



Bell, A. (2018) The neurobiology of acute pain. *Veterinary Journal*, 237, pp. 55-62.

There may be differences between this version and the published version. You are advised to consult the publisher's version if you wish to cite from it.

<http://eprints.gla.ac.uk/167619/>

Deposited on: 24 September 2019

Enlighten – Research publications by members of the University of Glasgow\_  
<http://eprints.gla.ac.uk>

1 **Review**

2 **The Neurobiology of Acute Pain**

3 Andrew Bell

4 *School of Veterinary Medicine, University of Glasgow, Glasgow, G61 1QH, UK*

5 *Email address: [andrew.bell@glasgow.ac.uk](mailto:andrew.bell@glasgow.ac.uk)*

6 **Abstract**

7 The mechanisms by which noxious stimuli produce the sensation of pain in animals are  
8 complex. Noxious stimuli are transduced at the periphery and transmitted to the CNS, where  
9 this information is subject to considerable modulation. Finally, the information is projected to  
10 the brain where it is perceived as pain. Additionally, plasticity can develop in the pain  
11 pathway and hyperalgesia and allodynia may develop through sensitisation both peripherally  
12 and centrally. A large number of different ion channels, receptors, and cell types are involved  
13 in pain perception, and it is hoped that through a better understanding of these, new and  
14 refined treatments for pain will result.

15

16 **Keywords:** Pain; Neurobiology; Nociception; Analgesia; Spinal cord

## 17 **Introduction**

18 Pain is the subjective experience of harm in a part of one's body, and is currently more  
19 strictly defined as "an unpleasant sensory and emotional experience associated with actual or  
20 potential tissue damage, or described in terms of such damage" (Williams and Craig, 2016).  
21 By this definition it is an experience, which therefore requires activity of structures in the  
22 brain to be perceived. This is in contrast to nociception which is defined as the encoding and  
23 processing of noxious stimuli in the nervous system. Pain has a fundamentally important  
24 protective role, alerting us to threats and providing an impetus for the preservation of the  
25 integrity of the body. However, in the context of veterinary treatment of illness or injury,  
26 acute pain may become an unwanted consequence that compromises the welfare of animals  
27 in our care. The implications of chronic pain, which is generally defined as pain that extends  
28 beyond the normal duration of healing, can be even more detrimental to wellbeing and is  
29 covered in another article within this issue.

30 A wide variety of thermal, chemical, mechanical and inflammatory stimuli can trigger pain  
31 and the experience of clinical pain is likely a complex amalgam of these stimuli perceived  
32 after significant modulation in the central nervous system. Pain may also result from a lesion  
33 or disease of the nervous system itself. However, the complex mechanisms of this so-called  
34 neuropathic pain, which have received significant attention from pain researchers, are not  
35 covered in detail in this review.

36 The nociceptive pathway that carries signals from the periphery to the brain where pain is  
37 perceived can be broken into components: *Transduction* of noxious stimuli at the periphery,  
38 *transmission* of those stimuli to the central nervous system (CNS), *central integration and*  
39 *modulation* of the signals at the CNS level, and finally *projection* to the brain followed by

40 *perception*. This article aims to detail the important cellular and molecular mechanisms of  
41 each of these stages.

## 42 **Transducing the stimulus at the periphery; the role of the nociceptor.**

43 Nociceptors are primary afferent neurons which project from tissues including skin, muscle,  
44 joints and viscera to the spinal cord or its trigeminal equivalent in the brainstem. Unlike other  
45 classes of primary afferents, e.g. those that convey touch, nociceptors preferentially  
46 transduce stimuli with intensities in the noxious range allowing them to respond to injurious  
47 stimuli (Basbaum et al., 2009). Anatomically, the cell bodies of nociceptors are located in  
48 the dorsal root ganglia (DRG) adjacent to the spinal cord or in the trigeminal ganglia in the  
49 case of sensory information arising from the face. Their axons arise from these cell bodies  
50 and have both a peripheral branch that innervates the tissues where stimuli are transduced,  
51 and central branch innervating the spinal cord. While other classes of sensory primary  
52 afferents may have complex peripheral apparatus for the detection of stimuli, e.g. Meissner's  
53 corpuscles for low threshold touch, nociceptors are present at the periphery as simple  
54 branched free nerve endings (Lumpkin and Caterina, 2007). Despite this apparently simple  
55 anatomical arrangement, nociceptors have a complex array of cellular and molecular  
56 machinery that enables stimulus transduction, as detailed below.

### 57 *Classifying Nociceptors*

58 Primary afferent neurons are classically characterised by their diameter and degree of  
59 myelination which both determine their conduction velocity (see Table 1). Most large  
60 myelinated afferents (termed A $\beta$  fibres) are low frequency mechanoreceptors which respond  
61 to touch or hair movement (Abraira and Ginty, 2013). Two major classes of nociceptor exist.  
62 The first are medium diameter, myelinated afferents (A $\delta$ ) and these are responsible for the  
63 transmission of well-localised 'fast' or 'first' pain (Ringkamp et al., 2013). The second are

64 unmyelinated, small C-fibre nociceptors which convey poorly localised ‘slow’ pain. The A $\delta$   
65 nociceptors can be further subdivided into two functional groups (Treede et al., 1998). Type I  
66 A $\delta$  nociceptors respond to mechanical and chemical stimuli but have high heat thresholds but  
67 will sensitise in the context of tissue injury. These fibres are probably responsible for ‘first’  
68 pain from mechanical stimuli such as a pin prick. Type II fibres have a much lower heat  
69 threshold and a high mechanical threshold and are involved in ‘fast’ pain responses to heat.

70 Mammalian C-fibre nociceptors can be further classified, not by their functional or  
71 conduction properties but on a molecular basis based on receptors and neurochemicals that  
72 they express. A wide range of markers has been studied with the aim of defining neuronal  
73 subpopulations and correlating these with the response properties of the nociceptors. It is  
74 common practice to divide the nociceptive C-fibres into two groups; the peptidergic fibres  
75 marked by the expression of calcitonin gene-related peptide (CGRP) and substance P, and the  
76 non-peptidergic group identified by their binding of isolectin B4 (IB4) (Snider and  
77 McMahon, 1998). A plethora of other single markers, such as the transient receptor potential  
78 channels (TRP channels) and the Mrg family of G-protein linked receptors, have been  
79 suggested to define functional populations (Zhang et al., 2013) but recent unbiased molecular  
80 strategies aiming to define classes within all sensory primary afferents indicate the situation  
81 is clearly more complex (Li et al., 2016; Usoskin et al., 2014). These studies have sought to  
82 define primary afferent heterogeneity using modern molecular techniques to analyse mRNA  
83 transcripts in the cell bodies of DRG neurons. Individual cells can then be classified, not by a  
84 single marker, but by the constellation of genes that they express. The result is that primary  
85 afferents are grouped on a molecular basis into ten or eleven subgroups and in one study the  
86 functional significance of these groups has been interrogated (Li et al., 2016).

87 It is of interest that not all sensory C-fibres are nociceptive. Small subgroups of C fibres  
88 appear to be specifically responsible for itch transduction and are termed pruritoceptors (Han

89 et al., 2013; Mishra and Hoon, 2013). Additionally, two small classes of C-fibres are low  
90 threshold afferents (termed C low threshold mechanoreceptors, (C-LTMR)) that are involved  
91 in the transduction of pleasant or gentle touch sensations (Seal et al., 2009; Vrontou et al.,  
92 2013).

93 Much of the knowledge about nociceptors is derived from those that innervate the skin, so  
94 called cutaneous nociceptors, rather than those that innervate the viscera and convey the  
95 impulses that can lead to the sensation of visceral pain. As such, much of what is presented in  
96 this section is relevant only to cutaneous nociceptors. In some respects, visceral sensory  
97 neurons with the capacity to convey nociceptive information are similar to cutaneous  
98 nociceptors; their cell bodies are present in the DRG (or nodose ganglia in the case of vagal  
99 visceral afferents), and they generally possess thinly myelinated or unmyelinated axons and  
100 small to medium sized cell bodies (i.e. A $\delta$  & C fibres). Some visceral afferents, however,  
101 traverse pre- and paravertebral ganglia en route to the spinal cord (Gebhart and Bielefeldt,  
102 2011). Importantly, the viscera are sparsely innervated compared to the non-visceral tissues,  
103 and visceral nociceptors have markedly different response properties. Specifically, visceral  
104 nociception and hence pain does not arise from cutting or burning of organs, rather it arises  
105 from distension, traction, ischaemia and through release of chemical mediators of  
106 inflammation. While our knowledge of the biochemical differences underlying these  
107 functional differences is incomplete, it is safe to say that the make-up of receptors and ion  
108 channels present on visceral nociceptors is unique, and several notable differences have been  
109 reported (Robinson and Gebhart, 2008).

#### 110 *Mechanisms of stimulus transduction*

111 Acute noxious stimuli may be thermal, mechanical or chemical and specific ion channels and  
112 G-protein linked receptors are involved in conversion of the stimulus into electrical signals in

113 the primary afferents. These channels generate an electrical current through either opening,  
114 hence allowing the influx of  $\text{Na}^+$  or  $\text{Ca}^{2+}$ , or closing if the channel is responsible for a  
115 hyperpolarising current (e.g. a  $\text{K}^+$  channel) (Gold, 2013). Many chemical stimuli act via G-  
116 protein linked receptors and in these cases intracellular signalling pathways indirectly modify  
117 ion channel activity.

118 The specific channels and receptors that transduce stimuli have been studied extensively with  
119 the transient receptor potential channels (TRP channels) being of importance. In the case of  
120 heat sensation the TRP Vanilloid 1 (TRPV1) channel would appear to play a prominent role  
121 (Cavanaugh et al., 2009). This is of particular interest as TRPV1 is the receptor for capsaicin,  
122 the active ingredient in chilli peppers, and there has been significant attention paid to  
123 developing drugs acting here (Brown, 2016). TRPV1 is one of some 30 or so members of the  
124 transient receptor potential family with other channels also important for stimulus  
125 transduction. TRP Melastatin 8 (TRPM8), the receptor for menthol, is proposed to have  
126 major roles in the transduction of cold stimuli (Bautista et al., 2007). Other thermotransducers  
127 also contribute to temperature sensation and these include two-pore potassium channels  
128 (K2P) (Noël et al., 2009) and voltage gated sodium channels (Zimmermann et al., 2007). A  
129 number of candidate proteins have also emerged as important contributors to noxious  
130 mechanosensation. These include the acid-sensitive ion channels (ASICs) (Omerbašić et al.,  
131 2015), Piezo channels (Coste et al., 2010) , TRP Ankyrin 1 (TRPA1) (Corey et al., 2004), and  
132 K2P channels , although the molecular basis for mechanotransduction requires further  
133 clarification (Basbaum et al., 2009).

134 The ability to detect chemical signals is an important requirement for an organism for  
135 avoiding both environmental noxious chemicals and also to detect endogenous irritants that  
136 may be produced as a result of injury and inflammation. The TRP channels are particularly  
137 important here acting as the receptors for capsaicin (TRPV1), mustards and garlic (TRPA1),



138 and a wide array of other chemical irritants (e.g. TRPA1 transduces the aversive smell of  
139 isoflurane). A number of ion channels (e.g. ASICs) and a wide variety of G-protein linked  
140 receptors are present on peripheral nociceptor terminals (Yaksh et al., 2015) which can sense  
141 the substances produced by the process of inflammation and sensitise the nociceptor giving  
142 rise to lower thresholds. These mechanisms are discussed in more details below. However, it  
143 is also worth stating that a number of mechanisms exist whereby nociceptor activity can be  
144 modulated peripherally (Pan et al., 2008). Antinociceptive G-protein receptors involved  
145 include opioid, cannabinoid, somatostatin, muscarinic acetylcholine, GABA<sub>B</sub>, and  $\alpha$ 2-  
146 adrenergic receptors and most appear to primarily act via modulation of Ca<sup>2+</sup> channels thus  
147 reducing Ca<sup>2+</sup> entry. This appears to be highly relevant following clinical administration of  
148 exogenous opioid agonists at peripheral sites where they can produce significant analgesic  
149 and anti-inflammatory effects (Iwaszkiewicz et al., 2013; van Loon et al., 2010).

#### 150 *Transducing the stimulus in the context of inflammation*

151 Disease and injury often results in pain that is not explained simply by the transduction of  
152 noxious stimuli as explained above. This so called inflammatory pain is the result of  
153 endogenously generated factors which can activate nociceptor terminals (Dawes et al., 2013).  
154 In addition, these substances can also sensitise nociceptors, which is to lower their response  
155 threshold and increase their response to a given stimulus. This process is termed peripheral  
156 sensitisation and the result for the animal are the phenomena of hyperalgesia, where painful  
157 stimuli are perceived as more painful, and allodynia, where non-noxious stimuli are perceived  
158 as painful.

159 Inflammatory mediators may be released by a multitude of non-neuronal cells including  
160 fibroblasts, keratinocytes, platelets and immune cells, as well as from the peripheral terminals  
161 of activated nociceptors themselves; so called neurogenic inflammation (Chiu et al., 2012).

162 These mediators include prostaglandins, leukotrienes, bradykinin, serotonin, histamine,  
163 CGRP, substance P, purines such as ATP, protons, free radicals, lipids, cytokines,  
164 chemokines, and neurotrophins such as nerve growth factor (NGF) (Yaksh et al., 2015).  
165 Sensitisation may then occur by one of 3 mechanisms; i) direct activation of cation channels  
166 causing nociceptor activation, ii) activation of intracellular regulatory pathways via G-  
167 proteins to indirectly alter (e.g. phosphorylate) membrane proteins, or iii) alterations to the  
168 transcriptional phenotype of the cell (Dawes et al., 2013). In the case of cytokines and  
169 chemokines, while some evidence exists to suggest a direct action on nociceptors, their  
170 proalgesic action most likely arises from the strengthening of the inflammatory response and  
171 consequent release of other mediators.

172 A number of factors present in the ‘inflammatory soup’ have received particular attention in  
173 terms of the development of therapeutics to target inflammatory pain and peripheral  
174 sensitisation. Prostaglandins, leukotrienes and thromboxanes, collectively termed  
175 eicosanoids, are thought to sensitise nociceptors rather than activate them directly (Pethő and  
176 Reeh, 2012) but the inhibition of prostaglandin synthesis by cyclooxygenase inhibitors is a  
177 common and efficacious approach to pain treatment (KuKanich et al., 2012). Inhibition of  
178 other lipid inflammatory mediators, such as the soluble epoxide hydrolase pathway shows  
179 promise (Guedes et al., 2017). Antagonism of the prostaglandin E2 receptor has also proved  
180 to be an efficacious treatment for osteoarthritis pain in dogs (Rausch-Derra et al., 2016),  
181 which may have a reduced risk of adverse effects compared to conventional cyclooxygenase  
182 inhibition. Nerve growth factor is released during inflammation and acts via the tyrosine  
183 kinase (TrkA) receptor expressed on nociceptors to produce hyperalgesia. A potential  
184 therapeutic approach to pain that is being investigated is the use of monoclonal antibodies to  
185 NGF to reduce its action (Lascelles et al., 2015).

186 *Transmission of the Stimulus*

187 Once the noxious stimulus has been transduced at the periphery it must be transmitted as an  
188 action potential to the central nervous system. Voltage-gated sodium and potassium channels  
189 are involved in the generation of this action potential. Different classes of sodium channels,  
190 such as Nav 1.1, Nav 1.6, Nav 1.7, Nav 1.8 and Nav 1.9, are expressed in sensory neurons,  
191 with the later three being predominantly expressed in nociceptors. Recently, mutations  
192 within the Nav 1.7 channel have been shown to underlie dramatic insensitivities to pain in  
193 human subjects (Cox et al., 2006) while gain of function mutations here have been shown to  
194 cause painful disorders. As sodium channel subtypes seem to be differentially expressed in  
195 nociceptive and non-nociceptive primary afferents, there has been interest in these targets for  
196 the development of novel analgesics (Emery et al., 2016). Voltage-gated calcium channels  
197 are involved in neurotransmitter release at central and peripheral terminals. Calcium channels  
198 are composed of  $\alpha 1$  pore forming subunits and  $\alpha 2$  modulatory subunits. The  $\alpha 2\delta$  subunit is  
199 highly expressed in C nociceptors particularly after nerve injury and is an analgesic target of  
200 gabapentin (Li et al., 2006).

### 201 **Integration in the Dorsal Horn of the Spinal Cord**

202 The dorsal horn of the spinal cord is the site of the first synapse of the primary afferent  
203 neuron and is a site of tremendous modulation and integration of sensory information before  
204 it is projected to the brain. Given the remarkable heterogeneity in the neurobiology of  
205 nociceptive primary afferents, we are at an early stage in terms of our understanding of how  
206 these inputs both transmit specific stimuli and how this is deciphered by the CNS thereby  
207 resulting in a pain percept. The dorsal horn undeniably plays a major role in this processing  
208 and whilst the theoretical frameworks for pain processing are beyond the scope of this review,  
209 those interested readers are referred to an excellent review by Moayed and Davis (2013).

210 The grey matter of the spinal cord is divided into laminae based on cytoarchitectonic criteria  
211 (fig 1) (Rexed, 1952). The superficial dorsal horn receives the majority of nociceptive  
212 afferent input and is composed of laminae I and II. In this area the dorsal horn contains 4  
213 basic neural components: the central terminals of primary afferent neurons, excitatory and  
214 inhibitory interneurons, projection neurons and descending modulatory axons (Todd, 2010).  
215 Primary afferents terminate in the dorsal horn in a well-ordered fashion determined by fibre  
216 type within a somatotopic arrangement. The input to the dorsal horn is also stratified by  
217 somatosensory modality.  $A\delta$  nociceptors end mainly in lamina I, peptidergic primary  
218 afferents arborize mainly in laminae I and II outer whereas most non-peptidergic C fibres  
219 form a band occupying the central part of lamina II (Fig 1). Non-nociceptive afferents such as  
220  $A\beta$  tactile fibres end mainly in the deeper laminae III-V (Abraira et al., 2017). Within this  
221 arrangement synapses with second-order neurons may have simple single synaptic  
222 arrangements or may form complex synaptic glomeruli which give rise to numerous synapses  
223 and receive axo-axonic inhibitory inputs from local interneurons (Ribeiro-da-Silva and  
224 Coimbra, 1982). Primary afferents all use glutamate as their principle neurotransmitter and  
225 hence all synapses with second order neurons are excitatory. These second order neurons  
226 receiving synapses may be interneurons which form complex circuits within the dorsal horn,  
227 or projection neurons.

228 Visceral nociceptive primary afferents arborise in a unique way in the dorsal horn and this  
229 pattern of innervation, alongside the fact that the viscera are comparatively sparsely  
230 innervated, underlies the often diffuse and poorly localised nature of visceral pain. These  
231 afferents project extensively to both superficial and deep laminae (I, II, V & X) but notably  
232 spread out over several spinal segments and may decussate on the opposite side of the cord  
233 (Gebhart and Bielefeldt, 2011). Viscerosomatic convergence is common, such that almost all  
234 second order spinal neurons receiving visceral input also receive somatic input from skin or

235 muscle. This provides an explanation for the phenomenon of referred pain, where visceral  
236 nociception is not perceived at the site of origin, rather at an adjacent or distant somatic site  
237 (Cervero, 1994). A good example of this is pain felt in the shoulder which results from gas  
238 accumulation in the abdomen and consequent diaphragmatic irritation following laparoscopy.

### 239 *Projection Neurons; the Output from the Dorsal Horn*

240 For a noxious stimulus to be perceived as painful, it must first be projected to higher centres  
241 in the brain. Nociceptive specific projection neurons are concentrated in lamina I and  
242 scattered through laminae III-VI. Projection neurons in deeper laminae (V) may not be  
243 specific to noxious stimuli and are termed wide dynamic range neurons (WDR) as they  
244 respond to a broad range of input and encode stimulus intensity (Sikandar et al., 2013).  
245 Despite their importance, projection neurons only comprise around 5% of the cells in the  
246 superficial dorsal horn (Spike et al., 2003).

247 The axons of these projection cells cross the midline, travel in ascending spinal white matter  
248 tracts and innervate various brainstem and thalamic nuclei. The white matter tracts involved  
249 in the projection of nociceptive information vary depending on species; the spinothalamic  
250 tract would appear to be most important in humans and primates, the spinocervicothalamic  
251 tract predominant in carnivores, and the spinoparabrachial tract most important in rodents  
252 (Dostrovsky and Craig, 2013). The brainstem and thalamic nuclei receiving projections  
253 include the medulla (caudal ventrolateral medulla (CVLM) & the rostral ventromedial  
254 medulla (RVM)), the parabrachial area (Pb), the periaqueductal grey (PAG), the nucleus of  
255 the solitary tract (NTS) and the thalamus (Figure 2). Each of these areas is believed to code  
256 for specific dimensions of the pain experience (West et al., 2015). The parabrachial area is  
257 thought to be particularly important in terms of the affective component of pain as its output  
258 provides for a rapid connection to the amygdala and hypothalamus. The thalamus has been

259 associated with the sensory-discriminatory aspects of pain due to its connections to the  
260 somatosensory cortex. Both the PAG and CVLM are thought to be upstream of other  
261 brainstem areas that control powerful descending inputs to the spinal cord (see below).  
262 The majority (80%) of lamina I projection neurons express the neurokinin 1 receptor (NK1r)  
263 upon which substance P acts. This receptor has attracted considerable interest as selective  
264 ablation of the cells expressing NK1r reduces hyperalgesia in inflammatory, cancer and  
265 neuropathic pain models (Mantyh et al., 1997) including in studies conducted in clinical  
266 canine patients (Brown and Agnello, 2013).

### 267 *Synaptic Mechanisms and Plasticity*

268 As mentioned above, all synapses from primary afferents onto second order neurons are  
269 excitatory and use glutamate as a neurotransmitter. Although glutamate is the primary  
270 neurotransmitter, it can be co-localised with the neuropeptides substance P and CGRP, which  
271 also play a role in nociceptive signalling (De Biasi and Rustioni, 1988). Glutamate acts on  
272 three ionotropic receptors; the kainate receptor, the alpha-amino3-hydroxy-5-methy-4-  
273 isoxazolepropionic acid (AMPA) receptor, and the N-methyl-D-aspartate (NMDA) receptor.  
274 It also acts via the metabotropic glutamate receptor (mGlu).

275 Synapses within the dorsal horn display activity-dependent plasticity in response to  
276 prolonged or high intensity noxious input (Sandkühler, 2009). A result of this inherent  
277 plasticity is facilitation of the signal such that the information relayed to higher centres is not  
278 coupled to the intensity or duration of the peripheral stimulus (Latremoliere and Woolf,  
279 2009). This phenomenon is commonly termed central sensitisation and results in hyperalgesia  
280 and allodynia. Central sensitisation can be readily and rapidly elicited in human volunteers,  
281 thus is important in the physiology of acute pain as well as being commonly present in many  
282 chronic pain syndromes in humans (Woolf, 2011). The desire to avoid the induction of

283 central sensitisation also underlies the desire of many anaesthetists to practice preemptive  
284 analgesia for surgery. Here, central sensitisation is reduced by the use of analgesics to block  
285 the intraoperative nociceptive barrage, thus reducing the magnitude of postoperative acute  
286 pain. While this concept has proved controversial and difficult to prove in the human pain  
287 literature (Katz et al., 2011), evidence in veterinary species is encouraging (Lascelles et al.,  
288 1997) and the concept has been validated experimentally (LaMotte et al., 1992).

289 Multiple mechanisms underlie central sensitisation but it is well established that the NMDA  
290 receptor plays an integral role as its antagonism inhibits activity-dependent plasticity  
291 (Bergadano et al., 2009; Dickenson and Sullivan, 1987). The NMDA receptor is usually  
292 blocked by a  $Mg^{2+}$  ion at resting membrane potential. When the post-synaptic neuron  
293 undergoes sustained depolarisation due to the actions of glutamate and also SP and CGRP,  
294 this blockade is lifted and glutamate can activate the receptor. This results in a greater influx  
295 of  $Na^+$  and  $Ca^{2+}$  and thus amplification of the signal. Subsequent maintenance of central  
296 sensitisation occurs due to increases in activity of a number of second messenger systems as a  
297 result of an increase in cytosolic  $Ca^{2+}$ . Increased activity of intracellular kinases serves to  
298 phosphorylate receptors, recruit new receptors and alter gene expression resulting in altered  
299 synaptic responses and continuation of sensitisation (Sandkühler and Gruber-Schoffnegger,  
300 2012).

301 Conversely, a number of endogenous mechanisms may reduce synaptic transmission; the  
302 three principle opioid receptors ( $\mu$ ,  $\delta$  and  $\kappa$ ) are present in high concentrations both  
303 pre and post-synaptically in the dorsal horn. Pre-synaptically, opioid receptor activation  
304 reduces neurotransmitter release via reducing calcium influx and this is the major mechanism  
305 of their analgesic action here (Kohno et al., 1999). It should be noted that opioids also exert a  
306 significant effect at supra-spinal levels, particularly at the level of the rostral ventromedial

307 medulla (RVM). Additional endogenous modulation of noxious inputs to the dorsal horn may  
308 occur via descending mechanisms, or inhibitory circuits as discussed below.

### 309 *Spinal Cord Circuitry*

310 The concept of the dorsal horn as a site of modulatory circuits for noxious signals dates back  
311 to Wall and Melzack's gate control theory (GCT) of pain (Melzack and Wall, 1965). GCT  
312 proposed that the extent to which a stimulus produced pain was not just a function of the  
313 magnitude of the signal in nociceptive specific primary afferents, rather this activity could be  
314 modulated at the level of the spinal cord by non-nociceptive afferents. At the centre of the  
315 gate control circuit lies an inhibitory interneuron which can be activated by the large fibres  
316 resulting in feed-forward inhibition of the action system. Nociceptive specific primary  
317 afferents not only activate the action system but also reduce the activity of the 'gating'  
318 inhibitory interneuron. It has been demonstrated that the selective inactivation of large  
319 populations of inhibitory interneurons in the dorsal horn results in spontaneous pain and itch  
320 behaviour and hyperalgesia (Duan et al., 2014; Foster et al., 2015). While these studies and  
321 many others previously provide support for the important role of spinal circuitry, the actual  
322 circuits involved may be significantly more complex than originally proposed (Mendell,  
323 2014; Peirs and Seal, 2016) containing both excitatory and inhibitory interneurons. The  
324 precise identities of spinal neurons and circuits that transmit and gate pain related information  
325 remain largely unknown (Zeilhofer et al., 2012).

326 One of the major barriers to identifying circuits that modulate noxious information is the  
327 difficulty in defining functional populations among the interneurons (Todd, 2017). As already  
328 stated there are two main types; inhibitory and excitatory interneurons. Inhibitory  
329 interneurons use  $\gamma$ -aminobutyric acid (GABA) or glycine as neurotransmitters although many  
330 co-express both (Todd, 2010) while excitatory interneurons use glutamate as a



331 neurotransmitter. A number of classification schemes for superficial dorsal horn interneurons  
332 have been suggested and electrophysiological techniques can be used to identify subtypes by  
333 input or firing patterns (Yasaka et al., 2010). However, a morphological classification scheme  
334 (Grudt and Perl, 2002) has become a widely adopted classification. This describes four  
335 morphologically distinguishable interneurons: islet, central, radial and vertical cells. Some of  
336 these cells types have been associated with specific roles in nociceptive circuitry (Lu et al.,  
337 2013; Lu and Perl, 2003) but this classification scheme leaves a substantial proportion of  
338 cells unclassified (Yasaka et al., 2007). An alternative approach to classification has been to  
339 use neurochemical markers such as neuropeptides, receptors, enzymes and calcium binding  
340 proteins that are expressed by discrete populations of interneurons. GABAergic interneurons  
341 have recently been divided into non-overlapping subpopulations defined by expression of  
342 galanin/dynorphin, neuropeptide Y, neuronal nitric oxide synthase and parvalbumin and these  
343 appear to be functionally as well as neurochemically distinct (Polgár et al., 2013; Duan et al.,  
344 2014; Bourane et al., 2015; Petitjean et al., 2015). Excitatory interneurons have also recently  
345 been divided into distinct groups based on their expression of somatostatin, substance P,  
346 gastrin-releasing peptide and protein kinase C gamma (PKC $\delta$ ) (Gutierrez-Mecinas et al.,  
347 2016, 2017).

348 Based on current knowledge, it is difficult to rationalise clinical or therapeutic decisions  
349 using our evolving knowledge of circuitry. However, manipulation of spinal circuits can  
350 powerfully influence nociceptive transmission in many ways. In particular circuits involving  
351 excitatory interneurons underlie the potential for touch information to be able to access  
352 nociceptive specific projection neurons via heterosynaptic facilitation, hence causing  
353 allodynia (Graham et al., 2007). Recent studies have demonstrated the important roles of  
354 excitatory interneurons expressing somatostatin (Christensen et al., 2016; Duan et al., 2014),  
355 PKC $\delta$  (Lu et al., 2013) and vesicular glutamate transporter 3 (VGLUT3) (Peirs et al., 2015)

356 in the generation of heightened pain states. Adequate inhibitory control of these pathways by  
357 the GABAergic or glycinergic interneuron population is important for prevention  
358 development of hyperalgesia and allodynia (Foster et al., 2015) . It is thought that a number  
359 of clinical chronic pain states may be due to disinhibition of spinal cord circuits. Quite why  
360 this may occur is not entirely clear but a promising mechanism may be due to downregulation  
361 of the neuronal potassium-chloride co-transporter (KCC2). This mechanism has been  
362 suggested to be driven by the microglial response in the dorsal horn seen after injury (Coull et  
363 al., 2005). While this mechanism is probably more important in chronic and neuropathic pain,  
364 it may still be important in acute inflammatory pain (McMahon and Malcangio, 2009).  
365 Interestingly, the spinal neuroimmune interactions underlying this sensitisation appear to  
366 differ between males and females in experimental studies, and this may underpin differences  
367 in clinical pain sensitivities between sexes (Mogil, 2012).

### 368 *Descending controls*

369 Descending control pathways from brainstem regions project to the dorsal horn of the spinal  
370 cord. They represent a mechanism through which the transmitted nociceptive signal may be  
371 facilitated - enhancing the pain experienced, or inhibited - reducing pain. They provide a top-  
372 down mechanism whereby cognitive, emotional or autonomic factors can regulate pain  
373 processing at the dorsal horn (Bannister and Dickenson, 2017). One can see the potential  
374 advantages of antinociceptive mechanisms which can be engaged during 'fight or flight'  
375 situations for example. Inputs from many brain regions activate areas of the PAG and this in  
376 turn feeds into the RVM and locus coeruleus (LC). From here neurons project bilaterally to  
377 the dorsal horn where they release 5-hydroxytryptamine (5-HT) and noradrenaline (NA).  
378 Noradrenaline acts via  $\alpha_2$ -adrenoceptors resulting in antinociceptive effects. The situation  
379 with 5-HT is somewhat more complex as it can result in anti- or pro-nociceptive effects  
380 through action at either the 5-HT<sub>7</sub> or 5-HT<sub>3</sub> receptors respectively (Dogrul et al., 2009).

381 Recent evidence points to RVM neurons releasing 5-HT having a predominantly facilitatory  
382 effect on nociceptive transmission (Cai et al., 2014). Notably, descending facilitation rather  
383 than inhibition predominates overall in young animals. This dominance alongside numerous  
384 other neurobiological changes may underlie the excitatory dominance that is a feature of pain  
385 in neonates and infants (Fitzgerald, 2015).

386 One consequence of this descending pain modulation system is that one pain can inhibit  
387 another, e.g. an ear pinch may reduce the pain from a toe pinch. This phenomenon is termed  
388 diffuse noxious inhibitory controls (DNIC) and this can be easily evoked and quantified in  
389 animals and man, thus allowing measurement of the activity of the descending pathways  
390 (Bannister and Dickenson, 2017). Reduced DNIC is indicative of altered descending  
391 modulation which may be predictive of the actions of analgesic drugs and the degree of pain  
392 morbidity after injury (Yarnitsky, 2010).

### 393 **Representation of Pain in the Brain**

394 As outlined above, information from the spinal cord is projected to various centres in the  
395 brain and the resultant neuronal activity gives rise to the multidimensional experience that is  
396 pain. Functional imaging studies in humans and animals have increased our knowledge of the  
397 brain areas involved hugely (Borsook and Becerra, 2011; Guillot et al., 2015; Tracey and  
398 Mantyh, 2007). This combined set of brain regions is often referred to as the "pain matrix"  
399 and encompasses areas involved with sensory-discriminative, affective and cognitive aspects.

400 However, the pain matrix is not entirely specific to pain in the way the visual cortex, for  
401 example, is to sight. Instead, none of the regions are unique to pain and many are involved in  
402 other aspects of perception and behaviour (Tracey and Johns, 2010). Recently the dorsal  
403 posterior insula, and its analogue in other animals, has been suggested as a fundamentally  
404 important site for tracking the intensity component of a noxious stimulus at a cerebral level

405 (Segerdahl et al., 2015). However, while similar regions are implicated in cerebral pain  
406 processing and perception in both animals and humans (Thompson and Bushnell, 2012), it is  
407 currently unknown how the complex interplay between these areas determines what  
408 constitutes a painful experience across the phylogeny. Cognitive and affective top-down  
409 regulation of pain via the ‘pain matrix’ also underlies the placebo and nocebo effects (Tracey,  
410 2010) which are reliant, at least in humans, on prior experience and expectation. The  
411 investigation of true placebo effects in animals is at a very early stage (Muñana et al., 2010).

412

### 413 **Conclusions**

414 Acute pain is common as a result of surgery, illness or injury and is undeniably unpleasant.  
415 Our knowledge of the circuits and cellular and molecular mechanisms underlying acute pain  
416 and plasticity is expanding rapidly. In parallel to this explosion of interest in nociceptive  
417 mechanisms, researchers are developing novel and highly specific pharmaceutical and  
418 genetic techniques to precisely manipulate biological systems. This knowledge should enable  
419 the development of new therapeutics and approaches to treating pain based on mechanistic  
420 principles with increased efficacy and fewer side effects; ultimately enabling veterinarians to  
421 treat pain and the resultant suffering in animals more effectively.

### 422 **Conflict of interest statement**

423 None of the authors of this paper has a financial or personal relationship with other people or  
424 organisations that could inappropriately influence or bias the content of the paper.

425

### 426 **References**

427 Abraira, V.E., Ginty, D.D., 2013. The Sensory Neurons of Touch. *Neuron* 79, 618–639.

- 428 Abraira, V.E., Kuehn, E.D., Chirila, A.M., Springel, M.W., Toliver, A.A., Zimmerman, A.L.,  
429 Orefice, L.L., Boyle, K.A., Bai, L., Song, B.J., Bashista, K.A., O'Neill, T.G., Zhuo,  
430 J., Tsan, C., Hoynoski, J., Rutlin, M., Kus, L., Niederkofler, V., Watanabe, M.,  
431 Dymecki, S.M., Nelson, S.B., Heintz, N., Hughes, D.I., Ginty, D.D., 2017. The  
432 Cellular and Synaptic Architecture of the Mechanosensory Dorsal Horn. *Cell* 168,  
433 295–310.e19.
- 434 Bannister, K., Dickenson, A.H., 2017. The plasticity of descending controls in pain:  
435 translational probing. *Journal of Physiology* 595, 4159–4166.
- 436 Basbaum, A.I., Bautista, D.M., Scherrer, G., Julius, D., 2009. Cellular and Molecular  
437 Mechanisms of Pain. *Cell* 139, 267–284.
- 438 Bautista, D.M., Siemens, J., Glazer, J.M., Tsuruda, P.R., Basbaum, A.I., Stucky, C.L., Jordt,  
439 S.-E., Julius, D., 2007. The menthol receptor TRPM8 is the principal detector of  
440 environmental cold. *Nature* 448, 204–208.
- 441 Bergadano, A., Andersen, O.K., Arendt-Nielsen, L., Theurillat, R., Thormann, W.,  
442 Spadavecchia, C., 2009. Plasma levels of a low-dose constant-rate-infusion of  
443 ketamine and its effect on single and repeated nociceptive stimuli in conscious dogs.  
444 *The Veterinary Journal* 182, 252–260.
- 445 Borsook, D., Becerra, L., 2011. CNS animal fMRI in pain and analgesia. *Neuroscience*  
446 *Biobehavioural Reviews* 35, 1125–1143.
- 447 Bourane, S., Duan, B., Koch, S.C., Dalet, A., Britz, O., Garcia-Campmany, L., Kim, E.,  
448 Cheng, L., Ghosh, A., Ma, Q., Goulding, M., 2015. Gate control of mechanical itch  
449 by a subpopulation of spinal cord interneurons. *Science* 350, 550–554.
- 450 Brown, D.C., 2016. Resiniferatoxin: The Evolution of the “Molecular Scalpel” for Chronic  
451 Pain Relief. *Pharmaceuticals (Basel)* 9.
- 452 Brown, D.C., Agnello, K., 2013. Intrathecal substance-p saporin in the dog: Efficacy in bone  
453 cancer pain. *Anesthesiology* 119.
- 454 Cai, Y.-Q., Wang, W., Hou, Y.-Y., Pan, Z.Z., 2014. Optogenetic activation of brainstem  
455 serotonergic neurons induces persistent pain sensitization. *Molecular Pain* 10, 70.
- 456 Cavanaugh, D.J., Lee, H., Lo, L., Shields, S.D., Zylka, M.J., Basbaum, A.I., Anderson, D.J.,  
457 2009. Distinct subsets of unmyelinated primary sensory fibers mediate behavioral  
458 responses to noxious thermal and mechanical stimuli. *Proceedings of the National*  
459 *Academy of Sciences* 106, 9075–9080.
- 460 Cervero, F., 1994. Sensory innervation of the viscera: peripheral basis of visceral pain.  
461 *Physiological Reviews*. 74, 95–138.
- 462 Chiu, I.M., von Hehn, C.A., Woolf, C.J., 2012. Neurogenic Inflammation – The Peripheral  
463 Nervous System’s Role in Host Defense and Immunopathology. *Nature Neuroscience*  
464 15, 1063–1067.
- 465 Christensen, A.J., Iyer, S.M., François, A., Vyas, S., Ramakrishnan, C., Vesuna, S.,  
466 Deisseroth, K., Scherrer, G., Delp, S.L., 2016. In Vivo Interrogation of Spinal  
467 Mechanosensory Circuits. *Cell Reports* 17, 1699–1710.
- 468 Corey, D.P., García-Añoveros, J., Holt, J.R., Kwan, K.Y., Lin, S.-Y., Vollrath, M.A.,  
469 Amalfitano, A., Cheung, E.L.-M., Derfler, B.H., Duggan, A., Géléoc, G.S.G., Gray,  
470 P.A., Hoffman, M.P., Rehm, H.L., Tamasauskas, D., Zhang, D.-S., 2004. TRPA1 is a

- 471 candidate for the mechanosensitive transduction channel of vertebrate hair cells.  
472 Nature 432, 723–730.
- 473 Coste, B., Mathur, J., Schmidt, M., Earley, T.J., Ranade, S., Petrus, M.J., Dubin, A.E.,  
474 Patapoutian, A., 2010. Piezo1 and Piezo2 are essential components of distinct  
475 mechanically-activated cation channels. Science 330, 55–60.
- 476 Coull, J.A.M., Beggs, S., Boudreau, D., Boivin, D., Tsuda, M., Inoue, K., Gravel, C., Salter,  
477 M.W., De Koninck, Y., 2005. BDNF from microglia causes the shift in neuronal  
478 anion gradient underlying neuropathic pain. Nature 438, 1017–1021.
- 479 Cox, J.J., Reimann, F., Nicholas, A.K., Thornton, G., Roberts, E., Springell, K., Karbani, G.,  
480 Jafri, H., Mannan, J., Raashid, Y., Al-Gazali, L., Hamamy, H., Valente, E.M.,  
481 Gorman, S., Williams, R., McHale, D.P., Wood, J.N., Gribble, F.M., Woods, C.G.,  
482 2006. An SCN9A channelopathy causes congenital inability to experience pain.  
483 Nature 444, 894–898.
- 484 Dawes, J., Andersson, D., Bennet, D., Bevan, S., McMahon, S.B., 2013. Inflammatory  
485 Mediators and Modulators of Pain. In: Wall and Melzack's Textbook of Pain (Editors  
486 McMahon, Koltzenburg, Tracey, and Turk). Elsevier, pp. 48–67.
- 487 De Biasi, S., Rustioni, A., 1988. Glutamate and substance P coexist in primary afferent  
488 terminals in the superficial laminae of spinal cord. Proceedings of the National  
489 Academy of Sciences 85, 7820–7824.
- 490 Dickenson, A.H., Sullivan, A.F., 1987. Evidence for a role of the NMDA receptor in the  
491 frequency dependent potentiation of deep rat dorsal horn nociceptive neurones  
492 following C fibre stimulation. Neuropharmacology 26, 1235–1238.
- 493 Dogrul, A., Ossipov, M.H., Porreca, F., 2009. Differential mediation of descending pain  
494 facilitation and inhibition by spinal 5HT-3 and 5HT-7 receptors. Brain Research  
495 1280, 52–59.
- 496 Dostrovsky, J., Craig, B., 2013. Ascending Projection Systems. In: FSB, S.M.Fm., FRCP,  
497 M.K.M., FRCA, I.T.M.P., PhD, D.C.T. (Eds.), Wall & Melzack's Textbook of Pain.  
498 Saunders, Philadelphia, PA, pp. 182–197.
- 499 Duan, B., Cheng, L., Bourane, S., Britz, O., Padilla, C., Garcia-Campmany, L., Krashes, M.,  
500 Knowlton, W., Velasquez, T., Ren, X., Ross, S.E., Lowell, B.B., Wang, Y., Goulding,  
501 M., Ma, Q., 2014. Identification of Spinal Circuits Transmitting and Gating  
502 Mechanical Pain. Cell 159, 1417–1432.
- 503 Emery, E.C., Luiz, A.P., Wood, J.N., 2016. Nav1.7 and other voltage-gated sodium channels  
504 as drug targets for pain relief. Expert Opinion Therapeutic Targets 20, 975–983.
- 505 Fitzgerald, M., 2015. What do we really know about newborn infant pain? Experimental  
506 Physiology 100, 1451–1457.
- 507 Foster, E., Wildner, H., Tudeau, L., Haueter, S., Ralvenius, W.T., Jegen, M., Johannssen, H.,  
508 Hösli, L., Haenraets, K., Ghanem, A., Conzelmann, K.-K., Bösl, M., Zeilhofer, H.U.,  
509 2015. Targeted ablation, silencing, and activation establish glycinergic dorsal horn  
510 neurons as key components of a spinal gate for pain and itch. Neuron 85, 1289–1304.
- 511 Gebhart, G.F., Bielefeldt, K., 2011. Physiology of Visceral Pain. In: Comprehensive  
512 Physiology. John Wiley & Sons, Inc.

- 513 Gold, M., 2013. Molecular Biology of Sensory Transduction. In: Wall and Melzack's  
514 Textbook of Pain (Editors McMahon, Koltzenburg, Tracey, and Turk). Elsevier, pp.  
515 31–47.
- 516 Graham, B.A., Brichta, A.M., Callister, R.J., 2007. Moving From an Averaged to Specific  
517 View of Spinal Cord Pain Processing Circuits. *Journal of Neurophysiology* 98, 1057–  
518 1063.
- 519 Grudt, T.J., Perl, E.R., 2002. Correlations between neuronal morphology and  
520 electrophysiological features in the rodent superficial dorsal horn. *Journal of*  
521 *Physiology* 540, 189–207.
- 522 Guedes, A.G.P., Aristizabal, F., Sole, A., Adedeji, A., Brosnan, R., Knych, H., Yang, J.,  
523 Hwang, S.-H., Morisseau, C., Hammock, B.D., 2017. Pharmacokinetics and  
524 antinociceptive effects of the soluble epoxide hydrolase inhibitor t-TUCB in horses  
525 with experimentally induced radiocarpal synovitis. *Journal of Veterinary*  
526 *Pharmacology and Therapeutics*.
- 527 Guillot, M., Chartrand, G., Chav, R., Rousseau, J., Beaudoin, J.-F., Martel-Pelletier, J.,  
528 Pelletier, J.-P., Lecomte, R., de Guise, J.A., Troncy, E., 2015. [(18)F]-  
529 fluorodeoxyglucose positron emission tomography of the cat brain: A feasibility study  
530 to investigate osteoarthritis-associated pain. *The Veterinary Journal* 204, 299–303.
- 531 Gutierrez-Mecinas, M., Bell, A.M., Marin, A., Taylor, R., Boyle, K.A., Furuta, T., Watanabe,  
532 M., Polgár, E., Todd, A.J., 2017. Preprotachykinin A is expressed by a distinct  
533 population of excitatory neurons in the mouse superficial spinal dorsal horn including  
534 cells that respond to noxious and pruritic stimuli. *Pain* 158, 440–456.
- 535 Gutierrez-Mecinas, M., Furuta, T., Watanabe, M., Todd, A.J., 2016. A quantitative study of  
536 neurochemically defined excitatory interneuron populations in laminae I–III of the  
537 mouse spinal cord. *Molecular Pain* 12, 1744806916629065.
- 538 Han, L., Ma, C., Liu, Q., Weng, H.-J., Cui, Y., Tang, Z., Kim, Y., Nie, H., Qu, L., Patel,  
539 K.N., Li, Z., McNeil, B., He, S., Guan, Y., Xiao, B., LaMotte, R.H., Dong, X., 2013.  
540 A subpopulation of nociceptors specifically linked to itch. *Nature Neurosci* 16, 174–  
541 182.
- 542 Iwaszkiewicz, K.S., Schneider, J.J., Hua, S., 2013. Targeting peripheral opioid receptors to  
543 promote analgesic and anti-inflammatory actions. *Front Pharmacol* 4.
- 544 Katz, J., Clarke, H., Seltzer, Z., 2011. Review article: Preventive analgesia: quo vadimus?  
545 *Anesthesia and Analgesia*. 113, 1242–1253.
- 546 Kohno, T., Kumamoto, E., Higashi, H., Shimoji, K., Yoshimura, M., 1999. Actions of  
547 opioids on excitatory and inhibitory transmission in substantia gelatinosa of adult rat  
548 spinal cord. *Journal of Physiology* 518, 803–813.
- 549 KuKanich, B., Bidgood, T., Knesl, O., 2012. Clinical pharmacology of nonsteroidal anti-  
550 inflammatory drugs in dogs. *Veterinary Anaesthesia and Analgesia* 39, 69–90.
- 551 LaMotte, R.H., Lundberg, L.E., Torebjörk, H.E., 1992. Pain, hyperalgesia and activity in  
552 nociceptive C units in humans after intradermal injection of capsaicin. *J. Physiol.*  
553 (Lond.) 448, 749–764.
- 554 Lascelles, B.D., Cripps, P.J., Jones, A., Waterman, A.E., 1997. Post-operative central  
555 hypersensitivity and pain: the pre-emptive value of pethidine for ovariohysterectomy.  
556 *Pain* 73, 461–471.

- 557 Lascelles, B.D.X., Knazovicky, D., Case, B., Freire, M., Innes, J.F., Drew, A.C., Gearing,  
558 D.P., 2015. A canine-specific anti-nerve growth factor antibody alleviates pain and  
559 improves mobility and function in dogs with degenerative joint disease-associated  
560 pain. *BMC Veterinary Research* 11, 101.
- 561 Latremoliere, A., Woolf, C.J., 2009. Central sensitization: a generator of pain  
562 hypersensitivity by central neural plasticity. *Journal of Pain* 10, 895–926.
- 563 Li, C.-L., Li, K.-C., Wu, D., Chen, Y., Luo, H., Zhao, J.-R., Wang, S.-S., Sun, M.-M., Lu, Y.-  
564 J., Zhong, Y.-Q., Hu, X.-Y., Hou, R., Zhou, B.-B., Bao, L., Xiao, H.-S., Zhang, X.,  
565 2016. Somatosensory neuron types identified by high-coverage single-cell RNA-  
566 sequencing and functional heterogeneity. *Cell Research* 26, 83–102.
- 567 Li, C.-Y., Zhang, X.-L., Matthews, E.A., Li, K.-W., Kurwa, A., Boroujerdi, A., Gross, J.,  
568 Gold, M.S., Dickenson, A.H., Feng, G., Luo, Z.D., 2006. Calcium Channel  $\alpha 2\delta 1$   
569 Subunit Mediates Spinal Hyperexcitability in Pain Modulation. *Pain* 125, 20–34.
- 570 Lu, Y., Dong, H., Gao, Y., Gong, Y., Ren, Y., Gu, N., Zhou, S., Xia, N., Sun, Y.-Y., Ji, R.-  
571 R., Xiong, L., 2013. A feed-forward spinal cord glycinergic neural circuit gates  
572 mechanical allodynia. *Journal of Clinical Investigation* 123, 4050–4062.
- 573 Lu, Y., Perl, E.R., 2003. A Specific Inhibitory Pathway between Substantia Gelatinosa  
574 Neurons Receiving Direct C-Fiber Input. *Journal of Neuroscience* 23, 8752–8758.
- 575 Lumpkin, E.A., Caterina, M.J., 2007. Mechanisms of sensory transduction in the skin. *Nature*  
576 445, 858–865.
- 577 Mantyh, P.W., Rogers, S.D., Honore, P., Allen, B.J., Ghilardi, J.R., Li, J., Daughters, R.S.,  
578 Lappi, D.A., Wiley, R.G., Simone, D.A., 1997. Inhibition of hyperalgesia by ablation  
579 of lamina I spinal neurons expressing the substance P receptor. *Science* 278, 275–279.
- 580 McMahon, S.B., Malcangio, M., 2009. Current Challenges in Glia-Pain Biology. *Neuron* 64,  
581 46–54.
- 582 Melzack, R., Wall, P.D., 1965. Pain mechanisms: a new theory. *Science* 150, 971–979.
- 583 Mendell, L.M., 2014. Constructing and deconstructing the gate theory of pain. *PAIN* 155,  
584 210–216.
- 585 Mishra, S.K., Hoon, M.A., 2013. The Cells and Circuitry for Itch Responses in Mice. *Science*  
586 340, 968–971.
- 587 Moayedi, M., Davis, K.D., 2013. Theories of pain: from specificity to gate control. *Journal of*  
588 *Neurophysiology* 109, 5–12.
- 589 Mogil, J.S., 2012. Sex differences in pain and pain inhibition: multiple explanations of a  
590 controversial phenomenon. *Nature Reviews. Neuroscience* 13, 859–866.
- 591 Muñana, K.R., Zhang, D., Patterson, E.E., 2010. Placebo effect in canine epilepsy trials.  
592 *Journal of Veterinary Internal Medicine*. 24, 166–170.
- 593 Noël, J., Zimmermann, K., Busserolles, J., Deval, E., Alloui, A., Diochot, S., Guy, N.,  
594 Borsotto, M., Reeh, P., Eschalièr, A., Lazdunski, M., 2009. The mechano-activated  
595 K<sup>+</sup> channels TRAAK and TREK-1 control both warm and cold perception. *EMBO*  
596 *Journal* 28, 1308–1318.
- 597 Omerbašić, D., Schuhmacher, L.-N., Bernal Sierra, Y.-A., Smith, E.S.J., Lewin, G.R., 2015.  
598 ASICs and mammalian mechanoreceptor function. *Neuropharmacology, Acid-*  
599 *Sensing Ion Channels in the Nervous System* 94, 80–86.



- 600 Pan, H.-L., Wu, Z.-Z., Zhou, H.-Y., Chen, S.-R., Zhang, H.-M., Li, D.-P., 2008. Modulation  
601 of Pain Transmission by G Protein-Coupled Receptors. *Pharmacology and*  
602 *Therapeutics* 117, 141–161.
- 603 Peirs, C., Seal, R.P., 2016. Neural circuits for pain: Recent advances and current views.  
604 *Science* 354, 578–584.
- 605 Peirs, C., Williams, S.-P.G., Zhao, X., Walsh, C.E., Gedeon, J.Y., Cagle, N.E., Goldring,  
606 A.C., Hioki, H., Liu, Z., Marell, P.S., Seal, R.P., 2015. Dorsal Horn Circuits for  
607 Persistent Mechanical Pain. *Neuron* 87, 797–812.
- 608 Pethő, G., Reeh, P.W., 2012. Sensory and Signaling Mechanisms of Bradykinin, Eicosanoids,  
609 Platelet-Activating Factor, and Nitric Oxide in Peripheral Nociceptors. *Physiological*  
610 *Reviews* 92, 1699–1775.
- 611 Petitjean, H., Pawlowski, S.A., Fraine, S.L., Sharif, B., Hamad, D., Fatima, T., Berg, J.,  
612 Brown, C.M., Jan, L.-Y., Ribeiro-da-Silva, A., Braz, J.M., Basbaum, A.I., Sharif-  
613 Naeni, R., 2015. Dorsal Horn Parvalbumin Neurons Are Gate-Keepers of Touch-  
614 Evoked Pain after Nerve Injury. *Cell Reports* 13, 1246–1257.
- 615 Polgár, E., Sardella, T.C.P., Tiong, S.Y.X., Locke, S., Watanabe, M., Todd, A.J., 2013.  
616 Functional differences between neurochemically defined populations of inhibitory  
617 interneurons in the rat spinal dorsal horn. *PAIN* 154, 2606–2615.
- 618 Rausch-Derra, L., Huebner, M., Wofford, J., Rhodes, L., 2016. A Prospective, Randomized,  
619 Masked, Placebo-Controlled Multisite Clinical Study of Grapiprant, an EP4  
620 Prostaglandin Receptor Antagonist (PRA), in Dogs with Osteoarthritis. *Journal of*  
621 *Veterinary Internal Medicine*. 30, 756–763.
- 622 Rexed, B., 1952. The cytoarchitectonic organization of the spinal cord in the cat. *Journal of*  
623 *Comparative Neurology* 96, 415–495.
- 624 Ribeiro-da-Silva, A., Coimbra, A., 1982. Two types of synaptic glomeruli and their  
625 distribution in laminae I-III of the rat spinal cord. *Journal of Comparative Neurology*  
626 209, 176–186.
- 627 Ringkamp, M., Srinivasa, R., Campbell, J., Meyer, R., 2013. Peripheral Mechanisms of  
628 Cutaneous Nociception. In: Wall and Melzack's Textbook of Pain (Editors McMahon,  
629 Koltzenburg, Tracey, and Turk). Elsevier, pp. 1–30.
- 630 Robinson, D.R., Gebhart, G.F., 2008. Inside information: the unique features of visceral  
631 sensation. *Molecular Interventions*. 8, 242–253.
- 632 Sandkühler, J., 2009. Models and Mechanisms of Hyperalgesia and Allodynia. *Physiological*  
633 *Reviews* 89, 707–758.
- 634 Sandkühler, J., Gruber-Schoffnegger, D., 2012. Hyperalgesia by synaptic long-term  
635 potentiation (LTP): an update. *Current Opinion Pharmacology* 12, 18–27.
- 636 Seal, R.P., Wang, X., Guan, Y., Raja, S.N., Woodbury, C.J., Basbaum, A.I., Edwards, R.H.,  
637 2009. Injury-induced mechanical hypersensitivity requires C-low threshold  
638 mechanoreceptors. *Nature* 462, 651–655.
- 639 Segerdahl, A.R., Mezue, M., Okell, T.W., Farrar, J.T., Tracey, I., 2015. The dorsal posterior  
640 insula subserves a fundamental role in human pain. *Nature Neuroscience* 18, 499–  
641 500.
- 642 Sikandar, S., Ronga, I., Iannetti, G.D., Dickenson, A.H., 2013. Neural coding of nociceptive  
643 stimuli—from rat spinal neurones to human perception. *Pain* 154, 1263–1273.

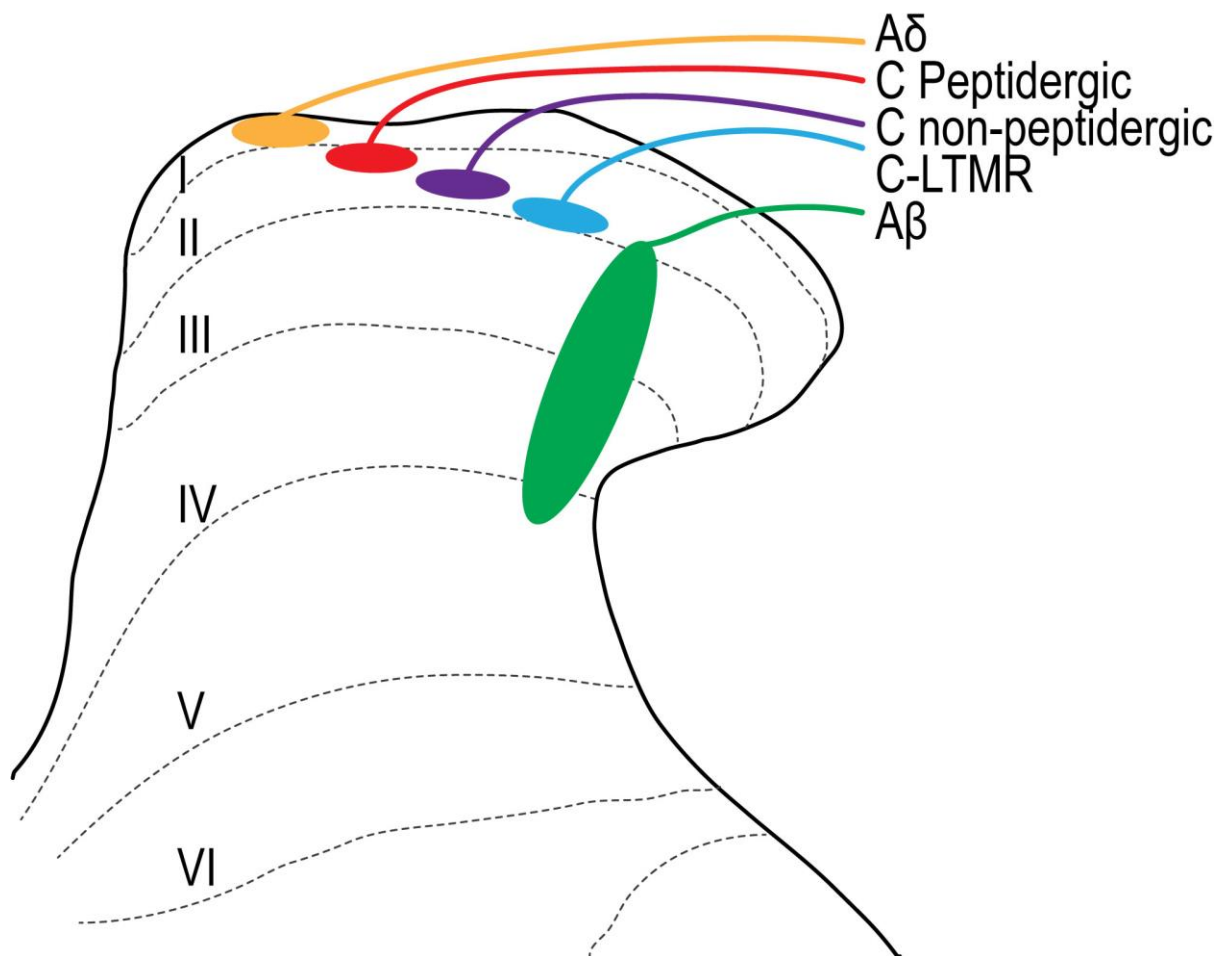
- 644 Snider, W.D., McMahon, S.B., 1998. Tackling Pain at the Source: New Ideas about  
645 Nociceptors. *Neuron* 20, 629–632.
- 646 Spike, R.C., Puskár, Z., Andrew, D., Todd, A.J., 2003. A quantitative and morphological  
647 study of projection neurons in lamina I of the rat lumbar spinal cord. *European*  
648 *Journal of Neuroscience* 18, 2433–2448.
- 649 Thompson, S.J., Bushnell, M.C., 2012. Rodent functional and anatomical imaging of pain.  
650 *Neuroscience Letters*, Neuroimaging of Pain: Insights into normal and pathological  
651 pain mechanisms 520, 131–139.
- 652 Todd, A.J., 2010. Neuronal circuitry for pain processing in the dorsal horn. *Nature Reviews*  
653 *Neuroscience* 11, 823–836.
- 654 Todd, A.J., 2017. Identifying functional populations among the interneurons in laminae I-III  
655 of the spinal dorsal horn. *Molecular Pain* 13, 1744806917693003.
- 656 Tracey, I., 2010. Getting the pain you expect: mechanisms of placebo, nocebo and reappraisal  
657 effects in humans. *Nature Medicine* 16, 1277.
- 658 Tracey, I., Johns, E., 2010. The pain matrix: reloaded or reborn as we image tonic pain using  
659 arterial spin labelling. *Pain* 148, 359–360.
- 660 Tracey, I., Mantyh, P.W., 2007. The Cerebral Signature for Pain Perception and Its  
661 Modulation. *Neuron* 55, 377–391.
- 662 Treede, R.D., Meyer, R.A., Campbell, J.N., 1998. Myelinated mechanically insensitive  
663 afferents from monkey hairy skin: heat-response properties. *Journal of*  
664 *Neurophysiology* 80, 1082–1093.
- 665 Usoskin, D., Furlan, A., Islam, S., Abdo, H., Lönnberg, P., Lou, D., Hjerling-Leffler, J.,  
666 Haeggström, J., Kharchenko, O., Kharchenko, P.V., Linnarsson, S., Ernfors, P., 2014.  
667 Unbiased classification of sensory neuron types by large-scale single-cell RNA  
668 sequencing. *Nature Neuroscience* 18, 145–153.
- 669 van Loon, J.P. a. M., de Grauw, J.C., van Dierendonck, M., L'ami, J.J., Back, W., van  
670 Weeren, P.R., 2010. Intra-articular opioid analgesia is effective in reducing pain and  
671 inflammation in an equine LPS induced synovitis model. *Equine Veterinary Journal*  
672 42, 412–419.
- 673 Vrontou, S., Wong, A.M., Rau, K.K., Koerber, H.R., Anderson, D.J., 2013. Genetic  
674 identification of C fibres that detect massage-like stroking of hairy skin in vivo.  
675 *Nature* 493, 669–673.
- 676 West, S.J., Bannister, K., Dickenson, A.H., Bennett, D.L., 2015. Circuitry and plasticity of  
677 the dorsal horn – Toward a better understanding of neuropathic pain. *Neuroscience*  
678 300, 254–275.
- 679 Williams, A.C., Craig, K.D., 2016. Updating the definition of pain. *Pain* 157, 2420-2423.
- 680 Woolf, C.J., 2011. Central sensitization: implications for the diagnosis and treatment of pain.  
681 *Pain* 152, S2-15.
- 682 Yaksh, T.L., Woller, S.A., Ramachandran, R., Sorkin, L.S., 2015. The search for novel  
683 analgesics: targets and mechanisms. *F1000Prime Reports* 7, 56.
- 684 Yarnitsky, D., 2010. Conditioned pain modulation (the diffuse noxious inhibitory control-like  
685 effect): its relevance for acute and chronic pain states. *Current Opinion in*  
686 *Anaesthesiology* 23, 611–615.

- 687 Yasaka, T., Kato, G., Furue, H., Rashid, M.H., Sonohata, M., Tamae, A., Murata, Y.,  
688 Masuko, S., Yoshimura, M., 2007. Cell-type-specific excitatory and inhibitory circuits  
689 involving primary afferents in the substantia gelatinosa of the rat spinal dorsal horn in  
690 vitro. *Journal of Physiology* 581, 603–618.
- 691 Yasaka, T., Tiong, S.Y.X., Hughes, D.I., Riddell, J.S., Todd, A.J., 2010. Populations of  
692 inhibitory and excitatory interneurons in lamina II of the adult rat spinal dorsal horn  
693 revealed by a combined electrophysiological and anatomical approach. *Pain* 151,  
694 475–488.
- 695 Zeilhofer, H.U., Wildner, H., Yevenes, G.E., 2012. Fast Synaptic Inhibition in Spinal Sensory  
696 Processing and Pain Control. *Physiological Reviews* 92, 193–235.
- 697 Zhang, J., Cavanaugh, D.J., Nemenov, M.I., Basbaum, A.I., 2013. The modality-specific  
698 contribution of peptidergic and non-peptidergic nociceptors is manifest at the level of  
699 dorsal horn nociresponsive neurons. *The Journal of Physiology* 591, 1097–1110.
- 700 Zimmermann, K., Leffler, A., Babes, A., Cendan, C.M., Carr, R.W., Kobayashi, J., Nau, C.,  
701 Wood, J.N., Reeh, P.W., 2007. Sensory neuron sodium channel Nav1.8 is essential for  
702 pain at low temperatures. *Nature* 447, 856–859.

703 **Figure Legends**

704 *Fig 1.*

705 *The dorsal horn of the spinal cord is divided into laminae. Different classes of cutaneous*  
706 *primary afferents have specific patterns of innervation of the dorsal horn. The figure*  
707 *demonstrates the stratification of the dorsal horn by somatosensory modality with those*  
708 *afferents carrying impulses resulting from noxious cutaneous stimuli primarily innervating*  
709 *laminae I and II, and those conveying touch signals projecting to the deeper laminae. (C-*  
710 *LTMR stands for C-low threshold mechanoreceptor.)*

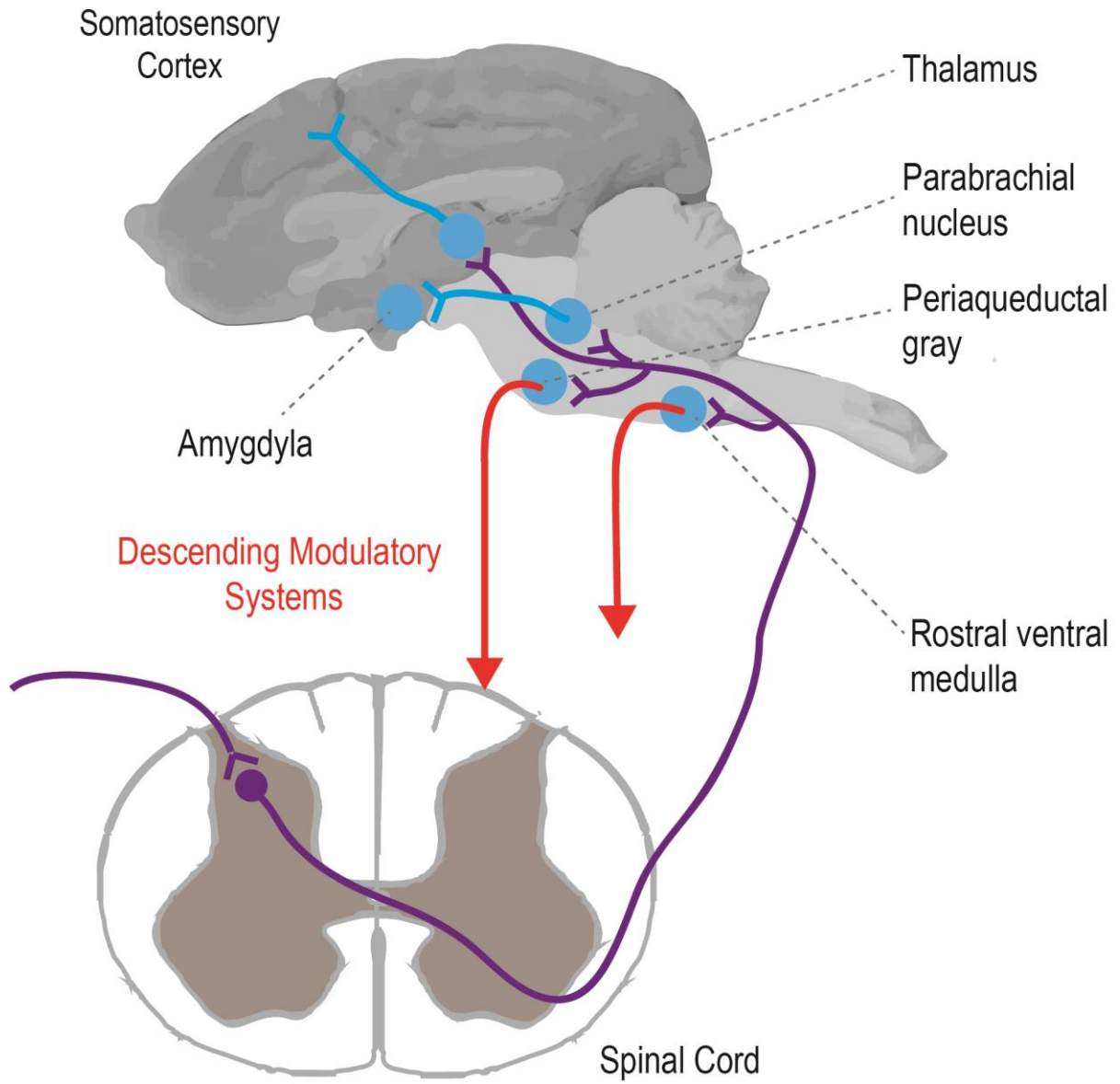


711

712 *Fig 2.*

713 *Projection neurons arise from the dorsal horn of the spinal cord. These innervate various*  
714 *higher centres responsible for aspects of the pain experience including the thalamus and the*  
715 *parabrachial nucleus. Neurons of the ventral medulla and midbrain periaqueductal gray are*  
716 *also engaged which may activate descending feedback systems. The simplified figure shows*  
717 *the sites of primary terminations from projection neurons. Initial activation of these areas*  
718 *forms the basis of a resultant coordinated activity in a multitude of brain regions which*  
719 *comprise the 'pain matrix'.*

720 The figure shows a single projection neuron, in this case a spinothalamic tract (STT) neuron.  
721 Note - species differences exist in the white matter tracts for the projection of nociceptive  
722 information as detailed in the text.



723

724 **Tables**

725 Table 1 – The classification of primary afferents by fibre diameter and conduction velocity.

<b>Classification</b>	<b>Diameter</b>	<b>Myelin</b>	<b>Conduction velocity</b>	<b>Sensory Function</b>
A $\beta$	Large (6–12 $\mu\text{m}$ )	Yes	>35 m/s	Touch
A $\delta$	Medium (1–5 $\mu\text{m}$ )	Thin	5-35 m/s	‘Fast’ pain
C	Small (0.2-1.5 $\mu\text{m}$ )	No	<2.0 m/s	‘Slow’ pain

726