

Neuroprotection by flavonoids

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Abstract

The high morbidity, high socioeconomic costs and lack of specific treatments are key factors that define the relevance of brain pathology for human health and the importance of research on neuronal protective agents. Epidemiological studies have shown beneficial effects of flavonoids on arteriosclerosis-related pathology in general and neurodegeneration in particular. Flavonoids can protect the brain by their ability to modulate intracellular signals promoting cellular survival. Quercetin and structurally related flavonoids (myricetin, fisetin, luteolin) showed a marked cytoprotective capacity in *in vitro* experimental conditions in models of predominantly apoptotic death such as that induced by medium concentrations (200 μ M) of H₂O₂ added to PC12 cells in culture. Nevertheless, quercetin did not protect substantia nigra neurons *in vivo* from an oxidative insult (6-hydroxydopamine), probably due to difficulties in crossing the blood-brain barrier. On the other hand, treatment of permanent focal ischemia with a lecithin/quercetin preparation decreased lesion volume, showing that preparations that help to cross the blood-brain barrier may be critical for the expression of the effects of flavonoids on the brain. The hypothesis is advanced that a group of quercetin-related flavonoids could become lead molecules for the development of neuroprotective compounds with multitarget anti-ischemic effects.

Key words

- Flavonoids
- Quercetin
- Focal ischemia
- Neuroprotection

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Brain vascular pathology and oxidative stress

It is known that brain pathology in the form of cerebrovascular and neurodegenerative disease is a leading cause of death all over the world, with an incidence of about 2/1000 and an 8% total death rate (1-3). Moreover, stroke and dementia are a source of high individual and family suffering mainly because of the lack of efficient therapeutic alternatives. The latter motivates research

efforts to identify the mechanisms of neuronal death and to discover new compounds to control them.

Neuronal death in stroke is a complex event involving failure of metabolic processes, excitotoxicity, loss of calcium homeostasis and oxidative stress, among other factors (4). During ischemic stroke, a decrease in metabolic energy in the form of ATP affecting membrane ionic pumps leads to an increase in intracellular Ca²⁺ and Na⁺ concentrations and to increased glutamate

release (5). The massive Ca^{2+} entry activates enzymes such as proteases, oxidases, phospholipases and endonucleases (6) that can hydrolyze the DNA molecule and destroy the cytoskeleton (7). Phospholipase A_2 activation favors the metabolism of arachidonic acid through lipoxygenases and eicosanoids in turn activate lipid peroxidation. Increased intracellular Ca^{2+} also activates protein kinase C that can modify the function of many ion channels (8) (Figure 1).

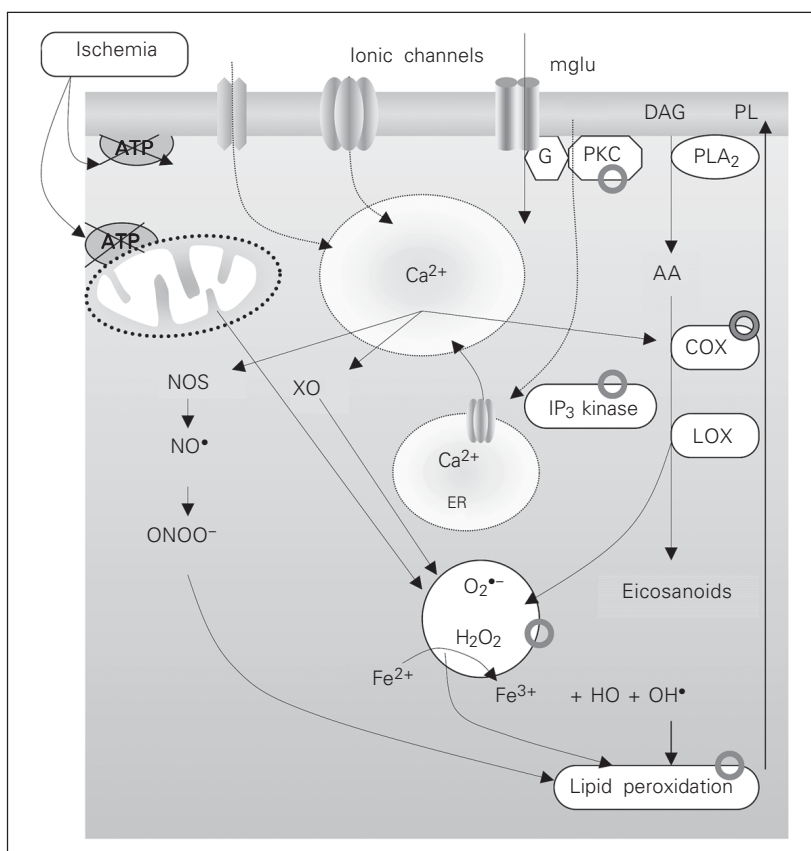


Figure 1. Schematic drawing of the ischemic process at the cellular level. The initial mitochondrial metabolic failure (ATP) leads to disruption of ionic pump functioning at the membrane level and massive neurotransmitter and glutamate release which in turn increase the Ca^{2+} entry. There is an activation of enzymes (XO, NOS, COX, LOX kinases). Reactive oxygen and nitrogen species production (such as $\text{O}_2^{\cdot-}$, NO^{\cdot} and H_2O_2) is part of this process, generating lipid peroxidation and nuclear DNA damage. Flavonoids act by inhibiting several steps in this process: XO, lipid peroxidation, IP_3 kinase, LOX, PKC and scavenging free radicals (marked with circles in the figure). AA = arachidonic acid; COX = cyclooxygenase; DAG = diacylglycerol; ER = endoplasmic reticulum; G = G protein; IP_3 = phosphatidylinositol-3; LOX = lipoxygenase; mglu = metabotropic glutamate receptors; NO = nitric oxide; NOS = nitric oxide synthase; PKC = protein kinase C; PL = phospholipids; PLA_2 = phospholipase A_2 ; XO = xanthine oxidase.

Several of these activated intracellular metabolic events lead to the generation of oxygen free radicals, which overcome the antioxidant defenses and provoke oxidative stress (9). Oxidative stress, in turn, provokes changes in macromolecules and in lipid membranes, generating a vicious cycle of more oxidation and more oxidative damage. Self-maintained oxidative reactions have been identified in arteriosclerosis, the main pathological condition leading to stroke (10,11).

Oxidative stress and natural antioxidants

In spite of the many lines of evidence linking oxidative stress to clinical symptoms related to arteriosclerosis in myocardial infarction and stroke (10,11), treatment with external antioxidants to regain oxidative equilibrium and to control the evolution of disease has provided controversial results. Some clinical trials with a recognized antioxidant like vitamin E did not show beneficial effects on the treatment of cardiovascular pathology risk (12) and doses higher than 500 mg of ascorbic acid as well as β -carotene seem to have negative effects (13,14).

On the other hand, several studies have shown that fruits, red wine, vegetables and some plants increase the total antioxidant capacity of blood (15,16) and the antioxidant actions of polyphenol metabolites have been suggested to explain these beneficial effects, particularly for the Mediterranean diet (17-19). The most important polyphenol compounds are the flavonoids, which are abundant components of the human diet (8,20,21). Quercetin, a key representative flavonoid molecule of the group, is present widely in vegetables and fruits, with a daily intake of up to 25 mg/day in a normal human diet (22). Other effects such as antitumoral, antithrombotic, anti-inflammatory and antiapoptotic ones, as well as effects inhibiting platelet aggregation and the growth of certain types

of cancer, have been described for quercetin and other flavonoids (23-28).

Quercetin actively participates in intracellular signaling, inhibiting phosphatidylinositol-3 kinase, protein kinase C, xanthine oxidase and NADPH diaphorase (8,29-31). Nevertheless, in spite of this multiplicity of actions, the cardiovascular and/or neuroprotective effects of flavonoids and quercetin are mainly explained by their antioxidant capacity and their ability to scavenge free radicals (18).

Natural antioxidants and cytoprotection

Nature has been a continuous source of pharmacologically active molecules and medicinal herbs have been used by countless human generations. Nevertheless, surprisingly few plant extracts have been demonstrated to be neuroactive. Kava and *Ginkgo biloba* extracts have been shown to have neuroprotective actions based on experimental evidence (17,32-35). However, adverse effects have also been reported for kava and *Ginkgo biloba* (36,37).

Among the very few investigators of the effects of isolated flavonoids on the brain *in vivo*, Shutenko et al. (38) characterized changes in brain nitric oxide levels in a model of global ischemia with reperfusion in the presence of quercetin, attributing the observed changes to the scavenger action of the flavonoid. Another report has described the beneficial effects of quercetin on endotoxic shock (39) and the authors explained these effects by lipoperoxidation inhibition and increases in glutathione peroxidase activity.

Direct scavenging of reactive oxygen species is one of the many antioxidant actions required to restore oxidative equilibrium once it is lost in different pathologies. The hypothesis that restoring redox equilibrium through activation of intracellular signals is also an important step of the

antioxidation process is gaining increasing support (40). It is likely that the trapping of free radical excess could restore redox equilibrium in the initial states of cellular oxidative stress. In massive cellular insults like ischemia, involving metabolic failure, loss of Ca^{2+} homeostasis and excitotoxicity, scavenger activity or one-target antioxidant mechanisms (NMDA receptor blockers, chain-breaking vitamin E or pure scavenger molecules such as boldine) may fail to protect cells from free radical damage.

Accordingly, in experimental conditions, the capacity of a given molecule to block the multiple sources of oxidative signals in situations like ischemia would be better assessed by its effectiveness in increasing cell survival. Hence, it could be hypothesized that the cytoprotective capacity of a given antioxidant would be critical to define a putative neuroprotective therapeutic activity. Protection of cells in culture against diverse insults (glutamate, AB peptide, and others) has proved to be a useful approach (18).

The oxidative insult with hydrogen peroxide (H_2O_2) has been widely used to assess cytoprotection, mainly in PC12 cells in culture (41-43). H_2O_2 offers the unique possibility of a graded action regulating the extent and severity of cell death by the selection of particular points of the H_2O_2 -cell interaction. Thus, it has been shown that exposure of PC12 cells to 200 μM H_2O_2 for 30 min resulted in 50% cell viability. Cell death was accompanied by DNA damage without lactic dehydrogenase release, suggesting a "non-necrotic" type of cell death (44). Under closely similar experimental conditions, H_2O_2 -induced cell death in PC12 was characterized as apoptotic (41). When various potent antioxidants (vitamin E, trolox, boldine, quercetin) were studied for their capacity to increase cell survival in the H_2O_2 -induced PC12 cell death, only quercetin or structurally similar flavonoids protected PC12 cells from the oxidative insult. Of several flavonoids structurally related to quercetin - myricetin,

Figure 2. PC12 cell viability after a 30-min H_2O_2 insult in the presence of diverse flavonoids. * $P < 0.01$ for H_2O_2 plus flavonoid treatment compared to only H_2O_2 treatment (ANOVA and multiple comparison Tukey test). The horizontal line behind the bars represents the control's mean and the gray area the standard deviation of control experiments.

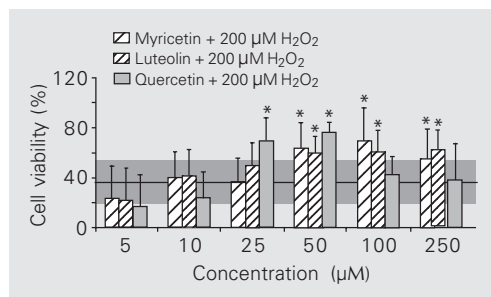
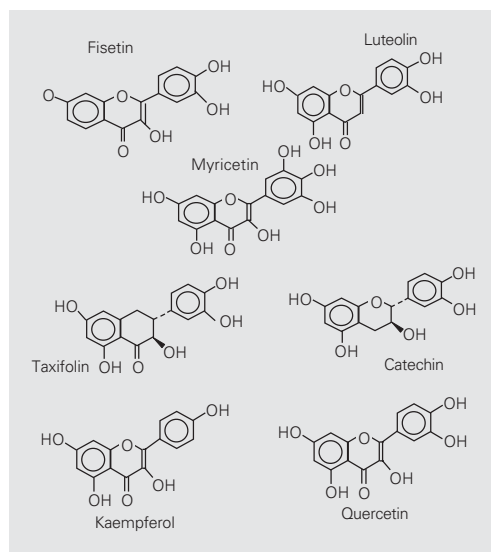


Figure 3. Structures of the flavonoids studied.



kaempferol, taxifolin, catechin, luteolin and fisetin - only fisetin, luteolin and myricetin increased cell survival (45; Figure 2).

The common structural features of the molecules that increased survival were a 3',4'-dihydroxy catechol structure in the B ring, coplanarity of the molecule and the presence of 2,3 unsaturation together with an oxo function at position 4 in the C ring. This structural profile is the one recognized as important also for antioxidant scavenger activity (19) (Figure 3).

The fact that quercetin and structurally related molecules increased cell survival in an oxidative stress model where scavenging antioxidants (vitamin E, boldine) failed to protect cells from the oxidative insult suggests that these molecules have a specific cell survival-increasing activity in addition to their scavenger activity. This cytoprotective capacity of flavonoids may be linked to their ability to activate intracellular molecules (kinases, phosphatases, gene promoters), which in turn activate components of intracellular cascades, promoting the expression of survival signals. This would be especially true for cellular protection in cases in which apoptosis is an important component of cell death.

Figure 4. Inhibition of spontaneous lipoperoxidation in isolated rat brain membranes by different antioxidants. Data are reported as IC_{50} (concentrations required to obtain 50% inhibition).

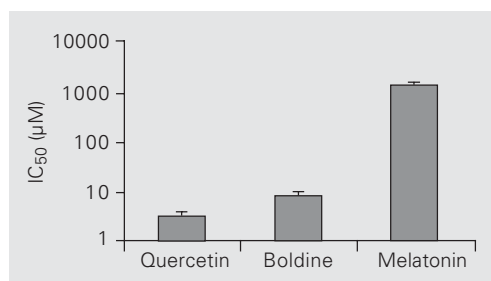
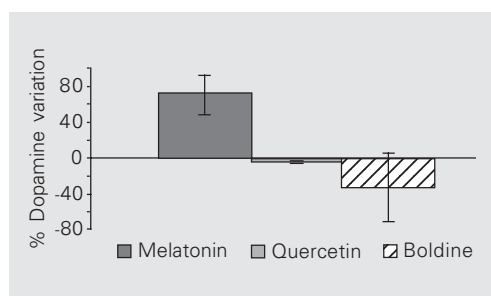


Figure 5. Dopamine tissue levels in the corpus striatum of rats injected with 6-hydroxydopamine (6-OHDA) into the substantia nigra. Data are reported as percent of the contralateral, nonlesioned side. Rats were treated *ip* with 5 mg/kg melatonin, 30 mg/kg quercetin, and 15 mg/kg boldine 30 min before the 6-OHDA lesion.



Flavonoids in experimental brain pathology

The antioxidant profile of flavonoids would be a strong basis for a neuron-protective activity in the brain. Their general bioavailability and particularly their presence in the brain *in vivo* appear to play an important role in the expression of the neuroprotective capacity of flavonoids. It is accepted that metabolic transformations (glucuronidation, methylation, etc.) are the rule and that a very small amount of a given intake of flavonoids are free as aglycones in blood (46,47). For the specific aim of neuroprotection, the blood-brain barrier appears to be an added obstacle to flavonoids reaching the brain.

Dajas and co-workers (48) administered

recognized antioxidants, including quercetin, in an experimental model of Parkinson to test the ability of flavonoids to cross the blood-brain barrier and act on the brain. Microinjection of 6-hydroxydopamine (6-OHDA) induces death of dopaminergic neurons in the substantia nigra with a concomitant loss of terminals in the corpus striatum. Assessment of dopamine in the latter after a 6-OHDA insult gives an idea of the extent of the lesion. In agreement with the prevalent hypothesis regarding the cause and/or progression of Parkinson's disease, 6-OHDA lesion is produced by oxidative stress (49). In the study of Dajas et al. (48), natural compounds such as boldine, quercetin and melatonin, with marked antioxidant potency and different mechanisms of action, were tested by being injected intraperitoneally in saline solution before 6-OHDA lesion. An interesting aspect was the comparison of the *in vitro* antioxidant capacity of these natural antioxidants with their neuroprotective activity in the experimental model of Parkinson's disease: boldine and quercetin had the highest antioxidant potency in the antilipoperoxidation assay, followed by melatonin (Figure 4). When the capacity to increase neuronal survival was tested, melatonin, the weaker antioxidant, reversed the dopamine levels in the striatum while quercetin and boldine did not (Figure 5). Accordingly, and in spite of its antioxidant potency *in vitro* and cytoprotective actions in cell cultures, quercetin did not protect substantia nigra neurons *in vivo* (50). These results would show that flavonoids like quercetin cross the blood-brain barrier poorly. The surprisingly few papers reporting effects of flavonoids in the brain *in vivo*, cited above, appear to confirm this fact. Most of the reports of neuroprotection by natural compounds from plants refer to complex extracts like those of *Ginkgo biloba* and not to single compounds.

In a later study, Dajas et al. (51) increased the possibility of quercetin crossing the blood-brain barrier by mixing it with lecithin, gen-

erating a liposomal preparation. This preparation is a recognized way of transporting molecules in the body, increasing the time of interaction of a given molecule with its target (46). The authors utilized the middle cerebral artery occlusion model in the rat (52). A single intraperitoneal dose of a lecithin/quercetin preparation (30 mg/kg) was administered 30 min after artery occlusion and rats were sacrificed 24 h after occlusion. Assessed by means of a computer (53), the volume of the ischemic lesion decreased 56% after treatment (Figures 6 and 7).

While the ischemic area extended over the striatum and parietal cortex (Figure 6), the recovery was more marked in the striatum. The decrease of the lesion area and volume corresponded to decreased edema and to neuronal survival as assessed by histology.

The decrease in lesion volume obtained with lecithin/quercetin was similar to the lesion improvement observed after excitotoxicity antagonism (54) or the use of cal-

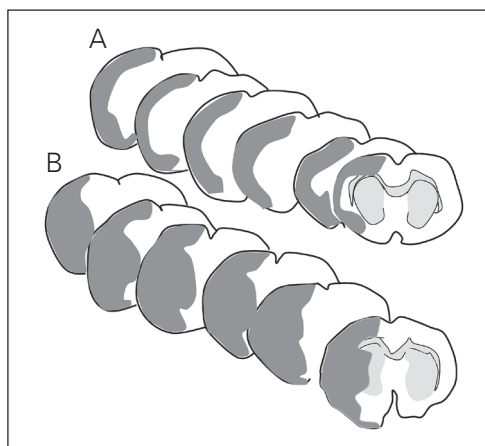


Figure 6. Computer-generated schematic drawings of representative frontal slices of 2,3,5 triphenyl tetrazolium chloride-stained coronal brain sections obtained 24 h after permanent middle cerebral artery occlusion in rats receiving an *ip* injection of either lecithin/quercetin preparation (A) or saline (B), 30 min after occlusion. The recovery of the ischemic areas is shown.

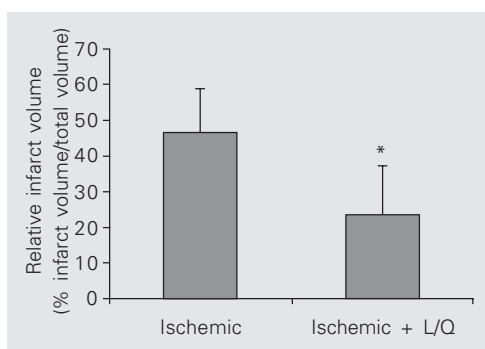


Figure 7. Relative infarct volume 24 h after permanent middle cerebral artery occlusion. Rats received an *ip* injection of saline (N = 5, ischemic in figure) or lecithin/quercetin (N = 7, ischemic + L/Q in figure) 30 min after middle cerebral artery occlusion. Values are reported as means \pm SD. *P < 0.05 compared to the saline group (ANOVA and Kruskal-Wallis test).

cium channel blockers (55). Accordingly, in contrast with the lack of effects in the oxidative lesion of experimental parkinsonism, the work on permanent ischemia showed a neuroprotective action of lecithin/quercetin in the brain, probably demonstrating the importance of the way of administering quercetin to assure the crossing of the blood-brain barrier in sufficient quantities to be beneficial against the oxidative damage. These preliminary results with quercetin would be showing a putative relationship, in a group of related flavonoids, between the cytoprotective potency in apoptotic models of cell death and central neuroprotective activity *in vivo*.

Up to now, neuroprotective strategies in ischemia that have focused on the development of molecules targeting one mechanism at a time have not proven to be successful in clinical trials. The multiple cell effects of flavonoid indicate that several targets could be reached with only one molecule by administering the flavonoid in a preparation that could cross the blood-brain barrier.

The capacity of flavonoids to inhibit the action of several enzymes should activate

survival signals exerting a net indirect antioxidant effect in addition to the direct scavenging of reactive oxygen species. Additionally, it is important to mention that the effects of quercetin and flavonoids are also exerted at the level of glia and vessels in the brain. At the microvessel level, antioxidation and anti-inflammation would be added to vasodilatation, improving blood flow and counteracting the ischemic process (56).

Although the exact explanation of the mechanisms of action of flavonoids on the brain has just started to be addressed and a wide diversity of questions remain open, flavonoids are likely to become leading compounds for the development of a new generation of molecules clinically effective in human brain ischemia.

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References

- Kolominsky-Rabas PL, Sarti C, Heuschmann PU, Graf C, Siemonsen S, Neundoerfer B, Katalinic A, Lang E, Gassmann KG & von Stockert TR (1998). A prospective community-based study of stroke in Germany. The Erlangen Stroke Project (ESPro): incidence and case fatality at 1, 3, and 12 months. *Stroke*, 29: 2501-2506.
- Samsa G, Bian J, Lipscomb J & Matchar D (1999). Epidemiology of recurrent cerebral infarction: a medicare claims-based comparison of first and recurrent strokes on 2-year survival and cost. *Stroke*, 30: 338-349.
- Leppälä J, Virtamo J, Fogelholm R, Albanes D & Heinonen O (1999). Different risk factors for different stroke subtypes: association of blood pressure, cholesterol, and antioxidants. *Stroke*, 30: 2535-2540.
- Alexi T, Borlongan C, Faull C, Williams C, Clark R, Gluckman P & Hughes P (2000). Neuroprotective strategies for basal ganglia degeneration: Parkinson's and Huntington's diseases. *Progress in Neurobiology*, 60: 409-470.
- Nicholls D & Attwell D (1990). The release and uptake of excitatory amino acids. *Trends in Pharmacological Sciences*, 11: 462-468.
- Nicotera P & Lipton S (1999). Excitotoxins in neuronal apoptosis and necrosis. *Journal of Cerebral Blood Flow and Metabolism*, 19: 583-591.
- Welch KMA, Kaplan LR, Reis DJ, Siesjo BK & Weir B (1997). *Primer on Cerebrovascular Diseases*. Academic Press, San Diego, CA, USA.
- Picq M, Dubois M, Munri-Silem Y, Prugent AF & Pacheco H (1989). Flavonoid modulation of protein kinase C activation. *Life Sciences*, 44: 1563-1571.
- Dugan LL & Choi DW (1999). Hypoxic-ischemic brain injury and oxidative stress. In: Siegel GJ, Agranoff BW, Albers RW, Fischer SK & Uhler MD (Editors), *Basic Neurochemistry*. Lippincott-Raven Publishers, Philadelphia, New York, chapter 34.
- Landemmer U & Harrison DG (2001). Oxidative stress and vascular damage in hypertension. *Coronary Artery Disease*, 12: 455-461.
- Landemmer U & Harrison DG (2001). Oxidant stress as a marker for cardiovascular events: Ox marks the spot. *Circulation*, 104: 2638-2640.
- Emmert D & Kirchner L (1999). The role of vitamin E in the prevention of heart disease. *Archives of Family Medicine*, 8: 537-542.
- Colqhoun D (2002). Nutraceuticals: vitamins and other nutrients in coronary heart disease. *Current Opinion in Lipidology*, 12: 639-646.
- Duthie G & Crozier A (2000). Plant derived phenolic antioxidants. *Current Opinion in Clinical Nutrition and Metabolic Care*, 3: 457-471.

15. Riso P, Pindler A, Santangelo A & Porrini M (1999). Does tomato consumption effectively increase the resistance of lymphocyte DNA to oxidative damage? *American Journal of Clinical Nutrition*, 69: 712-718.
16. Sung H, Nah J, Chun S, Park H, Yang SE & Min WK (2000). *In vivo* antioxidant effect of green tea. *European Journal of Clinical Nutrition*, 54: 527-529.
17. Bastianetto S & Quirion R (2002). Natural extracts as possible protective agents of brain aging. *Neurobiology of Aging*, 23: 891-897.
18. Esposito E, Rotilio D, Di Matteo V, Di Giulio C, Cacchio M & Algeri S (2002). A review of specific dietary antioxidants and the effects on biochemical mechanisms related to neurodegenerative processes. *Neurobiology of Aging*, 23: 719-735.
19. Rice Evans C (2001). Flavonoid antioxidants. *Current Medicinal Chemistry*, 8: 797-807.
20. Matter WF, Brown RF & Vlahos CJ (1992). The inhibition of phosphatidylinositol 3-kinase by quercetin and analogs. *Biochemical and Biophysical Research Communications*, 186: 624-631.
21. Chang WS, Lee YJ, Lu FJ & Chiang HC (1993). Inhibitory effects of flavonoids on xanthine oxidase. *Anticancer Research*, 13: 2165-2170.
22. NTP (1992). Technical report on the toxicology and carcinogenesis studies of quercetin in F344/N rats. NIH Publication No. 91-3140. U.S. Department of Health and Human Services, Public Health Service, Research Triangle Park, NC, USA.
23. Fujiki H, Horiuchi T & Yamashita K (1986). Inhibition of tumor promotion by flavonoids. In: Cody V, Middleton E & Harborne JB (Editors), *Plant Flavonoids in Biology and Medicine: Biochemical, Pharmacological and Structure-Activity Relationships*. Alan R. Liss, New York, 429-440.
24. Beretz A, Stierle A, Anton R & Cazenave JP (1981). Role of cyclic AMP in the inhibition of human platelet aggregation by quercetin, a flavonoid that potentiates the effect of prostacyclin. *Biochemical Pharmacology*, 31: 3597-3600.
25. Gryglewski RJ, Korbut R, Robak J & Swies J (1987). On the mechanism of antithrombotic action of flavonoids. *Biochemical Pharmacology*, 36: 317-322.
26. Scambia G, Ranalletti FO & Benederri Pacini P (1990). Inhibitory effect of quercetin on OVCA 433 cells and presence of type II oestrogen binding sites in primary ovarian tumors and cultured cells. *British Journal of Cancer*, 62: 942-947.
27. Yoshida M, Sakai T & Hosokawa N (1992). The effect of quercetin on cell cycle progression and growth of human gastric cancer cells. *FEBS Letters*, 260: 10-13.
28. Juurlink BH & Paterson PG (1998). Review of oxidative stress in brain and spinal cord injury: suggestions for pharmacological and nutritional management strategies. *Journal of Spinal Cord Medicine*, 21: 309-334.
29. Agullo G, Gamet-Payastre L, Manenti S, Viala C, Remesy C, Chap H & Payastre B (1997). Relationship between flavonoid structure and inhibition of phosphatidylinositol 3-kinase: a comparison with tyrosine kinase and protein kinase C inhibition. *Biochemical Pharmacology*, 53: 1649-1657.
30. Gschwendt M, Horn F, Kittstein W & Marks F (1983). Inhibition of the calcium and phospholipid dependent protein kinase activity from mouse brain cytosol by quercetin. *Biochemical and Biophysical Research Communications*, 117: 444-447.
31. Tamura M, Kagawa S, Tsuruo Y, Ishimura K & Morita K (1994). Effects of flavonoid compounds on the activity of NADPH diaphorase prepared from the mouse brain. *Japanese Journal of Pharmacology*, 66: 371-373.
32. Assemi M (2001). Herbs affecting the central nervous system: ginkgo, kava, St. John's wort and valerian. *Clinical Obstetrics and Gynecology*, 44: 824-835.
33. Backhauss C & Kriegelstein J (1992). Extract of kava (*Piper methysticum*) and its methysticin constituents protect brain tissue against ischemic damage in rodents. *European Journal of Pharmacology*, 215: 265-269.
34. Oyama Y, Fuchs PA, Katayama N & Noda K (1994). Myricetin and quercetin, the flavonoid constituents of *Ginkgo biloba* extract, greatly reduce oxidative metabolism in both resting and Ca²⁺-loaded brain neurons. *Brain Research*, 635: 125-129.
35. Lee EJ, Chen HY, Wu TW, Chen TY, Ayoub IA & Maynard KI (2002). Acute administration of *Ginkgo biloba* extract (Egb 761) affords neuroprotection against permanent and transient focal cerebral ischemia in Sprague-Dawley rats. *Journal of Neuroscience Research*, 68: 636-645.
36. Mashour NH, Lin G & Frishman WH (1998). Herbal medicines for the treatment of cardiovascular disease: clinical considerations. *Archives of Internal Medicine*, 158: 2225-2234.
37. Meseguer E, Taboada R, Sánchez V, Mena MA, Campos V & Garcia de Yebenes J (2002). Life threatening parkinsonism induced by kava-kava. *Movement Disorders*, 17: 195-196.
38. Shutenko Z, Henry Y, Pinard E, Seylaz J, Potier P, Berthet F, Girard P & Sercombe R (1999). Influence of the antioxidant quercetin *in vivo* on the level of nitric oxide determined by electron paramagnetic resonance in rat brain during global ischemia and reperfusion. *Biochemical Pharmacology*, 57: 199-208.
39. Abd-El-Gawad HM & Califa AE (2001). Quercetin, coenzyme Q10 and L-canavanine as protective agents against lipid peroxidation and nitric oxide generation in endotoxin-induced shock in rat brain. *Pharmacological Research*, 43: 257-263.
40. Schoroeter H, Clinton B, Jeremy PE, Robert JW, Cadenas E & Rice-Evans C (2002). MAPK signaling in neurodegeneration: influences of flavonoids and of nitric oxide. *Neurobiology of Aging*, 23: 861-880.
41. Yamakawa H, Ito Y, Naganawa T, Banno Y, Nakashima S, Yoshimura S, Sawada M, Nishimura Y, Nozawa Y & Sakai N (2000). Activation of caspase-9 and -3 during H₂O₂-induced apoptosis of PC12 cells independent of ceramide formation. *Neurological Research*, 22: 556-564.
42. Jiang D, Jha N, Boonplueang R & Andersen JK (2001). Caspase 3 inhibition attenuates hydrogen peroxide-induced DNA fragmentation but not cell death in neuronal PC12 cells. *Journal of Neurochemistry*, 76: 1745-1755.
43. Wang R, Zhou J & Tang XC (2002). Tacrine attenuates hydrogen peroxide-induced apoptosis by regulating expression of apoptosis-related genes in rat PC12 cells. *Brain Research. Molecular Brain Research*, 107: 1-8.
44. Arredondo F, Blasina F, Echeverry C, Morquio A, Ferreira F, Abin JA, Lafon L & Dajas F (2003). Cytoprotection by *Achyrocline satureioides* (Lam) DC and some of its flavonoids against oxidative stress. *Journal of Ethnopharmacology* (in press).
45. Dajas F, Rivera F, Blasina F, Arredondo F, Echeverry C, Lafon L, Morquio A & Heinzen H (2003). Cell culture protection and *in vivo* neuroprotective capacity of flavonoids. *Neurotoxicity Research*, 5: 377-384.
46. Azuma K, Ippoushi K, Ito H, Higashio H & Terao J (2002). Combination of lipids and emulsifiers enhances the absorption of orally administered quercetin in rats. *Journal of Agricultural and Food Chemistry*, 50: 1706-1712.

47. Oliveira EJ, Watson DG & Grant MH (2002). Metabolism of quercetin and kaempferol by rat hepatocytes and the identification of flavonoid glycosides in human plasma. *Xenobiotica*, 32: 279-287.
48. Dajas F, Costa G, Abin-Carriquiry JA, Echeverry C, Martínez-Borges A & Dajas-Bailador F (2001). Antioxidant and cholinergic neuroprotective mechanisms in experimental parkinsonism. *Functional Neurology*, 17: 37-44.
49. Dajas-Bailador F, Martínez A, Costa G, Abin A, Martignoni E, Nappi G & Dajas F (1998). Hydroxyl radical production in the substantia nigra after 6-hydroxydopamine and hypoxia-reoxygenation. *Brain Research*, 813: 18-25.
50. Costa G, Abin JA & Dajas F (2001). Nicotine prevents striatal dopamine loss produced by 6-hydroxydopamine lesion in the substantia nigra. *Brain Research*, 888: 336-342.
51. Dajas F, Rivera F, Blasina F, Urbanavicius J, Arredondo F, Lafon L, Costa G, Echeverry C, Ferreira M & Morquio A (2002). Mechanisms of neuroprotection. The contribution of quercetin in focal ischemia and cell culture. *XVII Annual Meeting of the Federação de Sociedades Brasileiras de Biologia Experimental (FESBE)*, Salvador, BA, Brazil, August 28-31, Abstract 1241.
52. Sydserff SG, Cross AJ & Green AR (1995). The neuroprotective effect of chlormethiazole on ischaemic neuronal damage following permanent middle cerebral artery ischaemia in the rat. *Neurodegeneration*, 4: 323-328.
53. Swanson RA, Morton MT, Tsao-Wu G, Saavalos RA, Davidson CY & Sharp FR (1990). A semiautomated method for measuring brain infarct volume. *Journal of Cerebral Blood Flow and Metabolism*, 10: 290-293.
54. Margail S, Parmentier J, Callebert M, Allix R & Boulu M (1996). Short therapeutic window for MK801 in transient focal cerebral ischemia in normotensive rats. *Journal of Cerebral Blood Flow and Metabolism*, 16: 107-113.
55. O'Neill M, Hicks CA, Ward MA et al. (2001). LY393615, a novel neuronal Ca²⁺ and Na⁺ channel blocker with neuroprotective effects in models of *in vitro* and *in vivo* cerebral ischemia. *Brain Research*, 888: 138-149.
56. Welton AF, Tobias LD & Fiedler-Nagy C (1986). Effect of flavonoids on arachidonic acid metabolism. *Progress in Clinical and Biological Research*, 213: 231-242.