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1 Long-term microplastic retention causes reduced body condition in the
2 Langoustine, *Nephrops norvegicus*

3

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11

12 **Abstract**

13 Microplastic represents a rising proportion of marine litter and is widely distributed throughout
14 a range of marine habitats. Correspondingly, the number of reports of species containing
15 microplastics increases annually. *Nephrops norvegicus* in the Firth of Clyde have previously been
16 shown to retain large aggregations of microplastic fibres. The potential for *N. norvegicus* to retain
17 plastic over an extended time period increases the likelihood of any associated negative impacts
18 to the individual. This study represents the longest observation of the impacts of microplastic
19 retention in invertebrates. We exposed *N. norvegicus* to plastic over eight months to determine
20 the impacts of extended exposure. Over this period we compared the feeding rate, body
21 mass, and nutritional state of plastic-fed *N. norvegicus* to that of fed and starved control
22 groups. Following the experimental period, the plastic-fed langoustine contained microplastic
23 aggregations comparable to those of small individuals from the Clyde Sea Area. Comparisons
24 between fed, unfed and plastic-fed individuals indicated a reduction in feeding rate, body mass,
25 and metabolic rate as well as catabolism of stored lipids in plastic contaminated animals. We
26 conclude that *N. norvegicus* exposed to high levels of environmental microplastic pollution may
27 experience reduced nutrient availability. This can result in reduced population stability and may
28 affect the viability of local fisheries.

29 *Capsule: Long term retention of microplastic aggregations reduces the nutritional state of*
30 *langoustine.*

31

32 **Keywords:** Microplastic, Nutrition, *Nephrops norvegicus*, feeding rate

33

34 **Introduction**

35 Uptake of plastic debris has been recorded in numerous vertebrate taxa, including birds (Burger
36 and Gochfeld, 2004; Ryan, 2008), fish (Lusher et al., 2013), and mammals (Baird and Hooker,
37 2000). However, as this large plastic debris breaks down it forms microplastic (<5mm), which
38 represents an increasing proportion of global marine litter (Barnes et al., 2009). This increase has
39 resulted in a rise in the number of records of plastic uptake by wild-caught invertebrates
40 (Devriese et al., 2015; Mathalon and Hill, 2014; Murray and Cowie, 2011; Welden and Cowie,
41 2016). The uptake of microplastics may occur through a variety of routes, including active
42 ingestion, a result of mistaking plastics for prey items; or passively, through contaminated prey
43 and/or sediment. Both of these routes have been observed in laboratory experiments (Bern,
44 1990; Besseling et al., 2012; Farrell and Nelson, 2013; Frias et al., 2014). Uptake has even been
45 observed through the gills during respiration (Watts et al., 2014b).

46 *The Impacts of Plastic Ingestion*

47 The ingestion of plastic by marine vertebrates has been seen to have a range of effects on the
48 organism. One of the most apparent impacts is mortality due to gut damage as observed in turtles
49 (Lutz, 1990; Tomás et al., 2002) and porpoises (Baird and Hooker, 2000), or starvation as a result
50 of plastic taking up the space of food in the stomach. Damage to the gut may also result in reduced
51 digestive efficiency and nutrient uptake, known as nutrient dilution (McCauley and Bjorndal,
52 1999). In birds, this has been linked to reduced body condition as contaminated individuals utilise
53 the body's stores to bridge the energy gap (Connors and Smith, 1982; Pierce et al., 2004; Ryan,
54 1988). Finally, animals may also be subject to the transfer of hydrophobic contaminants, as
55 previously observed in seabirds (Ryan et al., 1988). This is particularly concerning in areas where
56 plastics exposed to sea water containing hydrophobic contaminants may develop loads much
57 higher than that of the surrounding water (Endo et al., 2005; Teuten et al., 2009).

58 *Microplastic and Invertebrates*

59 Despite the previous work examining the impacts of plastic debris on both wild and captive
60 vertebrates, the effect of microplastic ingestion by invertebrates have yet to be determined. Of
61 those studies that address these impacts, most deal with short-term effects. For example,
62 microplastic consumption has been seen to reduce feeding in *Arenicola marina* (Besseling et al.,
63 2012; Wright et al., 2013) and shore crabs, *Carcinus maenas*, fed on chronic levels of plastic fibres
64 over four week periods showed a dramatic reduction in both feeding and scope for growth (Watts
65 et al., 2015). *A. marina* has also been seen to take up hydrophobic contaminants from plastics,
66 which may result in harmful effects to the animal (Besseling et al., 2012). Responses have also

67 been observed in terrestrial invertebrates. Examination of the impacts of polyethylene powder
68 (< 400µm) on the worm, *Lumbricus terrestris*, indicated reduced growth rate at all microplastic
69 concentrations and increased mortality at concentrations over 28% (Huerta Lwanga et al., 2016).

70 A small number of species have demonstrated minimal or no effects as the result of plastic
71 ingestion. The larvae of the sea urchin *Tripneustes gratilla* were seen to readily ingest
72 polyethylene spheres; however, this ingestion was not seen to significantly increase mortality,
73 despite a slight reduction in survivorship in the highest concentration (Kaposi et al., 2014). The
74 marine isopod *Idotea emarginata* also did not discriminate against plastics, ingesting beads,
75 fragments, and fibres. The isopods were able to pass ingested microplastic, with no significant
76 level of accumulation observed; as a result, it was determined that microplastic ingestion had no
77 negative impact on the health of the organism (Hämer et al., 2014). Such variation in the impact
78 of microplastic ingestion between species is to be expected. Biotic factors, such as microplastic
79 uptake rate, residence time of microplastic in the gut, and tolerance to low nutrient conditions
80 each influence the cumulative impact of exposure.

81 *Microplastic Uptake and Retention by N. norvegicus*

82 *Nephrops norvegicus* are decapod crustaceans found in fine sediments at depths between 20 and
83 800 meters across the Northeast Atlantic and in the Mediterranean. Their diet is mainly
84 composed of bivalve molluscs, polychaetes, echinoderms, fish, and crustaceans including
85 conspecifics (Aguzzi and Sardà, 2008). *N. norvegicus* recovered from the Clyde Sea Area have been
86 seen to contain large aggregations of microplastic fibres (Murray and Cowie, 2011; Welden and
87 Cowie, 2016), and may be at increased risk of the negative impacts of microplastic uptake. These
88 large aggregations are believed to be the result of the complex gut morphology of *N. norvegicus*.
89 The digestive system has three main portions, the foregut, mid gut and narrow hind gut, and a set
90 of chitinous structures known as the gastric mill (Welden et al., 2015). Examination of wild caught
91 *N. norvegicus* has shown microplastic aggregations directly anterior to the gastric mill and the
92 narrowing at the entrance to the hind gut; it is thought that these structures prevent some
93 microplastics from leaving the digestive tract (Welden and Cowie, 2016).

94 It has been seen that *N. norvegicus* may lose aggregated microplastic during moult, as the foregut
95 lining is shed (Welden and Cowie, 2016). *N. norvegicus* moult occurs biannually in male and
96 immature individuals, and annually in mature females. This extended period over which they may
97 retain microplastics suggests that the species are at increased risk of the negative impacts of
98 plastic ingestion. For *N. norvegicus* populations in areas of high environmental microplastic
99 contamination individuals may develop large aggregations of plastic fibres in the fore-gut, which
100 increases the likelihood and magnitude of these effects (Welden and Cowie, 2016).

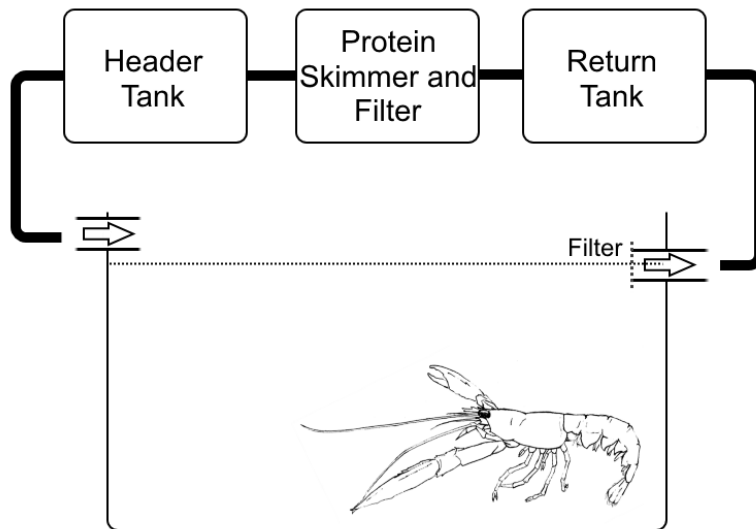
101 It is hypothesised that the level of microplastic contamination observed in *N. norvegicus* from the
102 Clyde Sea Area may result in numerous impacts on the nutritional state of *N. norvegicus*, with the
103 potential for false satiation and starvation in highly affected individuals. In this paper we aim to
104 assess the impact of large fibre aggregations on nutritional health and mortality of this
105 economically important population. In an eight month exposure trial, the impacts of long-term
106 exposure to microplastics on food consumption and growth in male *N. norvegicus* was examined.

107 **Methods**

108 Male *N. norvegicus* were sampled from the Clyde Sea Area in early spring using 50 mm mesh otter
109 trawls. Prior to the experiment, captured animals were kept in holding tanks (270 litres) supplied
110 with recirculating sea water, and were fed on whole squid mantle that had been rinsed thoroughly
111 with distilled water to remove any attached microplastic. After the group had moulted, 34
112 individuals were sacrificed for gut content analysis to confirm that the animals were free from
113 plastics, and 36 were transferred to lidded individual tanks (10 litres). Each tank contained rock
114 shelters, and was maintained at ambient temperature with a 12 hour light/dark regime.

115 A closed 68 litre header tank was used to circulate filtered seawater to prevent introduction of
116 foreign microplastics. Water leaving tanks passed through a 0.2mm filter to prevent plastics from
117 the treatment group being passed to control animals, and all water flowed through a secondary
118 protein skimmer and filter before re-entering the header tank (Fig. 1). The system was reduced
119 to 50% volume before being topped up with filtered seawater approximately every 2 weeks; more
120 frequently during warmer weather. Water levels were monitored on a daily basis and ad hoc
121 additions occasionally made to maintain constant volume. Air-stones and pumps were used to
122 ensure sufficient oxygenation of the water and slate shelters were provided for cover.

123



124

125 Fig. 1. Individual tank set up with model of sea water circulation system

126 After a month-long acclimatisation period in the experimental tanks, the carapace length and
 127 body mass of each individual was recorded. The average carapace length at day 0 was 31.357 mm,
 128 and the average mass was 19.28 g. Total haemolymph protein was also determined using a
 129 Bradford assay, described in full below.

130 The 36 individuals were then divided into three groups (12 individuals per group): the treatment
 131 group, fed a 1.5 g squid mantle seeded with five polypropylene fibres; the fed control group, fed
 132 with 1.5 g of squid mantle only; and the starved control group. Fibres were chosen for this
 133 experiment as they were the most abundant category of microplastic recovered in previous
 134 studies of ingestion by *N. norvegicus*. Similarly, they are the dominant microplastic category
 135 observed in environmental sampling.

136 Polypropylene rope was chosen owing to its widespread use in the fishing industry and
 137 prevalence in previously studied *N. norvegicus* from the Clyde Sea Area (Welden and Cowie,
 138 2016). Fibres were removed from twisted split film polypropylene rope supplied by Gaelforce.
 139 Individual fibres measured between 3 mm and 5 mm in length and were approximately 0.2 mm
 140 in diameter. *N. norvegicus* in the Clyde Sea Area are known to be subject to high aggregations of
 141 microplastic. Five fibres per feeding would expose the plastic-fed group to 360 fibres over the
 142 experimental period, although not all were expected to be ingested and it is uncertain what
 143 proportion of ingested plastic is retained in the foregut.

144 Previous observations of nutritional state in *N. norvegicus* have indicated that animals can survive
 145 for long periods without food (Watts et al., 2014a). The starved control was used to provide a
 146 baseline for reduced nutritional health. Over the eight month experimental period the treatment

147 group and fed control were fed every two days, and feeding rate in both groups was determined
148 by weighing the un-eaten food. After eight months, a second haemolymph sample was taken from
149 each animal, after which the animal was killed and immediately dissected; the stomach was
150 removed and transferred directly into 80% ethanol, and the hepatopancreas removed, weighed,
151 and frozen at -80°C.

152 *Foregut Microplastics*

153 Microplastic retention was determined by examining the foregut of each individual under stereo
154 microscope, as outlined in Murray and Cowie (2011) and Welden and Cowie (2016). Plastic
155 recovered were visually examined to determine that they originated from the experimental
156 condition rather than prior exposure. The weight of retained microplastic was recorded using a
157 Mettler MX5 balance (Mettler-Toledo international Inc., Columbus, USA).

158 *Bradford Assay*

159 Changes in the composition of the haemolymph and the level of stored lipids were used as
160 indicators of nutritional state. Reduction in haemolymph protein has been linked to metabolic
161 depression (Watts et al., 2014a). The results of the Bradford assay obtained following the
162 acclimatisation period were compared to a second assay carried out on haemolymph samples
163 taken at the end of the experimental period. The method followed that outlined by Hagerman
164 (1983), with 10 µl of haemolymph diluted with 990 µl of deionised water. 50 µl of the diluted
165 sample was then added to 950 µl of coomassie blue. The absorbance of the resulting solution was
166 determined at 562 nm using a spectrophotometer, calibrated using standardised solutions of
167 bovine serum albumen (BSA)(Hagerman, 1983).

168 *Copper in the Hepatopancreas*

169 The breakdown of haemocyanin, the protein responsible for oxygen transport, results in an
170 increase in copper levels in the hepatopancreas. To determine the level of copper in the
171 hepatopancreas, dehydrated tissue samples were subjected to atomic absorption spectrometry
172 (AAS). Hepatopancreas samples were freeze dried over five days and the dry weight recorded.
173 100 mg of the dry tissue was pre-digested at 95°C using 8 ml of nitric acid. The resulting material
174 was allowed to cool for 10 minutes, following which 3 ml of hydrogen peroxide were added. After
175 a minimum of 8 hours, the samples were made up to 10 ml with distilled water, and analysed
176 using an AA Analyst400 (Perkin Elmer Ltd, Cambridge, UK). Results were compared to standards
177 of copper nitrate (Sigma Aldrich) diluted to concentrations of 15, 10, 5, 2.5 and 1.25 ppm and a
178 distilled water blank (Watts et al., 2014a).

179 *Mass and Lipid Content of the Hepatopancreas*

180 Reduction of energy stores is an indication that the level of starvation exceeds that which can be
181 managed by metabolic depression alone. In *N. norvegicus*, the greatest concentration of stored
182 lipids is found in the hepatopancreas; as these are catabolised they are replaced with water to
183 maintain tissue volume. In a study of the nutritional value of pelleted and natural food sources
184 carried out by Mente (2010), the starved control group exhibited a reduction in lipid
185 concentration of 12.16% in over 8 months. Two indices were used as indicators of catabolism of
186 stored lipids. Hepatosomatic Index (HSI), which is the mass of the hepatopancreas in relation to
187 overall body mass (Mayrand and Dutil, 2008), and the percentage of water in the hepatopancreas
188 (% H₂O HP). HSI was calculated using the mass of the hepatopancreas recorded at dissection and
189 the final body weight of the animal. The percentage water of the hepatopancreas was calculated
190 from the final weight of the hepatopancreas and the weight of the freeze dried hepatopancreas
191 samples.

192 *Statistical Analysis*

193 Statistical analysis was carried out using Minitab15. Differences in food consumption between
194 the plastic-fed treatment group and the fed control group were examined at each month using a
195 Mann-Whitney U analysis. Comparisons of the aggregation of gut plastic, variation in body mass,
196 and the various indices of nutritional state between the three treatment groups at month eight
197 were conducted using a Kruskal-Wallis test. In the event of a significant result, the relationship
198 was explored using post hoc Mann-Whitney tests to determine the treatment group responsible
199 for the response.

200

201 **Results and Discussion**

202 Analysis of the gut content of plastic-fed individuals revealed aggregations weighing between
203 0.41 – 3.49 mg, (average 1.5 mg). One of the control animals was also observed to have plastic in
204 the foregut. This was a single pink fibre, distinctly different to those provided to the treatment
205 group. Despite this, there remained a highly significant difference in the level of contamination
206 between the three groups ($H = 16.77$, d.f. = 2, $P < 0.001$). The levels of plastic observed in the
207 treatment group at the end of the exposure period are comparable to aggregations found in highly
208 contaminated individuals from the Clyde Sea Area (Welden and Cowie, 2016). This comparable
209 level of plastic retention indicates that the effects outlined below are representative of those
210 experienced by animals in areas of high microplastic contamination.

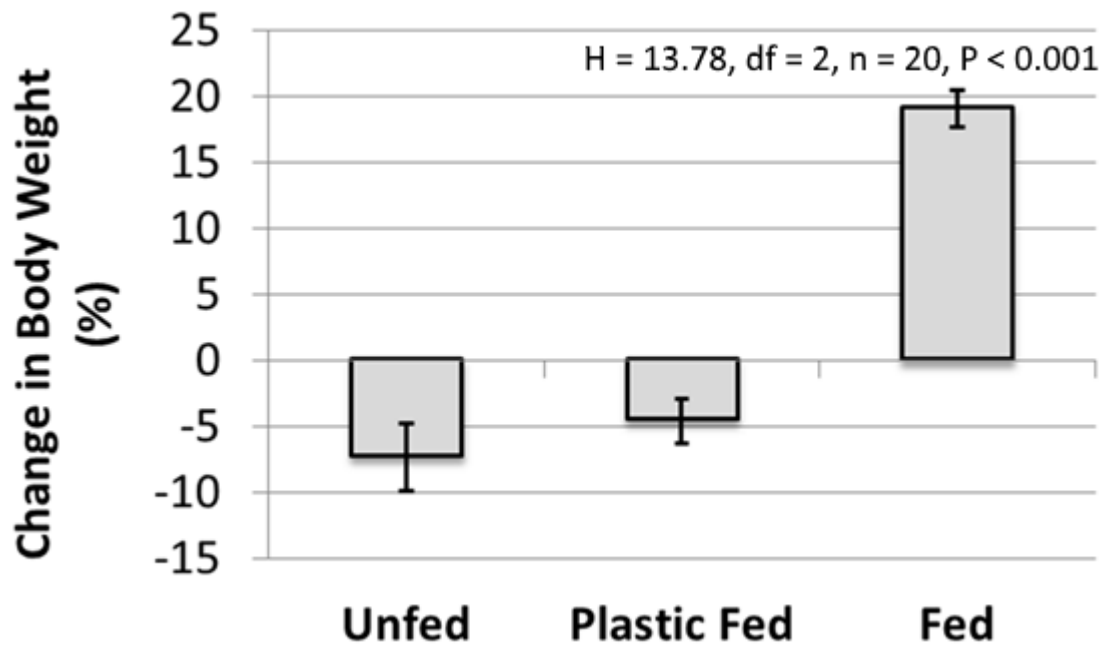
211 Survivorship varied between treatments groups, with the starved condition displaying the
212 highest mortality (58.3%), followed by plastic fed (41.6%), and then fed individuals (33.2%). The
213 mortality rate of plastic fed animals fell between the two control conditions, indicating that *N.*
214 *Norvegicus* were weakened by the presence of plastic in the diet. Mortality in the fed control was
215 higher than that which might be expected. It may be that *N. norvegicus* are less able to cope with
216 starvation under ambient temperature conditions than under a lower temperature regime, or
217 that the group were subject to an unidentified stressor, such as the presence of a pathogen.

218 Ingestion of large plastic debris has previously been identified as a cause of increased mortality
219 in both turtles (Lutz, 1990; Tomás et al., 2002) and cetaceans (Baird and Hooker, 2000), as a result
220 of blockage of the gut. Similarly, worms that were fed microplastics have also demonstrated an
221 increased mortality rate (Huerta Lwanga et al., 2016). The increased mortality in the plastic-fed
222 condition may be the result of starvation caused by reduced nutrient availability, or of direct
223 damage to the organism. However, as part of its natural diet *N. norvegicus* regularly ingest items
224 that are of irregular size and shape, which have equal potential to damage the animal's digestive
225 tract. As a result, reduced nutrient availability is considered the most likely factor influencing the
226 observed increase in mortality of the plastic fed animals.

227 *The effect of microplastic ingestion on growth*

228 Over the eight month survey period significant differences were observed in the body mass of the
229 three treatment groups, and statistical analysis revealed a significant difference between the
230 conditions ($H = 13.78$, d.f. = 2, $P < 0.001$) (Fig. 2). The normally fed control exhibited a mean
231 increase in body mass of 0.0795% per day, while starved individuals demonstrated a mean
232 decrease in body mass of -0.0303% per day. Animals fed microplastics showed a decrease in mean
233 body mass of -0.0189% per day, falling between the two control treatments.

234 As scope for growth is determined by food availability it is assumed that the decrease in body
235 mass of the plastic fed individuals is the result of lowered nutrient uptake. Reduction in the body
236 mass of both the starved and plastic fed groups occur as a result of the body's stores being
237 metabolised in the absence of nutrition from food sources (Connors and Smith, 1982). The mean
238 reduction in body mass recorded in plastic-fed individuals was not as marked as that in the
239 starved control, suggesting that a proportion of the nutrients consumed by the treatment group
240 were successfully absorbed. No significant correlation was found between the mass of plastic in
241 the gut and the reduction in body mass, suggesting that there is not a direct relationship between
242 the two; however, the sample size in this study is small, and a relationship may become apparent
243 in a larger experiment using more *N. norvegicus*.



244

245 Fig. 2. Mean change in body mass of animals from the three experimental groups observed at month eight
 246 (bars indicate standard error)

247

248 *Variation in Feeding Rate*

249 During the experimental period, the feeding rates of both the fed and the plastic fed groups were
 250 seen to vary on a monthly basis; however, the mass of food consumed by plastic-fed animals
 251 decreased in relation to that of the fed control. Over the 8-month experimental period, a decline
 252 in the mass of consumed food could be seen in the plastic-fed group (Fig. 3), indicating reduced
 253 feeding in plastic-fed individuals. However, this effect was not seen to be significant. ($W = 44.0, P$
 254 < 0.1824).

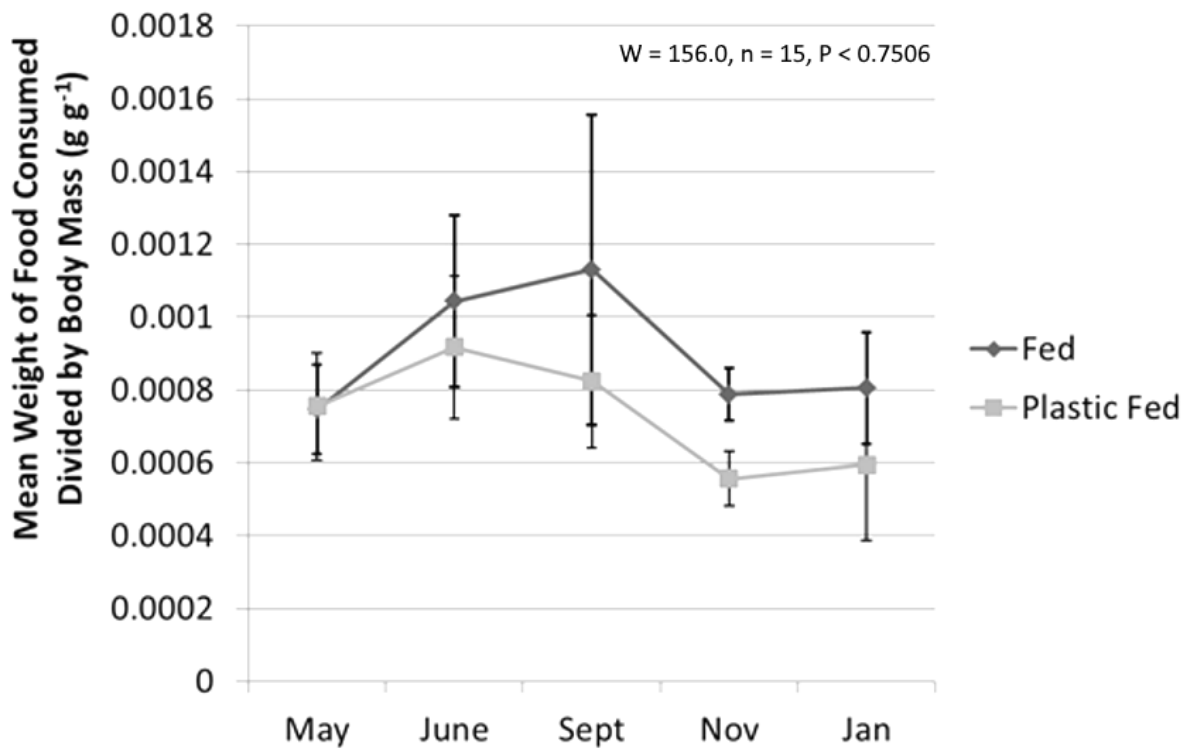
255 These results support previous observations of reduced feeding in microplastic-fed *C. maenas*
 256 reported by Watts et al., (2015), and microsphere-fed *A. marina* (Besseling et al., 2012) and *D.*
 257 *manga* (Besseling et al., 2014). The reduction in feeding rate is believed to be the result of false
 258 satiation, as plastic aggregations take up an increasing volume in the stomach. This effect has
 259 been observed in a number of vertebrate species, particularly birds (Azzarello and Fleet, 1987;
 260 Connors and Smith, 1982; Pierce et al., 2004; Ryan and Jackson, 1987).

261 Sustained reduction in feeding is known to result in reduced body condition (Watts et al., 2014a).
 262 However, it is not clear whether the reduced growth rate and increased mortality described
 263 above are solely the result of the decreased feeding rate or whether plastic in the gut also reduces

264 nutrient uptake by the digestive tract. Damage to or irritation of the gut membrane may reduce
265 the efficiency with which digested food is absorbed, reducing the nutritive value of the food that
266 is ingested.

267 For some species, microplastic ingestion may not result in false satiation. Organisms that
268 regularly take in indigestible material may possess compensatory mechanisms. Oysters exposed
269 to microspheres were seen to dramatically increase their food uptake, apparently in an attempt
270 to compensate for the ingestion of plastic (Sussarellu et al., 2014).

271



272

273 Fig. 3. Mean weight of squid mantle consumed by the treatment group and fed control over the
274 experimental period (bars indicate standard error)

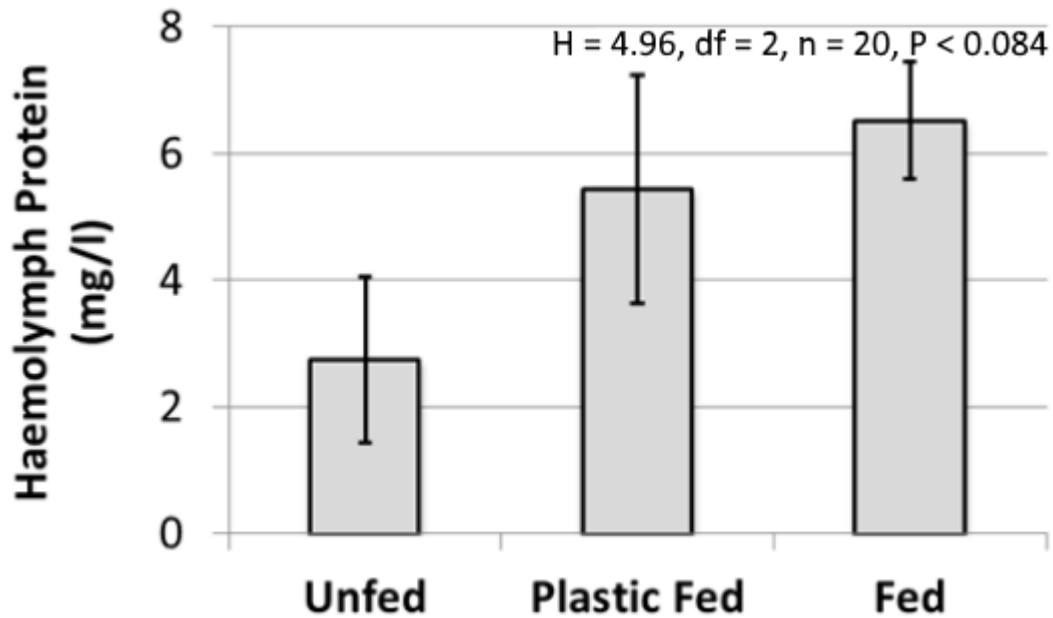
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276 *Metabolic Depression*

277 Analysis of the measures of metabolic depression indicated differences between the plastic-fed,
278 fed, and starved conditions. The level of protein in the haemolymph showed apparent variation
279 between the three groups (Fig. 4); however, this was not found to be significant when analysed
280 statistically (H = 4.96, d.f. = 2, P < 0.084). The minimal variation observed was predominantly
281 driven by the fed and starved control groups as indicated by post hoc Mann-Whitney analysis (W

282 = 21.0, $P < 0.05$), comparisons between the plastic-fed condition and the two control groups did
283 not reveal significant variation.

284



285

286 Fig. 4 Mean level of haemolymph protein of animals from the three experimental groups observed at month
287 eight (bars indicate standard error)

288

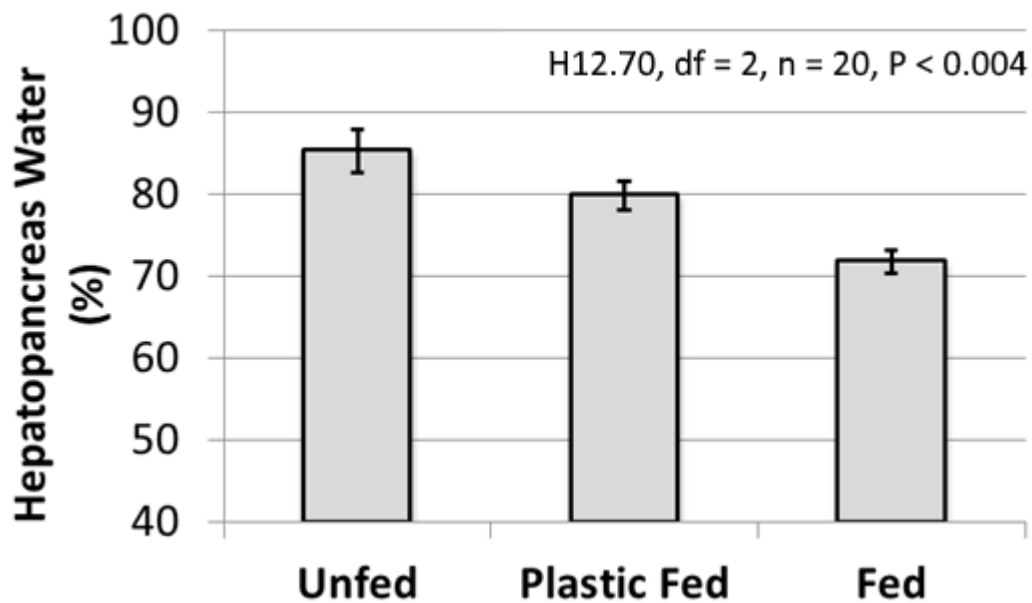
289 The breakdown of the main haemolymph protein, haemocyanin, releases two copper atoms
290 which build-up in the hepatopancreas. Examination of the level of copper in the hepatopancreas
291 revealed significant variation between the groups ($H = 7.96$, d.f. = 2, $P < 0.019$), although this was
292 driven by extraordinarily high levels of copper in two plastic-containing individuals. Mann-
293 Whitney analysis revealed differences between plastic-fed individuals and both controls;
294 however, these were only to 90 to 95% confidence (plastic-fed/starved: $W = 42.0$, $P < 0.0128$;
295 plastic-fed/fed: $W = 20.0$, $P < 0.0513$).

296 While the reduction in haemolymph protein in the plastic-fed individuals is not as marked as that
297 in the starved condition, it is clear that there is reduced nutrient uptake in *N. norvegicus*
298 contaminated with plastic, causing metabolic depression. A previous evaluation of starvation in
299 *N. norvegicus* carried out by Watts et al. (2014a) revealed that copper levels above $350.19 \mu\text{g g}^{-1}$
300 were indicative of starvation. The mean copper level in each group was above this threshold;
301 however, those of the plastic-fed groups were markedly higher, indicating that *N. norvegicus* in
302 the treatment group experienced metabolic depression.

303 *Use of Stored Lipids*

304 Metabolic depression is only effective for limited periods, prolonged episodes of insufficient
305 nutrients force *N. norvegicus* to utilise energy stores; first glycogen, and then lipids. Prolonged
306 starvation has previously been seen to result in a reduction in lipid in both the hepatopancreas
307 and tail (Barden, 1994).

308 All three treatment groups showed significant variation in nutritional state after eight months,
309 with an obvious reduction in the body condition of individuals exposed to microplastic. The
310 average water content of the hepatopancreas varied between the three conditions; 85.3% in the
311 starved condition, 79.9% in the plastic-fed condition, and 71% the fed condition (Fig. 5). This
312 variation was seen to be significant ($H = 12.70$, $d.f. = 2$, $P < 0.002$). Post hoc Mann-Whitney tests
313 revealed significant differences of at least 95% confidence between the three groups. The
314 relatively high water content in the starved and plastic condition indicates that these animals
315 have experienced a reduction in stored lipids which were then replaced by water.



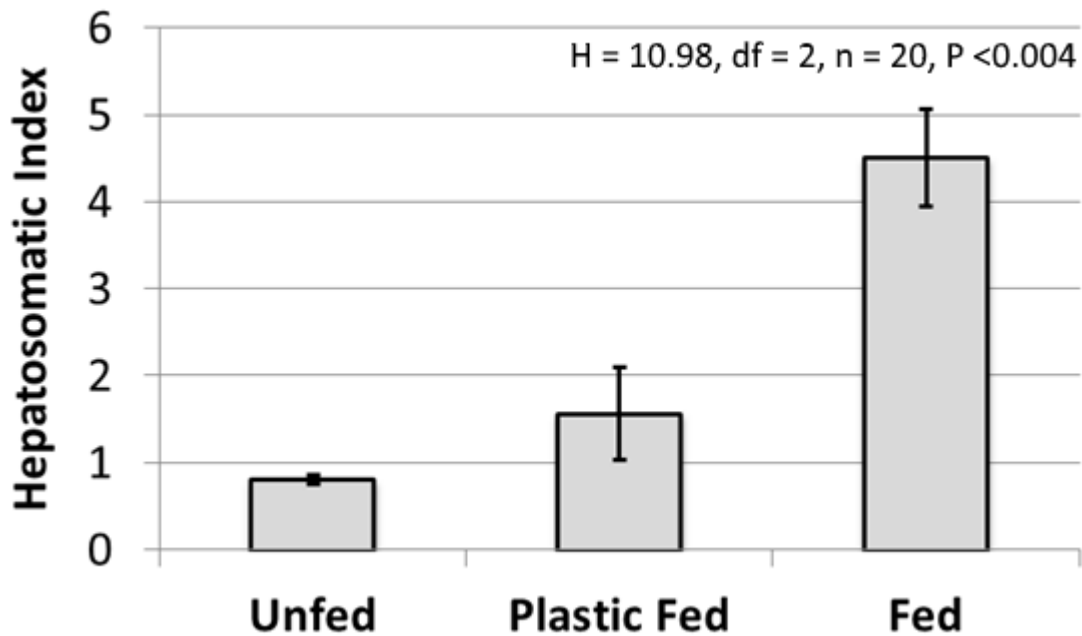
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317 Fig. 5. The mean level of water in the hepatopancreas (%) of animals from the three experimental groups
318 as observed at month eight (bars indicate standard error)

319

320 Correspondingly, the HSI of the plastic-fed condition was between that of the control groups. The
321 hepatosomatic index was seen to vary significantly between the three groups, ranging from 0.8%
322 in the starved condition to 4.5% in the fed condition ($H = 10.98$, $d.f. = 2$, $P < 0.004$) (Fig. 6). Post
323 hoc Mann-Whitney tests revealed differences between individuals in the fed and starved controls

324 (W = 76.0, P < 0.0043), and plastic-fed treatment and starved control (W= 86.0, P < 0.0128). The
325 lower HSI in the starved and plastic-fed conditions indicate that these animals have experienced
326 a reduction in available nutrients greater than that which could be mediated by metabolic
327 depression.



328

329 Fig. 6. The mean hepatosomatic index of animals from the three experimental groups observed at month
330 eight (bars indicate standard error)

331

332 *Further Impacts of Long Term Plastic Retention*

333 The results described above indicate that microplastic retention by *N. norvegicus* is linked to
334 reduced nutritional state; however, the prolonged exposure period experienced by *N. norvegicus*
335 may lead to secondary impacts of plastic ingestion beyond those of reduced feeding. The
336 intermoult period of *N. norvegicus* varies between the sexes; occurring every six months in males
337 and twelve months in ovigerous females (Farmer, 1973). As a result, females retain their
338 aggregations for longer periods and are at increased risk of developing large microplastic
339 aggregations as documented in Welden and Cowie (2016).

340 In addition to the impacts of low nutrient availability on metabolism and lipid stores discussed
341 above, starvation in crustaceans may cause reduced fecundity (Abelló and Sardá, 1982; Beyers
342 and Goosen, 1987; Hines, 1991; Lizárraga-Cubedo et al., 2003), limiting egg production and,
343 therefore, population growth. Such reduced fecundity has previously been observed in oysters

344 exposed to polystyrene microspheres (Sussarellu et al., 2014). This potential effect should be of
345 high concern in an economically important species such as *N. norvegicus*, landings of which are
346 regularly in the UK's top five highest grossing fisheries.

347 Retention of microplastic for long periods will also influence the uptake of additives and adsorbed
348 pollutants. Investigations of partitioning of hydrophobic contaminants have shown transfer
349 between gut plastic and animal tissues (Farrell and Nelson, 2013). In many of these species the
350 residence time of ingested plastics is believed to range from a number of hours (in marine
351 worms), to a number of days (in birds). The ability of *N. norvegicus* to retain plastic for a number
352 of months indicates an extended period over which hydrophobic contaminants may be
353 transferred to the organism and potentially contribute to deleterious population effects.

354

355 **Conclusions**

356 The results presented above indicate that microplastic aggregations reduce the nutritional health
357 of *N. norvegicus*. The reduction in mean body mass of the plastic-fed individuals indicates that
358 retention of microplastic results in lower growth rates. In wild populations experiencing
359 additional pressures, retention of microplastic may result in increased mortality and decreased
360 fecundity.

361

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