPIES

There are 2 classes of drugs aimed at modulating the amount of the incretin hormone GLP-1 in the circulation. These are first, the GLP-1R agonists that mimic the action of GLP-1 but are more resistant to rapid inactivation by DPP-4 and second, the DPP-4i that prevent the degradation of endogenous GLP-1³⁸. Found largely in pancreatic beta cells, GLP-1R are also amply present in the gut and central nervous system and to a lesser extent in the heart, lungs, kidney vasculature, pancreatic alpha cells and peripheral nervous system³³. The mechanism of incretin-based agents is glucose-dependent and so the risk of hypoglycemia is minimal.

In 2005 the first GLP-1R agonist therapy approved for use in humans was exenatide. It is a first-in-class diabetes therapy that uses an incretin hormone for metabolic control³⁹. Exenatide is a synthetic version of Exendin-4 that is a naturally occurring component of Gila monster venom first described by Raufman et al⁴⁰ in 1982 who then further explored the peptide in a subsequent study⁴¹. Liraglutide is long-acting GLP-1R agonist and has been found to inhibit cytokine production (IFN-gamma and IL-4) by activated CD4⁺ T cells. It has been shown to be effective not only at improving glucose homeostasis in T2D and promoting weight loss but also at preventing hepatic lipid buildup (hepatic steatosis) in mice⁴² and humans^{32,43}. All the incretin mimetics have similar effects such as stimulation of glucose-dependent insulin release, suppression of glucagon secretion and induction of weight loss. Slower gastric emptying has been associated with short-acting GLP-1R agonists such as short-acting exenatide⁴⁴ and therefore may slow the absorption of IS agents⁴⁵. Cardioprotective effects have been reported in clinical trials, including ELIXA, LEADER, SUSTAIN-6 and RE-WIND, that focused on the evaluation of GLP-1R agonists and cardiovascular (CV) disease in diabetic patients. It was reported that liraglutide, semaglutide and dulaglutide, but not lixisenatide, showed a significant reduction of macrovascular CV events^{32,37,38,46}. Table 2 lists the most common GLP-1R agonists and their brand names, SOTs that have included them in maintenance therapy, and some outcomes (Table 2).

The most common adverse effect of incretin mimetics seems to be nausea that usually resolves over time, but more troubling are reports of incidents of acute pancreatitis, pancreatic cancer, and other malignant neoplasms⁴⁷. These would seem to make GLP-1R agonists less than ideal. However, based on a meta-analysis of the outcomes of CV clinical trials that included incretin-based glucose-lowering medications, the actual increased risk of these incidents that can be attributed to GLP-1R agonists or DPP-4i therapy is negligible⁴⁷.

DPP-4i, also known as gliptins, act to extend incretin function by preventing the degradation of GLP-1 and a similar incretin, gastric inhibitory polypeptide (GIP). DPP-4i are currently used in combination with other T2D drugs to increase insulin secretion and decrease glucagon secretion and glucotoxicity. Studies have shown that DPP-4 has non-incretin substrates and immunomodulatory activity that cleaves cytokines, chemokines and neuropeptides involved in inflammation, immunity and vascular function⁴⁸. In light of this, DPP-4i may protect T2D patients against CVD and microvascular diabetic complications through both GLP-1 and non-GLP-1 dependent mechanisms^{49,50}. Moreover, in pre-clinical and clinical studies, DPP-4i were shown to exert anti-inflammatory and immunomodulatory effects that may contribute to graft survival and inhibit graft dysfunction^{51,52}. To date, there are a few reports of DPP-4i being used to alleviate PTDM. Jin et al⁵³ showed that a DPP-4i improved Tac-induced islet injury and hyperglycemia in rats. In a clinical retrospective cohort study targeting kidney transplant recipients with PTDM, the DPP-4i drug linagliptin was effective in controlling hyperglycemia and led to blood glucose levels comparable to those of kidney graft recipients without PTDM⁵⁴. In another study, lung graft recipients that included a DPP-4i in their treatment had a higher level of the anti-inflammatory cytokine IL-10 at 6 months after transplant resulting in less allograft dysfunction compared to recipients who did not

Table 2. GLP-1 receptor agonists and their clinical use as adjunctive therapy in organ recipients with PTDM.

Name	Alternative/ Brand names	Transplanted Organ	Outcomes	References
Liraglutide Long-acting (~13 hours)	Victoza [®] Saxenda [®]	Kidney Liver Heart Pancreas Lung	 Does not affect Tac levels Reduces risk of cardiovascular events May cause renal impairment Best at reducing HbA1c Reduces appetite Inhibits cytokine production by T cells Suppresses glucagon secretion and hepatic glucose production Increases insulin secretion Potential risk of thyroid tumor* 	Andersen et al ³² Caprio et al ³⁷ Nauck et al ³⁸ Ohki et al ⁴³ Pantalone et al ⁴⁶ Cehic et al ⁵⁷ Nauck et al ⁷⁵
Dulaglutide Long-acting (4-5 days)	Trulicity®	Kidney Liver Heart	 Suppresses glucagon secretion and hepatic glucose production Reduces risk of cardiovascular events Increases insulin secretion Potential risk of thyroid tumor* 	Andersen et al ³² Caprio et al ³⁷ Nauck et al ³⁸ Pantalone et al ⁴⁶ Cehic et al ⁵⁷ Nauck et al ⁷⁵
Exenatide Short-acting (~2.5 hours)	Exendin-4 Byetta [®] Bydureon [®]	Islets of Langerhans	 Nausea, vomiting and diarrhea that decrease with time Possible hypoglycemia Acute pancreatitis reported Renal impairment Reduces liver fat May inhibit beta-cell apoptosis Delays gastric emptying (reduces postprandial glucose excursions) Increases insulin secretion Potential risk of thyroid tumor* 	Parkes et al ³⁹ Raufman et al ⁴⁰ Raufman et al ⁴¹ Gentilella et al ⁴⁴ Vanhove et al ⁴⁵ Abd El Aziz et al ⁴⁷ Aroor et al ⁵⁰ Cehic et al ⁵⁷ Nauck et al ⁷⁵
Exenatide extended- release Long-acting (administered once weekly)	Bydureon BCise®	Islets of Langerhans	 Nausea, vomiting and diarrhea that decrease with time Possible hypoglycemia Acute pancreatitis reported Renal impairment Reduces liver fat May inhibit beta-cell apoptosis Reduces both fasting glucose and postprandial glucose Increases insulin secretion Potential risk of thyroid tumor* 	Parkes et al ³⁹ Raufman et al ⁴⁰ Raufman et al ⁴¹ Abd El Aziz et al ⁴⁷ Aroor et al ⁵⁰ Cehic et al ⁵⁷ Nauck et al ⁷⁵

Abbreviations: HbA1c = glycated hemoglobin, PTDM = post-transplant diabetes mellitus.

receive DPP-4i⁵². For a study involving DPP-4i after pancreatic transplant, the recipients receiving sitagliptin did not require insulin for a longer period of time than recipients receiving standard therapy⁵⁵. The sparse clinical studies mainly covered safety and efficacy in the short-term period. Although there seems to be beneficial outcomes for including DPP-4i in after-transplant therapy,

there is limited information available concerning the long-term use of DPP-4i in PTDM⁵⁶.

WHICH TRANSPLANT RECIPIENTS ARE AT RISK FOR PTDM?

Incidence of PTDM for the most common organ transplants are wide-ranging. Estimates included from different sources are as follows: kidney

^{*}Reported in animal studies, actual risk for humans is unclear.

4%-74%, heart 4%-40%, lung 20%-40% and liver 2.5%-40% ^{3,5,7}. There are several known risk factors for development of PTDM and many of them are the same as the risks for developing T2D such as increasing risk with age (>40 y), body mass index (BMI, >25kg/m²), genetics, family history of T2D, high-risk ethnicity, infections (hepatitis C and cytomegalovirus), cystic fibrosis and polycystic kidney disease. Post-transplant risk factors include type of organ transplanted, post-transplant weight gain, and large doses of IS agents such as prednisolone, Tac and sirolimus^{3,5,13,29}. PTDM has been reported to be decreasing in kidney transplant patients while occurrence in heart transplant patients seems to be increasing⁵⁷.

Pre-operation screening such as oral glucose tolerance tests of transplant candidates for insulin resistance/glucose intolerance could be used as an effective predictor of PTDM and prophylactic therapy (including GLP-1R agonists) may be initiated before the transplant surgery in order to alleviate the transplants-related metabolic consequences⁵⁸. Identification and early management of individuals that are susceptible to develop PTDM would improve quality of life for the patient, extend the functional life of the grafted organ and reduce mortality.

Are GLP-1R agonists effective in treating PTDM?

As yet, there have been very few targeted clinical trials that focus on the use of GLP-1R agonists in transplant recipients⁵⁹. There are, however, retrospective studies and animal experiments that establish the benefit of including a GLP-1R agonist as part of the post-transplant adjunctive therapy. Dai et al⁶⁰ showed that Exendin-4 was able to completely rescue Tac-induced beta cell dysfunction and partially rescue sirolimus-induced beta cell dysfunction in mice. Also in mice, Wang et al⁶¹ concluded that liraglutide alleviates heart graft vasculopathy and fibrosis partially by inhibiting transforming growth factor-β1 (TGF-β1) expression.

Several retrospective or observational clinical studies in T2D or PTDM patients show that treatment with a GLP-1R agonist improved glycemic control and promoted weight loss while having no effect on Tac levels. The PTDM patients included recipients of kidney, heart, pancreas and liver transplants⁶²⁻⁶⁶. Singh et al⁶⁴ reported improvement in chronic kidney disease after treatment with dula-

glutide, as well as a sustained reduction in weight, insulin requirement and improved HbA1c values in 63 SOT recipients, independent of type of transplant.

Is there any alternative to GLP-1R agonists?

Other, older drugs that are commonly used for control of T2D by improving insulin resistance are the thiazolidinediones (TZDs) also known as glitazones. TZDs such as rosiglitazone and pioglitazone are agonists of peroxisome proliferator-activated receptor gamma (PPAR-γ)⁵⁷. These are considered third-line medications and are usually not administered unless primary and secondary T2D medications, such as insulin, metformin and sulfonvlureas, are inadequate at achieving glucose control. TZDs are not known to interact with CNIs but their safety and efficacy for treatment of PTDM has yet to be established¹⁶. This class of drugs has been linked to serious adverse events such as weight gain, edema, heart failure and bladder cancer^{57,67} and are therefore not recommended for management of PTDM after heart transplantation (Table 3). Rosiglitazone has subsequently been withdrawn from several markets in the world due to higher CV risk including stroke⁶⁸.

Sodium-glucose cotransporter 2 (SGLT2) inhibitors reduce glucose reabsorption in the kidney and promote urinary glucose excretion. There is merit for including these agents in treating PTDM as they were tested in T2D patients and found to significantly reduce cardiovascular events and kidney disease progression⁶⁹ and cannot cause hypoglycemia because they work independently of beta-cell function and insulin secretion. Clinical trials using the SGLT2 inhibitor empagliflozin for PTDM after kidney^{70,71} and heart⁷² transplantation have shown promising preliminary results including patient safety, good metabolic control, and reduction in weight with no significant changes in blood pressure or renal function (Table 3).

Conclusions

GLP-1 agonists are very attractive in treating PTDM due to their low hypoglycemia risk and their lack of interaction with common immunosuppressive agents. Clearly, more clinical trials are needed to further explore the routine addition of incretin mimetics to immunosuppression regimens of those patients at risk of developing PTDM.

Table 3. Alternatives to GLP-1 receptor	agonists in organ	recipients with PTDM.
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Name	Alternative/ Brand names	Transplanted Organ	Outcomes	References
Thiazolidinediones (glitazones)	Rosiglitazone (Avandia®) Pioglitazone (Actos®)	Kidney	 Improved insulin sensitivity Edema Weight gain Macular edema Heart failure 	Penfornis et al ¹⁶ Cehic et al ⁵⁷ Zhu et al ⁶⁷ Lu et al ⁶⁸
SGLT2-inhibitors (gliflozins)	Empagliflozin (Jardiance®)	Kidney Heart	 Increased urinary/genital infections Hypoglycemia if combined with sulfonylureas or insulin Greater risk of ketoacidosis in transplant patients Reduced renal glucose reabsorption 	Chilton et al ⁶⁹ Halden et al ⁷⁰ Schwaiger et al ⁷¹ Cehic et al ⁵⁷ Cehic et al ⁷²

Abbreviations: PTDM = post-transplant diabetes mellitus, SGLT2 = Sodium-glucose cotransporter 2.

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The authors declare that they have no conflict of interest to disclose.

REFERENCES

- Jasiak NM, Park JM. Immunosuppression in solid organ transplantation essentials and practical tips. Crit Care Nurs O 2016; 39: 227-240.
- Black CK, Termanini KM, Aguirre O, Hawksworth JS, Sosin M. Solid organ transplantation in the 21(st) century. Ann Transl Med 2018; 6: 409-421.
- 3. Shivaswamy V, Boerner B, Larsen J. Post-transplant diabetes mellitus: causes, treatment and impact on outcomes. Endo Rev 2016; 37: 37-61.
- Davidson J, Wilkinson A; International expert panel on new-onset diabetes after transplantation. New-onset diabetes after transplantation 2003 international consensus guidelines: an endocrinologist's view. Diabetes Care 2004; 27: 805-812.
- Jenssen T, Hartmann A. Post-transplant diabetes mellitus in patients with solid organ transplants. Nat Rev Endocrinol 2019; 15: 172-188.
- Kgosidialwa O, Blake K, O'Connell O, Egan J, O'Neill J, Hatunic M. Post-transplant diabetes mellitus associated with heart and lung transplant. Irish J Med Sci 2019; 189: 185-189
- Munshi VN, Saghafian S, Cook CB, Werner KT, Chakkera HA. Comparison of post-transplantation diabetes mellitus incidence and risk factors between kidney and liver transplantion patients. PLoS One 2020; 15: e0226873.

- Seoane-Pillado MT, Pita-Fernandez S, Valdes-Canedo F, Seijo-Bestillerio R, Pertega-Diaz S, Fernandez-Rivera C, Alonso-Hernandez A, Gonzalez-Martin C, Balboa-Barreiro V. Incidence of cardiovascular events and associated risk factors in kidney transplant patients: a compelling risks survival analysis. BMC Cardiovasc Disord 2017; 17: 72-79.
- Eide IA, Halden TA, Hartmann A, Asberg A, Dahle DO, Reisaeter AV, Jennsen T. Mortality risk in post-transplantation diabetes mellitus based on glucose and HbAlc diagnostic criteria. Transpl Int 2016; 29: 568-578.
- Roccaro GA, Goldberg DS, Hwang WT, Judy R, Thomasson A, Kimmel SE, Forde KA, Lewis JD, Yang YX. Sustained posttransplantation diabetes is associated with long-term major cardiovascular events following liver transplantation. Am J Transplant 2018; 18: 207-215.
- Starzl TE, Marchioro TL, Dickinson TC, Rifkind D, Stonington OG, Waddell WR. Technique of renal homotransplantation. Experience with 42 cases. Arch Surg 1964; 89: 87-104.
- Ducloux D, Courivaud C, Bamoulid J, Crepin T, Gaiffe E, Laheurte C, Vauchy C, Rebibou JM, Saas P, Borot S. Immune phenotype predicts new onset diabetes after kidney transplantation. Hum Immunol 2019; 80: 937-942.
- Chowdhury TA. Post-transplant diabetes mellitus. Clin Med 2019; 19: 392-395.
- 14. Shapiro R, Jordan ML, Scantlebury VP, Vivas C, Fung JJ, McCauley J, Randhawa P, Demetris AJ, Irish W, Mitchell S, Hakala TR, Simmons RL, Starzl TE. A prospective randomized trial of FK506-based Immunosuppression after renal transplantation. Transplantation 1995; 59: 485-490.
- McMahon M, Gerich G, Rizza R. Effects of glucocorticoids on carbohydrate metabolism. Diabetes Metab Review 1988; 4: 17-30.
- 16. Penfornis A, Kury-Paulin S. Immunosuppressive drug-induced diabetes. Diabetes Metab 2006; 32: 539-546.

- 17. Hahn HJ, Dunger A, Laube F, Besch W, Radloff E, Kauert C, Kotzke G. Reversibility of the acute toxic effect of cyclosporin A on pancreatic B cells of Wistar rats. Diabetologia 1986; 29: 489-494.
- Teutonico A, Schena PF, Di Paolo S. Glucose metabolism in renal transplant recipients: effect of calcineurin inhibitor withdrawal and conversion to sirolimus. J Am Soc Nephrol 2005; 16: 3128-3135.
- Farouk SS, Rein JL. The many faces of calcineurin inhibitor toxicity-What the FK? Adv Chronic Kidney Dis 2020; 27: 56-66.
- 20. Terrec F, Jouve T, Naciri-Bennani H. Benhamou PY, Malvezzi P, Janbon B, Giovannini D, Rostaing L, Noble J. Late conversion from calcineurin inhibitors to belatacept in kidney-transplant recipients has a significant beneficial impact on glycemis parameters. Transplant Direct 2019; 6: e517.
- Lawrence MC, Bhatt HS, Watterson JM, Easom RA. Regulation of insulin gene transcription by a Ca2-responsive pathway involving calcineurin and nuclear factor of activated cells. Mol Endocrinol 2001; 15: 1758-1767.
- 22. Grimsby J, Sarabu R, Corbett WL, Haynes NE, Bizzarro FT, Coffey JW, Guertin KR, Hilliard DW, Kester RF, Mahaney PE, Marcus L, Qi L, Spence CL, Tengi J, Magnuson MA, Chu CA, Dvorozniak MT, Matschinsky FM, Grippo JF. Allosteric activators of glucokinase: potential role in diabetes therapy. Science 2003; 301: 370-373.
- 23. Pechhold K, Koczwara K, Zhu X, Harrison VS, Walker G, Lee J, Harlan DM. Blood glucose levels regulate pancreatic beta-cell proliferation during experimentally-induced and spontaneous autoimmune diabetes in mice. PLoS One 2009; 4: e4827.
- 24. Porat S, Weinberg-Corem N, Tornovsky-Babaey S, Schyr-Ben-Haroush R, Hija A, Stolovich-Rain M, Dadon D, Granot Z, Ben-Hur V, White P, Girard CA, Karni R, Kaestner KH, Ashcroft FM, Magnuson MA, Saada A, Grimsby J, Glaser B, Dor Y. Control of pancreatic β cell regeneration by glucose metabolism. Cell Metab 2011; 13: 440-449.
- 25. Salpeter SJ, Klochendler A, Weinberg-Corem N, Porat S, Granot Z, Shapiro AM, Magnuson MA, Eden A, Grimsby J, Glaser B, Dor Y. Glucose regulates cyclin D2 expression in quiescent and replicating pancreatic beta-cells through glycolysis and calcium channels. Endocrinology 2011; 152: 2589-2598.
- Yu H, van Berkel TJ, Biessen EA. Therapeutic potential of VIVT, a selective peptide inhibitor of nuclear factor of activated T cells, in cardiovascular disorders. Cardiovasc Drug Rev 2007; 25: 175-187.
- 27. Goodyer WR, GuX, Liu Y, Bottino R, Crabtree GR, Kim SK. Neonatal beta cell develop in mice and humans is regulated by calcineurin/FAT. Dev Cell 2012; 23: 21-34.
- 28. Bauerle KT, Harris C. Glucocorticoids and diabetes. Mol Med 2016; 113: 378-383.
- 29. Ahmed SH, Biddle K, Augustine T, Azmi S. Post-transplant diabetes mellitus. Diabetes Ther 2020; 11: 779-801.
- Suh S, Park MK. Glucocorticord-induced diabetes mellitus: an important but overlooked problem. Endocrinol Metab 2017; 32: 180-189.

- 31. MacDonald PE, El-Kholy W, Riedel MJ, Salapatek AM, Light PE, Wheeler MB. The multiple actions of GLP-1 on the process of glucose-stimulated insulin secretion. Diabetes 2002; 51: S434-442.
- Andersen A, Lund A, Knop FK, Vilsboll T. Glucagon-like peptide 1 in health and disease. Nat Rev Endocrinol 2018; 14: 390-403.
- 33. Reed J, Bain S, Kanamarlapudi V. Recent advances in understanding the role of glucagon-like peptide 1. F1000Res 2020; 9: F1000.
- 34. Heit JJ, Apelqvist AA, Gu X, Winslow MM, Neilson JR, Crabtree GR, Kim SK. Calcineurin/NFAT signalling regulates pancreatic β-cell growth and function. Nature 2006; 443: 345-349.
- 35. Dai C, Hang Y, Shostak A, Poffenberger G, Hart N, Prasad N, Phillips N, Levy SE, Greiner DL, Shultz LD, Bottino R, Kim SK, Powers AC. Age-dependent human beta cell proliferation induced by glucagon-like peptide 1 and calcineurin signaling. JCI 2017; 127: 3835-3844.
- Scheen AJ. Investigational insulin secretagogues for type
 diabetes. Expert Opin Investig Drugs 2016; 25: 405-422.
- 37. Caprio M, Vitale C, Rosano GMC. From glucose lowering to treatment of cardiovascular disease: the repositioning of glucose-lowering agents. Eur Heart J Cardiovasc Pharmacother 2020; 16: pvaa019.
- 38. Nauck MA, Meier JJ, Cavender MA, Abd El Aziz M, Drucker DJ. Cardiovascular actions and clinical outcomes with glucagon-like peptide-1 receptor agonists and dipeptidyl peptidase-4 inhibitors. Circulation 2017; 136: 849-870.
- 39. Parkes DG, Mace KF, Trautmann ME. Discovery and development of exenatide: the first antidiabetic agent to leverage the multiple benefits of the incretin hormone, GLP-1. Expert Opin Drug Discov 2013; 8: 219-244.
- Raufman JP, Jensen RT, Sutliff VE, Pisano JJ, Gardner JD. Actions of Gila monster venom on dispersed acini from guinea pig pancreas. Am J Physiol 1982; 242: G470-474.
- 41. Raufman JP. Bioactive peptides from lizard venom. Regul Pept 1996; 61: 1-18.
- 42. Itoh A, Irie J, Tagawa H, Kusumoto Y, Kato M, Kobayashi N, Tanaka K, Kikuchi R, Fujita M, Nakajima Y, Wu Y, Yamada S, Kawai T, Ridgway WM, Itoh H. GLP-1 receptor agonist, liraglutide, ameliorates hepatosteatosis induced by anti-CD3 antibody in female mice. J Diabetes Complications 2017; 31: 1370-1375.
- 43. Ohki T, Isogawa A, Iwamoto M, Ohsugi M, Yoshida H, Toda N, Tagawa K, Omata M, Koike K. The effectiveness of liraglutide in nonalcoholic fatty liver disease patients in type 2 diabetes mellitus compared to sitagliptin and pioglitazone. Sci World J 2012; 2012: 496453.
- 44. Gentilella R, Pechtner V, Corcos A, Consoli A. Glucagon-like peptide-1 receptor agonists in type 2 diabetes treatment: are they all the same? Diabetes Metab Res Rev 2019; 35: e3070.
- 45. Vanhove T, Remijsen Q, Kuypers D, Gillard P. Drugdrug interactions between immunosuppressants and antidiabetic drugs in the treatment of post-transplant diabetes mellitus. Transplant Rev (Orlando) 2017; 31: 69-77.

- 46. Pantalone KM, Munir K, Hasenour CM, Atisso CM, Varnado OJ, Maldonado JM, Konig M. Cardiovascular outcomes trials with glucagon-like peptide-1 receptor agonists: A comparison of study designs, populations and results. Diabetes Obes Metab. 2020 Aug 3. doi: 10.1111/ dom.14165. Epub ahead of print.
- 47. Abd El Aziz M, Cahyadi O, Meier JJ, Schmidt WE, Nauck MA. Incretin-based glucose-lowering medications and the risk of acute pancreatitis and malignancies: a meta-analysis based on cardiovascular outcomes trials. Diabetes Obes Metab 2020; 22: 699-704.
- Fadini GP, Avogaro A. Cardiovascular effects of DPP-4 inhibition: beyond GLP-1. Vascul Pharmacol 2011; 55: 10-16.
- Rankovic M, Jeremic N, Srejovic I, Radonjic K, Stojanovic A, Glisic M, Bolevich S, Bolevich S, Jakovljevic V. Dipeptidyl peptidase-4 inhibitors as new tools for cardioprotection. Heart Fail Rev 2020 Jul 21. doi: 10.1007/ s10741-020-10005-5. Epub ahead of print.
- 50. Aroor AR, Sowers JR, Jia G, DeMarco VG. Pleiotropic effects of the dipeptidylpeptidase-4 inhibitors on the cardiovascular system. Am J Physiol Heart Circ Physiol 2014; 307: H477-492.
- 51. Wang X, Zheng P, Huang G, Yang L, Zhou Z. Dipeptidyl peptidase-4(DPP-4) inhibitors: promising new agents for autoimmune diabetes. Clin Exp Med 2018; 18: 473-480.
- Yamada Y, Nishikawa S, Tanaka S, Hamaji M, Nakajima D, Ohsumi A, Chen-Yoshikawa TF, Date H. CD26/DPP4 Inhibitor: a novel prophylactic drug for chronic allograft dysfunction after clinical lung transplantation. J Heart Lung Transplant 2020; 39: S66-S71.
- 53. Jin L, Lim SW, Doh KC, Piao SG, Jin J, Heo SB, Chung BH, Yang CW. Dipeptidyl Peptidase IV inhibitor MK-0626 attenuates pancreatic islet injury in tacrolimus-induced diabetic rats. PLoS One 2014; 9: e100798.
- 54. Thiruvengadam S, Hutchison B, Lim W, Bennett K, Daniels G, Cusack N, Jacques A, Cawley B, Thiruvengadam S, Chakera A. Intensive monitoring for post-transplant diabetes mellitus and treatment with dipeptidyl peptidase-4 inhibitor therapy. Diabetes Metab Syndr 2019; 13: 1857-1863.
- 55. Ergin AB, Poggio E, Krishnamurthi V, Jaber T, Hatipoglu BA. DPP-4 inhibitor therapy in patients after pancreatic transplant. Endocr Pract 2015; 21: 567-573.
- 56. Abdelaziz TS, Ali AY, Fatthy M. Efficacy and Safety of Dipeptidyl peptidase-4 inhibitors in kidney transplant recipients with post-transplant diabetes mellitus (PTDM)a systematic review and meta-analysis. Curr Diabetes Rev 2020; 16: 580-585.
- Cehic MG, Nundall N, Greenfield JR, Macdonald PS. Management strategies for posttransplant diabetes mellitus after heart transplantation: a review. J Transplant 2018: 2018: 1025893.
- 58. Lai X, Zhang L, Fang J, Li G, Xu L, Ma J, Xiong Y, Liu L, Chen Z. OGTT 2-hour serum C-peptide index as a predictor of post-transplant diabetes mellitus in kidney transplant recipients. Ann Transl Med 2019; 7: 538-546.

- 59. Tsai SF, Chen CH. Management of diabetes mellitus in normal renal function, renal dysfunction and renal transplant recipients, focusing on glucagon-like peptide-1 agonist: a review based upon current evidence. Int J Mol Sci 2019; 20: 3152-3161.
- 60. Dai C, Walker JT, Shostak A, Padgett A, Spears E, Wisniewski S, Poffenberger G, Aramandla R, Dean ED, Prasad N, Levy SE, Greiner DL, Shultz LD, Bottino R, Powers AC. Tacrolimus- and sirolimus-induced human beta cell dysfunction is reversible and preventable. JCI Insight 2020; 5: e130770.
- 61. Wang ZM, Wang M, Hu X, Li Y, Ma D-X, Li S-L, Zhao G-Y, Xie Y-N, Shu Y, Yang J. Liraglutide, a glucagon-like peptide-1 receptor agonist, attenuates development of cardiac allograft vasculopathy in a murine heart transplant model. Transplantation 2019; 103: 502-511.
- 62. Thangavelu T, Lyden E, Shivaswamy V. A retrospective study of glucagon-like peptide 1 receptor agonists for the management of diabetes after transplantation. Diabetes Ther 2020; 11: 987-994.
- 63. Kukla A, Hill J, Merzkani M, Bentall A, Lorenz EC, Park WD, D'Costa M, Kudva YC, Stegall MD, Shah P. The use of GLP1R agonists for the treatment of type 2 diabetes in kidney transplant recipients. Transplant Direct 2020; 6: e524.
- 64. Singh P, Pesavento TE, Washburn K, Walsh D, Meng S. Largest single-centre experience of dulaglutide for management of diabetes mellitus in solid organ transplant recipients. Diabetes Obes Metab 2019; 21: 1061-1065.
- Cariou B, Bernard C, Cantarovich D. Liraglutide in whole-pancreas transplant patients with impaired glucose homoeostasis: a case series. Diabetes Metab 2015; 41: 252-257.
- 66. Liou JH, Liu YM, Chen CH. Management of diabetes mellitus with glucagonlike peptide-1 agonist liraglutide in renal transplant recipients: a retrospective study. Transplant Proc 2018; 50: 2502-2505.
- 67. Zhu J, Yu X, Zheng Y, Li J, Wang Y, Lin Y, He Z, Zhao W, Chen C, Qiu K, Wu J. Association of glucose-lowering medications with cardiovascular outcomes: an umbrella review and evidence map. Endocrinol 2020; 8: 192-205
- 68. Lu CJ, Sun Y, Muo CH, Chen RC, Chen PC, Hsu CY. Risk of stroke with thiazolidinediones: a ten-year nation-wide population-based cohort study. Cerebrovasc Dis 2013; 36: 145-151.
- 69. Chilton RJ. Effects of sodium-glucose cotransporter-2 inhibitors on the cardiovascular and renal complications of type 2 diabetes. Diabetes Obes Metab 2020; 1: 16-29.
- 70. Halden TAS, Kvitne KE, Midtvedt K, Rajakumar L, Robertsen I, Brox J, Bollerslev J, Hartmann A, Asberg A, Jenssen T. Efficacy and safety of empagliflozin in renal transplant recipients with posttransplant diabetes mellitus. Diabetes Care 2019; 42: 1067-1074.
- 71. Schwaiger E, Burghart L, Signorini L, Ristl R, Kopecky C, Tura A, Pacini G, Wrba T, Antlanger M, Schmaldienst S, Werzowa J, Säemann MD, Hecking M. Empagliflozin in posttransplantation diabetes mellitis: a prospective,

- interventional pilot study on glucose metabolism, fluid volume, and patient safety. Am J Transplant 2019; 19: 907-919.
- 72. Cehic MG, Muir CA, Greenfield JR, Hayward C, Jabbour A, Keogh A, Kotlyar E, Muthiah K, Macdonald PS. Efficacy and safety of empagliflozin in the management of diabetes mellitus in heart transplant recipients. Transplant Direct 2019; 5: e450.
- 73. Barlow AD, Nicholson ML, Herbert TP. Evidence for rapamycin toxicity in pancreatic β-cells and a review of the underlying molecular mechanisms. Diabetes 2013; 62: 2674-2682.
- 74. Bianchi A, Ciccarelli O. Daclizumab-induced encephalitis in multiple sclerosis. Mult Scler 2019; 25: 1557-1559.
- 75. Nauck MA, Friedrich N. Do GLP-1-based therapies increase cancer risk? Diabetes Care 2013; 36: S245-S252.