



King Saud University  
Saudi Journal of Biological Sciences

[www.ksu.edu.sa](http://www.ksu.edu.sa)  
[www.sciencedirect.com](http://www.sciencedirect.com)



## ORIGINAL ARTICLE

# *In vitro* importance of probiotic *Lactobacillus plantarum* related to medical field



Mariadhas Valan Arasu<sup>a</sup>, Naif Abdullah Al-Dhabi<sup>a</sup>, Soundharrajan Ilavenil<sup>b</sup>,  
Ki Choon Choi<sup>b,\*</sup>, Srisesharam Srigopalram<sup>b</sup>

<sup>a</sup> Department of Botany and Microbiology, Addiriyah Chair for Environmental Studies, College of Science, King Saud University, P.O. Box 2455, Riyadh 11451, Saudi Arabia

<sup>b</sup> Grassland and Forage Division, National Institute of Animal Science, RDA, Seonghwan-Eup, Cheonan-Si 330 801, Chungnam, Republic of Korea

Received 30 March 2015; revised 12 September 2015; accepted 14 September 2015

Available online 9 October 2015

## KEYWORDS

*Lactobacillus plantarum*;  
Medical applications

**Abstract** *Lactobacillus plantarum* is a Gram positive lactic acid bacterium commonly found in fermented food and in the gastro intestinal tract and is commonly used in the food industry as a potential starter probiotic. Recently, the consumption of food together with probiotics has tremendously increased. Among the lactic acid bacteria, *L. plantarum* attracted many researchers because of its wide applications in the medical field with antioxidant, anticancer, anti-inflammatory, antiproliferative, anti-obesity and antidiabetic properties. The present study aimed to investigate the *in vitro* importance of *L. plantarum* toward medical applications. Moreover, this report short listed various reports related to the applications of this promising strain. In conclusion, this study would attract the researchers in commercializing this strain toward the welfare of humans related to medical needs.

© 2015 Production and hosting by Elsevier B.V. on behalf of King Saud University. This is an open access article under the CC BY-NC-ND license (<http://creativecommons.org/licenses/by-nc-nd/4.0/>).

## 1. Introduction

The human digestive system contains approximately four hundred different bacterial species and its abundance differs between individuals. Among them few probiotic *Lactobacillus*

species namely, *Lactobacillus acidophilus*, *Lactobacillus pentosus*, *Lactobacillus brevis*, *Lactobacillus lactis*, *Lactobacillus amylovorus*, *Lactobacillus casei*, *Lactobacillus bulgaricus*, *Lactobacillus fermentum*, *Lactobacillus plantarum* and *Lactobacillus rhamnosus* specifically produce extracellular proteins, exopolysaccharides, bacteriocins and lipoteichoic acids which influence the health and physiology of the host by interacting with the epithelial cells and enhance the host immune system (Sanchez et al., 2010). *Lactobacillus* strains are recognized as safe for consumption because of their presence in food and their role in the gut defense mechanism. Of the *Lactobacillus* strains, *L. plantarum* is a Gram positive, short-rod, micro-aerophilic, acid-tolerant, non-spore forming,

\* Corresponding author. Tel.: +82 41 580 6752; fax: +82 41 580 6779.

E-mail address: [choiwh@korea.kr](mailto:choiwh@korea.kr) (K.C. Choi).

Peer review under responsibility of King Saud University.



Production and hosting by Elsevier

non-respiring, low G + C content, hetero-fermentative group of lactobacilli with a range of applications in the food industry as a starter culture and preservatives (Arasu et al., 2013). It is a non-spore forming bacterium which produces organic acids such as acetic acid, succinic acid and lactic acid as major metabolites. The antibacterial, antifungal and probiotic properties of LAB strains have been widely studied (Rejiniemon et al., 2015). *L. plantarum* grow under low buffering capacity in the stomach and other complex bile salt secretions in humans and other mammals. Besides applications in the food industry, *L. plantarum* has wide applications in the pharma industry by contributing significantly to human medicine without contributing to any side effects. Recently, *L. plantarum* has been applied in medical fields for the treatment of various chronic and cardiovascular diseases such as Alzheimer's, Parkinson's, diabetes, obesity, cancer, hypertension, urogenital complications, liver disorders, etc. (Woo et al., 2014). The present study aimed to investigate the in vitro importance of *L. plantarum* related to the medical field.

## 2. Materials and methods

### 2.1. Isolation of *Lactobacillus* strains

Novel *L. plantarum* was isolated from the silage (Arasu et al., 2013). For the isolation process, the silage sample was serially diluted and spread on de Man–Rogosa (MRS) agar and incubated at 37 °C for three days. Morphological, biochemical and physiological characteristics of the strains were examined by following the reported literature. Phenotypic characteristics were studied using API 50CHB kits. The growth and fermentation pattern of the strains under various sugars were determined by following the standard method.

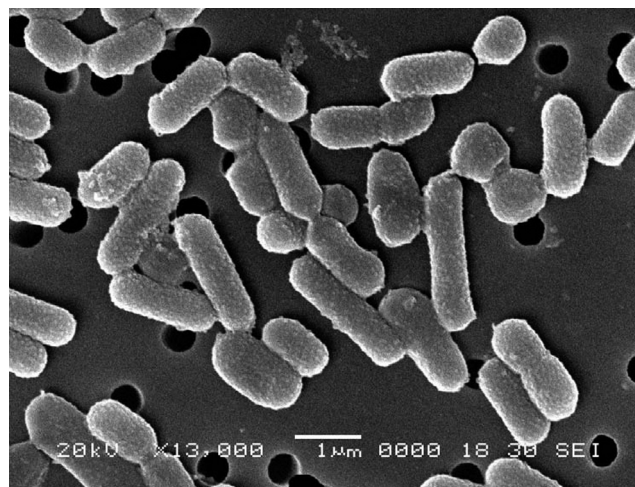
### 2.2. Importance of *L. plantarum*

The in vitro application of the *L. plantarum* was reported by many researchers. Besides the applications of these strains in our lab, the importance of these strains reported by other researchers was summarized in this report.

## 3. Results and discussion

### 3.1. Identification and characterization of *L. plantarum* strains

The fermented food is commonly known for the presence of *Lactobacillus* strains. Besides protecting the nutritional quality, *Lactobacillus* strains were used for protecting the fermented food from various fungal pathogens. Many beneficial species of *Lactobacillus* strains were derived from the fermented food and other silage samples. The routine microbiological identification methods and 16S rRNA gene amplification followed by sequencing identified the strain as *L. plantarum* (Arasu et al., 2013). The physiological and biochemical identification study showed that the strain was catalase negative, Gram positive (Fig. 1). This novel strain was tolerant to a different range of salts especially NaCl and bile salts, pH of 4.0–8.0, temperatures of 28–45 °C, and with optimum cell growth at a temperature of 37 °C and pH 7.0 respectively. Similar to the literature the identified strain



**Figure 1** Micro-morphological image of *Lactobacillus plantarum* strain.

survived various biological barriers such as low pH, lytic enzymes, and bile salts in the upper GI tract (Vijayakumar et al., 2015). Carbohydrate assimilation test concluded that the strain was able to utilize a wide range of sugars especially monosaccharide's and disaccharides respectively. Moreover, the production of extracellular enzymes such as amylase and protease was to its advantage. The above mentioned results were commonly observed in *L. plantarum* strains.

The bile salt tolerance level of the *Lactobacillus* strains was induced by the expression of proteins such as GshR4, Cfa2, Bsh1, OpuA, and AtpH (Hamon et al., 2011). Besides the tolerance level, the antagonistic properties of the novel *Lactobacillus* strains are important to prevent the spreading of the intestinal infections. In general, the probiotic *Lactobacillus* strains exhibited significant antimicrobial activity against various GI tract pathogens. The antimicrobial properties of the strains were mainly related to the secretion of the extracellular metabolites such as lactic acid, acetic acid, succinic acid, and bacteriocins.

### 3.2. Importance *L. plantarum* strains

Literature claimed that *L. plantarum* strains were widely studied for their applications in the medical field (Table 1). Especially, these strains were reported to possess the down regulation of the risk of cardiovascular diseases (Ahren et al., 2014), produce pro-inflammatory cytokines in the intestinal epithelial cells (Murofushi et al., 2015), produce varied concentrations of exopolysaccharide with anticancer property (Wang et al., 2014), reduce kidney stones (Sasikumar et al., 2014), enhance splenocytes in dendritic cells (Ku et al., 2014) and reduce the cholesterol level in the adipose tissue (Li et al., 2014). Recently, Ilavenil et al. (2015) claimed that the phenyl lactic acid recovered from *L. plantarum* promotes adipogenic activity in 3T3-L1. Interestingly, *L. plantarum* significantly induces mucosal, humoral and cellular immune responses (Shi et al., 2014) and protects against symptoms of irritable bowel syndrome (Stevenson et al., 2014). It inhibited the production of pro-inflammatory cytokines such as NF-κB and suppresses atherosclerotic plaque inflammation

**Table 1** Medical and pharmacological applications of *Lactobacillus plantarum*.

Key finding	References
This study showed that <i>L. plantarum</i> together with blueberries significantly reduced hypertension and blood pressure. Therefore, this strain might be used for the down regulation of the risk of cardiovascular diseases	Ahren et al. (2014)
The exopolysaccharides obtained from <i>L. plantarum</i> significantly decreased the production of pro-inflammatory cytokines in the intestinal epithelial cells in a RP105/MD1-dependent manner	Murofushi et al. (2015)
This study reveals that the cell bound exopolysaccharide isolated from <i>L. plantarum</i> 78010 showed significant anticancer activity	Wang et al. (2014)
This study demonstrated that the <i>L. plantarum</i> expressing oxalate decarboxylase gene significantly degrades calcium oxalate in the kidney, thus protects the kidney from stones	Sasikumar et al. (2014)
The extra cellular products of <i>L. plantarum</i> revealed anticancer effects by increased trans-epithelial electrical resistance of H4 cells and decreased the secretion of pro-inflammatory cytokines IL-6 and IL-8	Dimitrovski et al. (2014)
Oral supplementation of <i>L. plantarum</i> stimulated the expression of IL-12 and IFN- $\gamma$ in splenocytes and activates MHC class II markers, CD80 and CD 86 in dendritic cells. This study confirmed that the probiotic strain has immune-modulatory effects	Ku et al. (2014)
This study showed that the <i>L. plantarum</i> ameliorates colitis by inhibiting the TLR-4-linked NF- $\kappa$ B and MAPK signaling pathways	Jang et al. (2014)
Administration of <i>L. plantarum</i> , regulates lipid metabolism in adipose tissues by lowering cholesterol level	Li et al. (2014)
<i>Lactobacillus plantarum</i> enhances the antiproliferative activity in the vascular smooth muscle cell through the suppression of cell cycle progression and expression of cell cycle-related proteins	Lee et al. (2014)
This study states that the oral administration of <i>L. plantarum</i> expressing the hemagglutinin (HA) gene of H9N2 AIV significantly induces the mucosal, humoral and cellular immune responses. Therefore, this vaccine could be used to prevent the spreading of H9N2 avian influenza virus and also transmission of AIV	Shi et al. (2014)
The reported data confirmed that the administration of <i>L. plantarum</i> significantly reduced levels of the total cholesterol, $\gamma$ -glutamyl transpeptidase, low-density lipoprotein, glucose, homocysteine and interleukin-6 in postmenopausal women. These symptoms in postmenopausal women are an important risk factor for cardiovascular morbidity, especially stroke and coronary heart disease	Barreto et al. (2014)
The study demonstrates for the first time the protective role of <i>L. plantarum</i> on symptoms of irritable bowel syndrome	Stevenson et al. (2014)
This study confirmed that the oral administration of <i>L. plantarum</i> stimulates high levels of pro-inflammatory cytokine IL-12 and low levels of anti-inflammatory cytokine IL-10, whereas in hepatic and renal cells it induces the levels of alanine amino transferase, gamma glutamyl transferase, plasmatic triglycerides, total cholesterol, creatinine and urea concentrations	Salah et al. (2013)
Lipoteichoic acid obtained from <i>L. plantarum</i> inhibited the production of pro-inflammatory cytokines such as NF- $\kappa$ B and suppresses the atherosclerotic plaque inflammation	Kim et al. (2013)
This study confirmed that the administration of heat killed or live <i>L. plantarum</i> attenuates the symptom of Crohn's disease and ulcerative colitis	Chiu et al. (2013)
<i>L. plantarum</i> recovered from Korean traditional pickle exhibited significant antioxidant activity. This study concluded that the administration of this strain has various oxidative effects	Arasu et al. (2014)
<i>Lactobacillus plantarum</i> derived from the intestine induces the enhanced production of cytokine	Salah et al. (2013)
Regular intake of the diet with <i>Lactobacillus</i> strains reduces the body weight and white cell size of the adipose tissue	Grover et al. (2012)
This study depicts that the supplementation of <i>L. plantarum</i> prevents dermatitis by increment of type 1 helper T cell activation and regulatory T cell activation	Won et al. (2012)
<i>Lactobacillus plantarum</i> exerts anti-cancer effects in 1,2-dimethyl hydrazine (DMH)-induced colorectal cancer	Kumar et al. (2012)
Oral administration of <i>L. plantarum</i> K68 prevents the spreading of ulcer and exhibited comparatively better anti-inflammatory and immune modulatory activities by inhibiting the synthesis of factor- $\alpha$ and prostaglandin E(2) in macrophage	Liu et al. (2011)
This study documented that the plantaricin A produced by <i>L. plantarum</i> stimulates in vitro proliferation and migration of human keratinocytes	Pinto et al. (2011)
This study concluded that the <i>L. plantarum</i> strains are mainly involved in the T-cell differentiation, thereby improving the immune responses toward antigens	Vissers et al. (2010)
Tight junction formation in the intestine is stimulated by in-vivo administration of <i>L. plantarum</i>	Anderson et al. (2010)
Bacterial infection causes a serious problem in curing wounds especially in the ulcer stage. In this report, external application of <i>L. plantarum</i> on the ulcer patient cures the wounds of the diabetic patient. The investigations concluded that levels of polymorphonuclear, apoptotic and necrotic cells were completely decreased	Pera et al. (2010)
Oral administrations of <i>L. plantarum</i> cure obstructive jaundice and protect the liver from different barriers	Zhang et al. (2010)
<i>Lactobacillus plantarum</i> coated with proteins and polysaccharides exhibited interesting hypocholesterolemic effects. This combination speeds up the degradation of hepatic cholesterol into bile acids	Jeun et al. (2010)
<i>Lactobacillus plantarum</i> may improve human colon cancer. This is the first report about the pharmacological application of probiotic bacteria in the treatment of colon cancer and the mechanism of intestinal epithelial cells in immune responses	Paolillo et al. (2009)

(Kim et al., 2013) and induces the enhanced production of cytokines in the human intestine (Salah et al., 2012). On the other hand, *L. plantarum* strains are mainly involved in the T-cell differentiation thereby improving immune responses toward antigens (Visser et al., 2010) and preventing dermatitis by increment of type 1 helper T cell activation and regulatory T cell activation (Won et al., 2012).

#### 4. Conclusion

In conclusion, among the lactic acid bacteria, *L. plantarum* plays an important role in medical applications. The summarized recent report related to *L. plantarum* would be useful to the pharmaceutical industry for the preparation of medical formulations without side effects.

#### Acknowledgement

The Project was full financially supported by King Saud University, through Vice Deanship of Research Chairs.

#### References

- Ahren, I.L., Jie, X., Önnings, G., Olsson, C., Ahrné, S., Molin, G., 2014. Antihypertensive activity of blueberries fermented by *Lactobacillus plantarum* DSM 15313 and effects on the gut microbiota in healthy rats. *Clin. Nutr.*, 1–14
- Anderson, R.C., Cookson, A.L., McNabb, W.C., Park, Z., McCann, M.J., Kelly, W.J., Roy, N.C., 2010. *Lactobacillus plantarum* MB452 enhances the function of the intestinal barrier by increasing the expression levels of genes involved in tight junction formation. *BMC Microbiol.* 10, 316.
- Arasu, M.V., Jung, M.W., Ilavenil, S., Jane, M., Kim, D.H., Lee, K.D., Park, H.S., Hur, T.Y., Choi, G.J., Lim, Y.C., Al-Dhabi, N.A., Choi, K.C., 2013. Isolation and characterization of antifungal compound from *Lactobacillus plantarum* KCC-10 from forage silage with potential beneficial properties. *J. Appl. Microbiol.* 115, 1172–1185.
- Arasu, M.V., Kim, D.H., Kim, P.I., Jung, M.W., Ilavenil, S., Jane, M., Lee, K.D., Al-Dhabi, N.A., Choi, K.C., 2014. In vitro antifungal, probiotic and antioxidant properties of novel *Lactobacillus plantarum* K46 isolated from fermented sesame leaf. *Ann. Microbiol.* 64 (3), 1333–1346.
- Barreto, F.M., Simão, A.N.C., Morimoto, H.K., Lozovoy, M.A.I.B., Dichi, I., Miglioranza, L.H.S., 2014. Beneficial effects of *Lactobacillus plantarum* on glycemia and homocysteine levels in postmenopausal women with metabolic syndrome. *Nutrition* 30 (7–8), 939–942.
- Chiu, Y.-H., Lu, Y.-C., Ou, C.-C., Lin, S.-L., Tsai, C.-C., Huang, C.-T., Lin, M.-Y., 2013. *Lactobacillus plantarum* MYL26 induces endotoxin tolerance phenotype in Caco-2 cells. *BMC Microbiol.* 13, 190.
- Dimitrovski, D., Cencič, A., Winkelhausen, E., Langerholc, T., 2014. *Lactobacillus plantarum* extracellular metabolites: in vitro assessment of probiotic effects on normal and cancerogenic human cells. *Int. Microbiol. J.* 39 (2), 293–300.
- Grover, S., Rashmi, H.M., Srivastava, A.K., Batish, V.K., 2012. Probiotics for human health – new innovations and emerging trends. *Gut Pathog.* 4, 15.
- Hamon, E., Horvatovich, P., Izquierdo, E., Bringel, F., Marchioni, E., Aoudé-Werner, D., Ennahar, S., 2011. Comparative proteomic analysis of *Lactobacillus plantarum* for the identification of key proteins in bile tolerance. *BMC Microbiol.* 29 (11), 63.
- Ilavenil, S., Kim, D.H., Arasu, M.V., Srigopalram, S., Sivanesan, R., Choi, K.C., 2015. Phenyllactic acid from *Lactobacillus plantarum* promotes adipogenic activity in 3T3-L1 adipocyte via up-regulation of PPAR- $\gamma$ 2. *Molecules* 20 (8), 15359–15373. <http://dx.doi.org/10.3390/molecules200815359>.
- Jang, S.-E., Han, M.J., Kim, S.-Y., Kim, D.-H., 2014. *Lactobacillus plantarum* CLP-0611 ameliorates colitis in mice by polarizing M1 to M2-like macrophages. *Int. Immunopharmacol.* 21 (1), 186–192.
- Jeun, J., Kim, S., Cho, S.Y., Jun, H.J., Park, H.J., Seo, J.G., Chung, M.J., Lee, S.J., 2010. Hypocholesterolemic effects of *Lactobacillus plantarum* KCTC3928 by increased bile acid excretion in C57BL/6 mice. *Nutrition* 26 (3), 321–330.
- Kim, J.Y., Kim, H., Jung, B.J., Kim, N.R., Park, J.E., Chung, D.K., 2013. Lipoteichoic acid isolated from *Lactobacillus plantarum* suppresses LPS-mediated atherosclerotic plaque inflammation. *Mol. Cells* 35 (2), 115–124.
- Ku, H.-K., Lee, H., Choi, I.D., Ra, J.-H., Kim, T.-Y., Jeong, J.-W., Kim, S.-H., Sim, J.-H., Ahn, Y.-T., 2014. Immuno-stimulatory effect of *Lactobacillus plantarum* HY7712 via toll-like receptor 2 signaling pathway. *Cytokine* 70 (1), 52.
- Kumar, R.S., Kanmani, P., Yuvaraj, N., Paari, K.A., Pattukumar, V., Thirunavukkarasu, C., Arul, V., 2012. *Lactobacillus plantarum* ASI isolated from south Indian fermented food kallappam suppress 1,2-dimethyl hydrazine (DMH)-induced colorectal cancer in male wistar rats. *Appl. Biochem. Biotechnol.* 166 (3), 620–631.
- Lee, J.-J., Kwon, H., Lee, J.-H., Kim, D.-G., Jung, S.-H., Ma, J.Y., 2014. Fermented soshiho-tang with *Lactobacillus plantarum* enhances the antiproliferative activity in vascular smooth muscle cell. *BMC Complement. Altern. Med.* 14, 78.
- Li, C., Nie, S.-P., Ding, Q., Zhu, K.-X., Wang, Z.-J., Xiong, T., Gong, J., Xie, M.-Y., 2014. Cholesterol-lowering effect of *Lactobacillus plantarum* NCU116 in a hyperlipidaemic rat model. *J. Funct. Food* 8, 340–347.
- Liu, Y.W., Su, Y.W., Ong, W.K., Cheng, T.H., Tsai, Y.C., 2011. Oral administration of *Lactobacillus plantarum* K68 ameliorates DSS-induced ulcerative colitis in BALB/c mice via the anti-inflammatory and immunomodulatory activities. *Int. Immunopharmacol.* 11 (12), 2159–2166.
- Murofushi, Y., Villena, J., Morie, K., Kanmani, P., Tohno, M., Shimazu, T., Aso, H., Suda, Y., Hashiguchi, K., Saito, T., Kitazawa, H., 2015. The toll-like receptor family protein RP105/MD1 complex is involved in the immunoregulatory effect of exopolysaccharides from *Lactobacillus plantarum* N140. *Mol. Immunol.* 64 (1), 63–75.
- Paolillo, R., Carratelli, C.R., Sorrentino, S., Mazzola, N., Rizzo, A., 2009. Immunomodulatory effects of *Lactobacillus plantarum* on human colon cancer cells. *Int. Immunopharmacol.* 9 (11), 1265–1271.
- Pera, M.C., Rachid, M.M., Gobbato, N.M., Martinez, M.A.H., Valdez, J.C., 2010. Interleukin-8 production by polymorphonuclear leukocytes from patients with chronic infected leg ulcers treated with *Lactobacillus plantarum*. *Clin. Microbiol. Infect.* 16 (3), 281–286.
- Pinto, D., Marzani, B., Minervini, F., Calasso, M., Giuliani, G., Gobetti, M., Angelis, M.D., 2011. Plantaricin A synthesized by *Lactobacillus plantarum* induces in vitro proliferation and migration of human keratinocytes and increases the expression of TGF- $\beta$ 1, FGF7, VEGF-A and IL-8 genes. *Peptides* 32 (9), 1815–1824.
- Rejiniemon, T.S., Hussain, R.R., Rajamani, B., 2015. In-vitro functional properties of *Lactobacillus plantarum* iso-lated from fermented ragi malt. *South Ind. J. Biol. Sci.* 1, 15–23.
- Salah, R.B., Trabelsi, I., Mansour, R.B., Lassoued, S., Chouayekh, H., Bejar, S.A., 2012. New *Lactobacillus plantarum* strain, TN8, from the gastro intestinal tract of poultry induces high cytokine production. *Anaerobe* 18 (4), 436–444.
- Salah, R.B., Trabelsi, I., Hamden, K., Chouayekh, H., Bejar, S., 2013. *Lactobacillus plantarum* TN8 exhibits protective effects on lipid, hepatic and renal profiles in obese rat. *Anaerobe* 23, 55–61.

- Sanchez, B., Urdaci, M.C., Margolles, A., 2010. Extracellular proteins secreted by probiotic bacteria as mediators of effects that promote mucosa–bacteria interactions. *Microbiology* 156, 3232–3242.
- Sasikumar, P., Gomathi, S., Anbazhagan, K., Abhishek, A., Paul, E., Vasudevan, V., Sasikumar, S., Selvam, G.S., 2014. Recombinant *Lactobacillus plantarum* expressing and secreting heterologous oxalate decarboxylase prevents renal calcium oxalate stone deposition in experimental rats. *J. Biomed. Sci.* 21, 86.
- Shi, S.-H., Yang, W.-T., Yang, G.-L., Cong, Y.-L., Huang, H.-B., Wang, Q., Cai, R.-P., Ye, L.-P., Hu, J.-T., Zhou, J.-Y., Wang, C.-F., Li, Y., 2014. Immunoprotection against influenza virus H9N2 by the oral administration of recombinant *Lactobacillus plantarum* NC8 expressing hemagglutinin in BALB/c mice. *Virology* 464–465, 166–176.
- Stevenson, C., Blaauw, R., Fredericks, E., Visser, J., Roux, S., 2014. PP137-SUN: randomized clinical trial: effect of *Lactobacillus plantarum* 299V on symptoms of irritable bowel syndrome. *Clin. Nutr.* 33 (1), S71.
- Vijayakumar, M., Ilavenil, S., Kim, D.H., Arasu, M.V., Priya, K., Choi, K.C., 2015. In-vitro assessment of the probiotic potential of *Lactobacillus plantarum* KCC-24 isolated from Italian rye-grass (*Lolium multiflorum*) forage. *Anaerobe* 32, 90–97.
- Visser, Y.M., Snel, J., Zuurendonk, P.F., Smit, B.A., Wichers, H.J., Savelkoul, H.F.J., 2010. Differential effects of *Lactobacillus acidophilus* and *Lactobacillus plantarum* strains on cytokine induction in human peripheral blood mononuclear cells. *FEMS Immun. Med. Microbiol.* 59 (1), 60–70.
- Wang, K., Li, W., Rui, X., Chen, X., Jiang, M., Dong, M., 2014. Characterization of a novel exopolysaccharide with antitumor activity from *Lactobacillus plantarum* 70810. *Int. J. Biol. Macromol.* 63, 133–139.
- Won, T.J., Kim, B., Lee, Y., Bang, J.S., Oh, E.S., Yoo, J.-S., Hyung, K.E., Yoon, J., Hwang, S., Park, E.S., Park, S.-Y., Hwang, K.W., 2012. Therapeutic potential of *Lactobacillus plantarum* CJLP133 for house-dust mite-induced dermatitis in NC/Nga mice. *Cell. Immunol.* 277 (1–2), 49–57.
- Woo, J.-Y., Gu, W., Kim, K.-A., Jang, S.-E., Han, M.J., Kim, D.-H., 2014. *Lactobacillus pentosus* var. *plantarum* C29 ameliorates memory impairment and inflammaging in a D-galactose-induced accelerated aging mouse model. *Anaerobe* 27, 22–26.
- Zhang, M., Wang, X.Q., Zhou, Y.K., Ma, Y.L., Shen, T.Y., Chen, H.Q., Chu, Z.X., Qin, H.L., 2010. Effects of oral *Lactobacillus plantarum* on hepatocyte tight junction structure and function in rats with obstructive jaundice. *Mol. Biol. Rep.* 37 (6), 2989–2999.