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### **Paper:**

Zhang, B., Tang, S., Chen, X., Zhang, G., Zhang, W., Chen, T., Liu, G., Li, S., Dos Santos, L., et. al. (2018).  
*Streptomyces qaidamensis* sp. nov., isolated from sand in the Qaidam Basin, China. *The Journal of Antibiotics*  
<http://dx.doi.org/10.1038/s41429-018-0080-9>

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*Streptomyces qaidamensis* sp. nov., isolated from sand in the Qaidam  
Basin, China

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1 **Abstract**

2 A novel actinobacterial strain, designated S10<sup>T</sup>, was isolated from a sand sample  
3 collected from the Qaidam Basin in Qinghai province, China. The strain S10<sup>T</sup> exhibited  
4 antibacterial activity against MRSA. The taxonomic position of the strain S10<sup>T</sup> was  
5 determined by a polyphasic approach. There were 6 copies of 16S rDNA in S10<sup>T</sup> which were  
6 not same exactly (MH257693-MH257698). Phylogenetic analysis of 16S rRNA gene  
7 sequences indicated the strain belonging to the genus *Streptomyces* and it showed high  
8 sequence similarities with *Streptomyces chartreusis* NBRC 12753<sup>T</sup> (99.31%), *Streptomyces*  
9 *phaeoluteigriseus* DSM 41896<sup>T</sup> (99.24%), *Streptomyces variegatus* NRRL B-16380<sup>T</sup> (99.17%)  
10 and *Streptomyces flavovariabilis* NRRL B-16367<sup>T</sup> (99.17%) comparing with MH257693,  
11 MH257695, MH257696, MH257697, and MH257698. Similarities with *Streptomyces*  
12 *kunmingensis* NBRC14463<sup>T</sup> (98.82%), *Streptomyces bungoensis* DSM 41781<sup>T</sup> (98.76%), *S.*  
13 *chartreusis* NBRC 12753<sup>T</sup> (98.69%) and *S. phaeoluteigriseus* DSM 41896<sup>T</sup> (98.62%) with  
14 MH257694. Whole-genome average nucleotide identity (ANI) values between strain S10<sup>T</sup>  
15 and *S. chartreusis* NBRC 12753<sup>T</sup>, *S. phaeoluteigriseus* DSM 41896<sup>T</sup>, *S. variegatus* NRRL B-  
16 16380<sup>T</sup>, *S. flavovariabilis* NRRL B-16367<sup>T</sup>, *S. kunmingensis* NBRC 14463<sup>T</sup>, *S. bungoensis*  
17 DSM 41781<sup>T</sup> were 83.63%, 82.89%, 92.55%, 92.51%, 79.29 and 82.87%, respectively,  
18 suggesting that the strain S10<sup>T</sup> represented a new species. A phylogenetic analysis comparing  
19 the S10<sup>T</sup> genome with those of 336 other sequenced *Streptomyces* genomes confirmed its  
20 relatedness with *Streptomyces variegatus* NRRL B-16380<sup>T</sup> and *Streptomyces flavovariabilis*  
21 NRRL B-16367<sup>T</sup>. Strain S10<sup>T</sup> contained LL-diaminopimelic acid in the cell wall. The  
22 predominant menaquinones were MK-9(H<sub>6</sub>) and MK-9(H<sub>8</sub>) and the major fatty acids were  
23 iso-C<sub>15:0</sub>, anteiso-C<sub>15:0</sub>, iso-C<sub>16:0</sub>, and anteiso-C<sub>17:0</sub>. Phospholipids detected were diphosphatidyl  
24 glycerol, phosphatidyl ethanolamine, phosphatidyl choline, three unknown phospholipids, an  
25 unknown aminophospholipid and an unknown phosphatidyl glycolipid. On the basis of these  
26 genotypic and phenotypic data, it is proposed that isolate S10<sup>T</sup> (=JCM 31184<sup>T</sup> =CGMCC  
27 4.7315<sup>T</sup>) should be classified in the genus *Streptomyces* as *Streptomyces qaidamensis* sp. nov.

## 28 INTRODUCTION

29 The genus *Streptomyces* was first described by Waksman and Henrici.<sup>1</sup> Members of the  
30 genus *Streptomyces* are aerobic, Gram-positive filamentous bacteria which can form branched  
31 substrate and aerial mycelia. They have LL-diaminopimelic acid with no characteristic sugars  
32 in the cell wall<sup>2</sup> and have high genomic DNA G+C contents.<sup>3,4</sup> *Streptomyces* strains are an  
33 important source of a broad range of bioactive secondary metabolites, which are widely used  
34 in the fields of food, agriculture and pharmaceutical industries.<sup>5,6</sup>

35 *Streptomyces* can live in a range of environments, including both fertile and barren  
36 soils.<sup>7-9</sup> When we investigated the diversity of actinobacteria in the west of China, a novel  
37 actinobacterium producing antibiotic activity against Methicillin-resistant *Staphylococcus*  
38 *aureus* (MRSA) was isolated from sand collected from the Qaidam Basin, China. MRSA is a  
39 major cause of hospital-acquired infections, and has acquired resistance to many current  
40 frontline antibiotic classes, and consequently it is becoming increasingly difficult to combat  
41 MRSA infections.<sup>10</sup> The strain S10<sup>T</sup> may produce new bioactive compounds with anti-MRSA  
42 activity. Consequently, it is of considerable interest for further research.

43

## 44 MATERIALS AND METHODS

### 45 Bacterial strains and isolation

46 Strain S10<sup>T</sup> was isolated from a sand sample collected in the Qaidam Basin in Qinghai  
47 province, China, by using Gause's synthetic agar medium (20.0 g soluble starch, 1.0 g KNO<sub>3</sub>,  
48 0.5 g K<sub>2</sub>HPO<sub>4</sub>·3H<sub>2</sub>O, 0.5 g MgSO<sub>4</sub>·7H<sub>2</sub>O, 0.001 g FeSO<sub>4</sub>, 0.5 g NaCl and 20.0 g agar in 1.0  
49 liter tap water, pH 7.2), supplemented with nalidixic acid (25 µg ml<sup>-1</sup>) incubated for 7 days at  
50 28 °C. The strain was stored at -86 °C in the presence of 20 % (v/v) glycerol. The reference  
51 strains were *Streptomyces chartreusis* NBRC 12753<sup>T</sup>, *Streptomyces variegatus* NRRL B-  
52 16380<sup>T</sup>, *Streptomyces flavovariabilis* NRRL B-16367<sup>T</sup> and *Streptomyces kunmingensis* NBRC  
53 14463<sup>T</sup>.

### 54 Morphological, physiological and biochemical tests

55 The morphology of spore-chains and hyphae were determined by light microscopy (BH-

56 2; Olympus) and scanning electron microscopy (SU8010, Hitachi; JSM-5600, JEOL) using  
57 cultures grown on Gause's synthetic agar medium at 30 °C for 20 days. Cultural  
58 characteristics was examined after growth on standard media ISP 2-7<sup>11</sup>, Czapek's agar<sup>12</sup> and  
59 nutrient agar after incubation at 30 °C for 14 days. The utilization of sole carbon and nitrogen  
60 sources, and metabolism of starch and cellulose, were examined as described previously.<sup>13,14</sup>  
61 Growth at various temperatures (4, 10, 15, 20, 30, 37, 40, 45 and 50 °C) and NaCl  
62 concentrations (0-10 %) was examined on yeast extract-malt extract (ISP 2). The pH range  
63 and the optimum pH were determined by incubating at 28 °C in ISP 2 broth with the pH  
64 adjusted to between 4 – 12 by addition of KH<sub>2</sub>PO<sub>4</sub>/HCl, KH<sub>2</sub>PO<sub>4</sub>/K<sub>2</sub>HPO<sub>4</sub> and  
65 K<sub>2</sub>HPO<sub>4</sub>/NaOH (at intervals of 1.0 pH unit). The antibacterial activity of strain S10<sup>T</sup> was  
66 determined using a cylinder plug antibacterial bioassay using a clinical methicillin-resistant  
67 *Staphylococcus aureus* isolate, a gift of Jodi Lindsay, St George's Hospital, London, as the  
68 indicator strain.<sup>15</sup> This indicator strain, EMRSA-8, was resistant to a range of beta-lactam  
69 antibiotics, ciprofloxacin, erythromycin, rifampicin, tetracycline and trimethoprim, but  
70 sensitive to vancomycin.

## 71 **Chemotaxonomy**

72 Hyphae for chemotaxonomic studies were prepared by growing the strain in TSB  
73 medium in shake flasks for 10 days at 30 °C. The hyphae were harvested by centrifugation  
74 and washed twice with distilled water. Then the cells were recentrifuged and freeze-dried. The  
75 diaminopimelic acid isomers in the cell wall and whole-cell sugars were analyzed with the  
76 method described by Lechevalier and Lechevalier<sup>2</sup> and Stanek and Roberts<sup>16</sup>, respectively.  
77 The menaquinones were analyzed by the method of Collins *et al.*<sup>17</sup> and analysed by HPLC<sup>18</sup>.  
78 The polar lipids were examined using two-dimensional TLC and identified according to  
79 method of Minnikin *et al.*<sup>19</sup> The cellular fatty acids methyl esters were extracted by the  
80 method of Sasser<sup>20</sup> and analysis by according to the standard protocol of the Sherlock  
81 Microbial identification (MIDI) system.<sup>21</sup>

## 82 **Molecular analysis**

83 The genomic DNA of strain S10<sup>T</sup> was extracted and the 16S rRNA gene was amplified

84 as described by Harunari *et al.*<sup>22</sup> Closely related 16S rRNA gene sequences to that of strain  
85 S10<sup>T</sup> were identified using the EzTaxon-e server.<sup>23</sup> A phylogenetic tree was generated using  
86 the neighbour-joining,<sup>24</sup> maximum-parsimony<sup>25</sup> and maximum-likelihood<sup>26</sup> algorithms in  
87 MEGA5.0.<sup>27</sup> Evolutionary distances were calculated using the model of Jukes and Cantor.<sup>28</sup>  
88 Topologies of the resultant tree were evaluated by bootstrap analyses<sup>29</sup> based on 1000  
89 resamplings. The G+C content of the DNA was examined by HPLC according to the method  
90 of Tamaoka and Komagata.<sup>30</sup> The genomic DNA of strain S10<sup>T</sup> was also used to obtain a draft  
91 genome sequence using Illumina sequencing; the draft genome consisted of 1 contigs with an  
92 estimated genome size of 8.66 Mb(CP015098). The whole-genome average nucleotide  
93 identity (ANI) value were calculated by Goris, *et al.*<sup>31</sup> A maximum-composite likelihood tree  
94 was performed using PhyloPhlAn and the method of Segata *et al.*<sup>32</sup> Initially, all protein  
95 sequences from all annotated *Streptomyces* genomes (.faa files) were retrieved autonomously  
96 from the GenBank FTP site (last accessed November 2016) using the term “*Streptomyces*” as  
97 a query. Ortholog identification and alignment was performed in PhyloPhlAn using the “-u”  
98 command. A maximum likelihood phylogeny was reconstructed from the concatenated  
99 alignments in FastTree MP implemented by the Cipres Science Gateway Server<sup>33</sup>. The tree  
100 was drawn using the JTT+CAT model with 20 discrete categories (-cat 20). Topology  
101 refinement was performed using the follow parameters: -nni 10 -spr 2 -sprlength 10. Nodal  
102 support was inferred from 1000 bootstrap pseudoreplicates.

### 103 **Nucleotide sequence accession number**

104 The GenBank/EMBL/DDBJ accession number for the 16S rRNA gene sequence of strain  
105 S10<sup>T</sup> are MH257693- MH257698. The GenBank accession number for the genome of S10<sup>T</sup> is  
106 CP015098.1.

## 107 **RESULTS AND DISCUSSION**

### 108 **Morphological, cultural and physiological characteristics**

109 Strain S10<sup>T</sup> formed extensively branched substrate hyphae, and the aerial hyphae formed  
110 straight spore chains (Figure 1). The spores were cylindrical with a rough-textured surface  
111 (Figure 1A). The growth characteristics of strain S10<sup>T</sup> cultured on different growth media

112 were compared to those of the reference strains *Streptomyces chartreusis* NBRC 12753<sup>T</sup>,  
113 *Streptomyces variegatus* NRRL B-16380<sup>T</sup>, *Streptomyces flavovariabilis* NRRL B-16367<sup>T</sup> and  
114 *Streptomyces kunmingensis* NBRC 14463<sup>T</sup> (Table S1). Strain S10<sup>T</sup> grew well on ISP medium  
115 2-7, Czapek agar and nutrient agar. Sporulation was poor on ISP 5 and nutrient agar, and no  
116 sporulation or growth of aerial hyphae was observed for cultures grown on ISP 6 medium.  
117 Black soluble pigments were produced on ISP 6 and brown pigments were produced on  
118 nutrient agar. Prior to sporulation, aerial mycelia produced on all media except ISP 6 were  
119 white. The morphological features of isolate S10<sup>T</sup> were consistent with its classification in the  
120 genus *Streptomyces*<sup>34</sup> and were distinct from those of the reference strains.

121 The physiological properties of strain S10<sup>T</sup>, *Streptomyces chartreusis* NBRC 12753<sup>T</sup>,  
122 *Streptomyces variegatus* NRRL B-16380<sup>T</sup>, *Streptomyces flavovariabilis* NRRL B-16367<sup>T</sup> and  
123 *Streptomyces kunmingensis* NBRC13368<sup>T</sup> were compared and found to be different (Table 1).  
124 Strain S10<sup>T</sup> could utilize L-*myo*-inositol, L-arabinose, D-fructose, D-lactose, D-mannitol, D-  
125 raffinose, L-rhamnose or D-xylose as sole carbon sources. It also utilized L-alanine, L-  
126 asparagine or L-histidine as sole nitrogen sources, but not L-leucine and L-cysteine (Table 1).  
127 The strain S10<sup>T</sup> could degrade starch, cellulose, gelatin, tween 20, tween 80 or urea. The  
128 temperature range for growth of strain S10<sup>T</sup> was 20-40 °C (optimum temperature 30 °C). The  
129 pH range for growth was 6–11 (optimum pH 8.0). The maximum NaCl concentration for  
130 growth was 10% (w/v) (optimum 1%).

131 Cylinder plugs of strain S10<sup>T</sup> grown for 10 days on ISP 4 medium produced clearing  
132 zones of 10 (+/- 2) mm, compared to zones of 13 mm (+/- 2) mm produced when 10 µl of a  
133 50 mg/ml solution of vancomycin was applied to a 10 mm diameter sterile filter disc on the  
134 lawn of an MRSA strain bacteria, indicating that the strain S10<sup>T</sup> produces antibiotic activity  
135 against MRSA.

### 136 **Chemotaxonomic characteristics**

137 Cell wall analysis indicated that strain S10<sup>T</sup> contained LL-diaminopimelic acid, as is  
138 characteristic for the genus *Streptomyces*. The whole-cell hydrolysate contained galactose and  
139 ribose. The predominant isoprenoid quinone compound were MK-9(H<sub>6</sub>) (61.3%) and MK-

140 9(H<sub>8</sub>) (21.5%). The polar lipids detected were diphosphatidyl glycerol, phosphatidyl  
141 ethanolamine, phosphatidyl choline, three unknown phospholipids, an unknown  
142 aminophospholipid and an unknown phosphatidyl glycolipid (Figure S1). The major fatty  
143 acids were anteiso-C<sub>15:0</sub> (31.3%), anteiso-C<sub>17:0</sub> (17.5%), iso-C<sub>15:0</sub> (12.8%) and iso-C<sub>16:0</sub> (11.7%);  
144 the fatty acid composition was different to *Streptomyces chartreusis* NBRC 12753<sup>T</sup>,  
145 *Streptomyces variegatus* NRRL B-16380<sup>T</sup> and *Streptomyces flavovariabilis* NRRL B-16367<sup>T</sup>  
146 (Table 2). The DNA G+C content of strain S10<sup>T</sup> was 71.3 mol%.

#### 147 **Molecular analysis**

148 There were 6 copies of 16S rDNA in S10<sup>T</sup> which were not same exactly. There were high  
149 sequence similarities between strain S10<sup>T</sup> and *Streptomyces chartreusis* NBRC 12753<sup>T</sup>  
150 (99.31%), *Streptomyces phaeoluteigriseus* DSM 41896<sup>T</sup> (99.24%), *variegatus* NRRL B-  
151 16380<sup>T</sup> (99.17%) and *Streptomyces flavovariabilis* NRRL B-16367<sup>T</sup> (99.17%) comparing with  
152 MH257693, MH257695, MH257696, MH257697, and MH257698. Similarities with  
153 *Streptomyces kunmingensis* NBRC14463<sup>T</sup> (98.82%), *Streptomyces bungoensis* DSM  
154 41781<sup>T</sup> (98.76%), *S. chartreusis* NBRC 12753<sup>T</sup> (98.69%) and *S. phaeoluteigriseus* DSM  
155 41896<sup>T</sup> (98.62%) with MH257694. Based on the 16S rRNA gene sequence, phylogenetic  
156 analysis also confirmed that strain S10<sup>T</sup> represented a member of the genus *Streptomyces*  
157 (Figure 2). The whole-genome average nucleotide identity (ANI) value between strain S10<sup>T</sup>  
158 and *S. chartreusis* NBRC 12753<sup>T</sup>, *S. phaeoluteigriseus* DSM 41896<sup>T</sup>, *S. variegatus* NRRL B-  
159 16380<sup>T</sup>, *S. flavovariabilis* NRRL B-16367<sup>T</sup>, *S. kunmingensis* NBRC 14463<sup>T</sup>, *S. bungoensis*  
160 DSM 41781<sup>T</sup> were 83.63%, 82.89%, 92.55%, 92.51%, 79.29% and 82.87%, respectively.  
161 These values were below the species demarcation threshold of 95–96% ANI suggested for  
162 prokaryotic species<sup>35</sup>, indicating that S10<sup>T</sup> was a new *Streptomyces* species. A phylogenetic  
163 tree based on 16S rRNA sequences alone was found to be unstable. As a consequence, a  
164 maximum-composite likelihood tree based on analysis of 400 protein sequences from 336  
165 sequenced *Streptomyces* genomes was constructed. This analysis indicated strain S10<sup>T</sup> shared  
166 highest similarity with *S. variegatus* NRRL B-16380<sup>T</sup> and *S. flavovariabilis* NRRL B-16367<sup>T</sup>  
167 (Figure S2).



168 Based on its phenotypic, phylogenetic and chemotaxonomic characteristics, strain S10<sup>T</sup>  
169 represents a novel species within the genus *Streptomyces*, for which the name *Streptomyces*  
170 *qaidamensis* sp. nov. is proposed.

171

172 **Description of *Streptomyces qaidamensis* sp. nov.**

173 *Streptomyces qaidamensis* (qai.dam.en'sis. N.L. masc. adj. *qaidamensis* pertaining to  
174 Qaidam, China, where the type strain was isolated).

175 Cells are aerobic and Gram-stain-positive. The substrate hyphae are branched and aerial  
176 mycelia formed straight spore chains. The spores are cylindrical with a rough-textured surface.  
177 It grow well on ISP medium 2-7, Czapek agar and nutrient agar. Sporulation is poor on ISP 5  
178 and nutrient agar, and not detected on ISP 6. Aerial hyphae are white on all media except ISP  
179 6. Production of black soluble pigments is observed on ISP 6 and brown pigments are  
180 produced on nutrient agar. Growth occur between 20-40 °C (optimum temperature 30 °C), at  
181 pH 6-11 (optimum pH 8.0) and with 0-10% (w/v) NaCl. It utilize *myo*-inositol, L-arabinose,  
182 D-fructose, D-lactose, D-mannitol, D-raffinose, L-rhamnose or D-xylose as sole carbon  
183 sources. It also utilize L-alanine, L-asparagine or L-histidine as sole nitrogen sources, but not  
184 L-leucine and L-cysteine. The strain S10<sup>T</sup> degrade starch, cellulose, gelatin, tween 20, tween  
185 80 or urea. The cell wall contain LL-diaminopimelic acid, galactose and ribose. The  
186 predominant isoprenoid quinone compounds are MK-9(H<sub>6</sub>) and MK-9(H<sub>8</sub>). The polar lipids  
187 detected are diphosphatidyl glycerol, phosphatidyl ethanolamine, phosphatidyl choline, three  
188 unknown phospholipids, an unknown aminophospholipid and an unknown phosphatidyl  
189 glycolipid. The major fatty acids are anteiso-C<sub>15:0</sub>, anteiso-C<sub>17:0</sub>, iso-C<sub>15:0</sub> and iso-C<sub>16:0</sub>. The  
190 DNA G+C content of strain S10<sup>T</sup> is 71.3 mol%.

191 The type strain, S10<sup>T</sup> (=JCM 31184<sup>T</sup> =CGMCC 4.7315<sup>T</sup>) is isolated from a sand sample  
192 collected from the Qaidam Basin, China.

193

194 **Acknowledgments**

195 This work was funded by the National Science Foundation of China (No. 31470544,  
196 31570498), National Science Foundation of Gansu (17JR5RA308), SKLCS-OP-2018-10,  
197 BBSRC, UK (grant BB/J020419/1) and CAPES, Brazil (88887.125175/2015-00).

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Table 1. Phenotypic properties of strain S10<sup>T</sup> and related type species

Strains: 1, S10<sup>T</sup>; 2, *S. chartreusis* NBRC 12753<sup>T</sup>; 3, *S. kunmingensis* NBRC13368<sup>T</sup>(data from Li *et al.*, 2013, Ruan *et al.*, 1985);<sup>36-37</sup> 4, *S. flavovariabilis* B-16367<sup>T</sup> (data from Zhang *et al.*, 2016);<sup>38</sup> 5, *S. variegatus* NRRL B-16380<sup>T</sup> (data from Zhang *et al.*, 2016).<sup>38</sup> All data were obtained in this study except where indicated otherwise. Abbreviations: +, positive; w, weakly positive; -, negative; N, not determined. All strains were positive for utilization of L-arabinose and D-fructose.

Characteristics	1	2	3	4	5
NaCl for growth(% w/v)	0-10	0-5	5	>=7	>=7
Carbon source utilization (1.0%, w/v)					
myo-inositol	+	+	+	N	N
D-lactose	+	-	+	-	+
D-raffinose	+	-	+	+	+
L-rhamnose	+	+	+	+	-
D-mannitol	+	+	+	+	-
D-xylose	+	-	+	-	+
Starch	+	+	-	+	+
Nitrogen source utilization (0.1%, w/v)					
L-leucine	-	-	N	N	N
L-cysteine	-	+	N	N	N
L-alanine	+	+	N	N	N
L-asparagine	+	+	N	N	N
L-histidine	+	+	N	N	N
Degradation					
Tween 20	+	-	N	N	N
Tween 80	+	-	N	N	N
Cellulose	+	-	-	-	+
Gelatin liquefaction	+	+	-	+	+
Urease test	+	+	+	+	+

Table 2. Cellular fatty acid composition of strain S10<sup>T</sup> and related type species.

Strains: 1, S10<sup>T</sup>; 2, *S. chartreusis* NBRC 12753<sup>T</sup>; 3, *S. flavovariabilis* B-16367<sup>T</sup> (data from Zhang *et al.*, 2016);<sup>38</sup> 4, *S. variegatus* NRRL B-16380<sup>T</sup> (data from Zhang *et al.*, 2016).<sup>38</sup> All data were obtained in this study except where indicated otherwise.

Fatty acid	1	2	3	4
C <sub>14:0</sub>	0.31	0.92	2.7	0.3
C <sub>16:0</sub>	5.89	7.13	8.5	2.9
iso-C <sub>14:0</sub>	1.23	6.36	10.5	2.4
iso-C <sub>15:0</sub>	12.82	5.49	8.9	4.0
anteiso-C <sub>15:0</sub>	31.30	12.09	8.7	27.8
iso-C <sub>16:0</sub>	11.71	32.07	28.8	22.6
iso-C <sub>16:1</sub> H	1.45	9.59	4.4	5.8
iso-C <sub>17:0</sub>	4.10	0.96	1.1	1.0
cyclo-C <sub>17:0</sub>	ND	1.05	ND	ND
anteiso-C <sub>17:0</sub>	14.53	5.44	1.9	15.5
anteiso-C <sub>17:1</sub> ω9c	5.09	4.80	1.1	9.0
C <sub>17:1</sub> ω8c	1.30	0.34	0.2	ND
Sum In Feature 3	4.98	6.58	18.4	3.2
Sum In Feature 9	3.21	2.69	1.5	2.2

ND no-detected.

The amount of fatty acid was omitted when they all less than 1%.

Summed Feature 3: C<sub>16:1</sub> ω7c/ω6c or C<sub>16:1</sub> ω6c/ω7c; Summed Feature 9: C<sub>16:0</sub> 10-methyl or iso-C<sub>17:1</sub>ω9c.

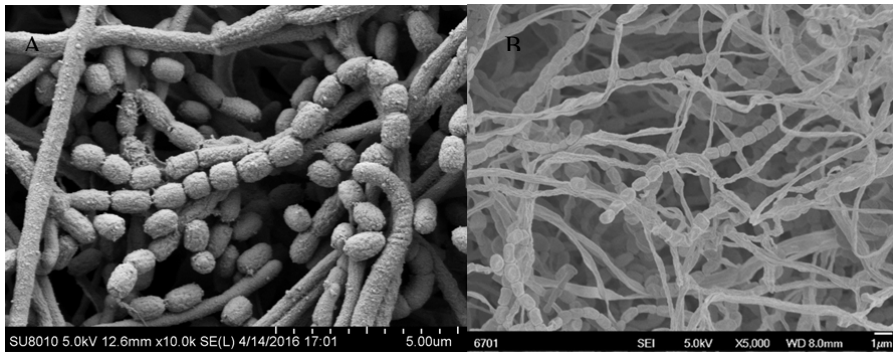


Figure 1 Scanning electron micrograph of strain S10<sup>T</sup> cultivated on Gause's synthetic agar at 30°C for 20 days. A taking with SU8010, B taking with JSM-5600.

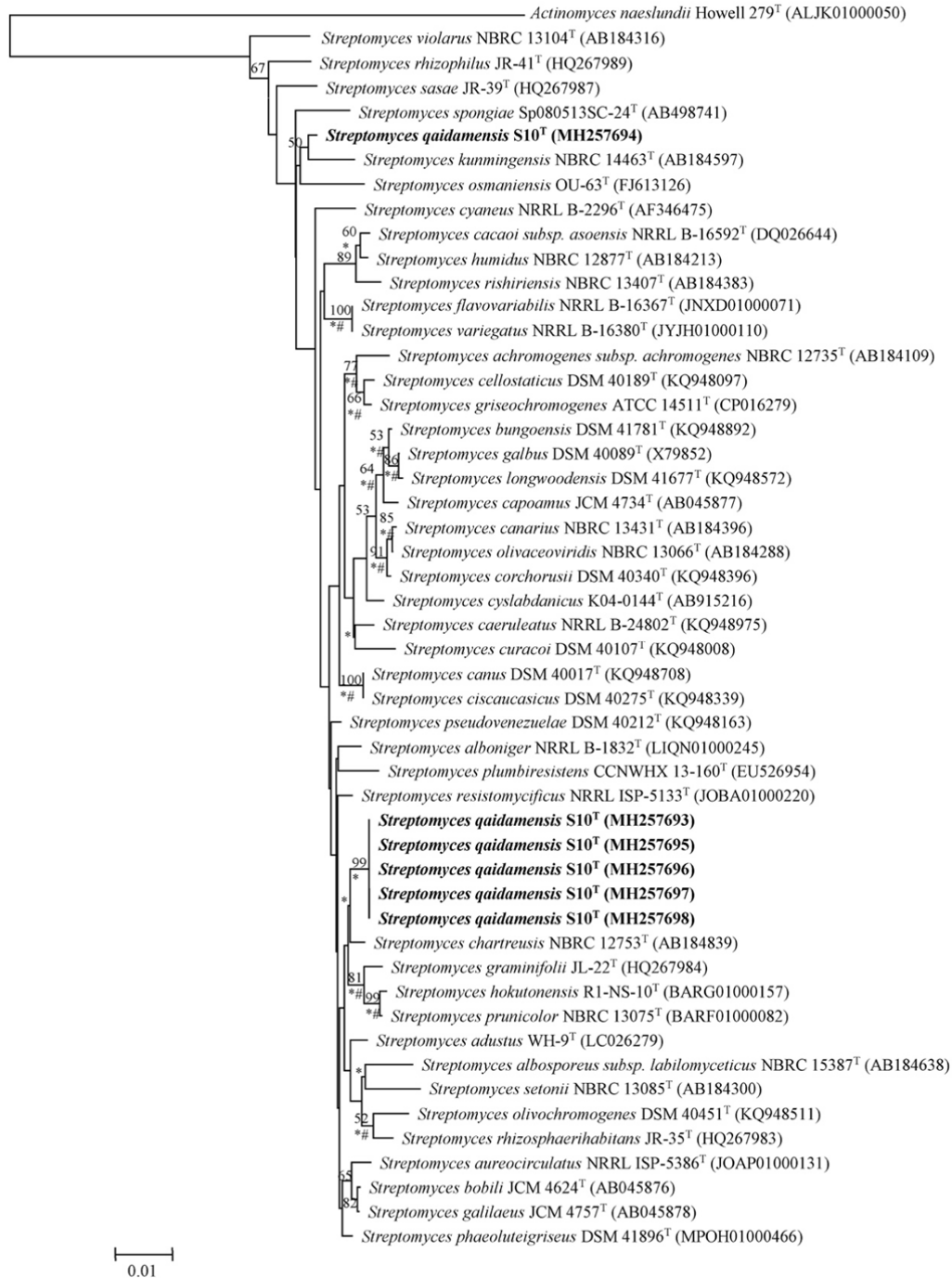


Figure 2 Neighbor-joining phylogenetic tree, based on nearly complete 16S rRNA gene sequences, showing the relationships between strain S10<sup>T</sup> and strains of related species of the genus *Streptomyces*. Numbers at nodes are bootstrap values based on 1000 re-samplings (only values above 50% are shown). Asterisks(\*, #) indicate that the clades are recovered in maximum-likelihood and maximum-parsimony trees, respectively.