



Welden, N. and Cowie, P. R. (2016) Environment and gut morphology influence microplastic retention in langoustine, *Nephrops norvegicus*. *Environmental Pollution*, 214, pp. 859-865. (doi:[10.1016/j.envpol.2016.03.067](https://doi.org/10.1016/j.envpol.2016.03.067))

This is the author's final accepted version.

There may be differences between this version and the published version. You are advised to consult the publisher's version if you wish to cite from it.

<http://eprints.gla.ac.uk/121326/>

Deposited on: 20 September 2018

Enlighten – Research publications by members of the University of Glasgow
<http://eprints.gla.ac.uk>

1 Environment and Gut Morphology influence Microplastic Retention in
2 Langoustine, *Nephrops norvegicus*
3 Natalie A.C. Welden*[‡], Phillip R. Cowie^α

4 * Corresponding author: natalie.welden@york.ac.uk Present Address: Stockholm Environment Institute, University of
5 York, Heslington, York, YO10 5DD

6
7 ‡ College of Medical, Veterinary and Life Sciences, University of Glasgow, Glasgow, G12 89QQ, Scotland

8 * University Marine Biological Station, Marine Parade, Millport, KA28 0EF, Scotland

9 ^α FSC Millport, Marine Parade, Millport, North Ayrshire, KA28 0EF, Scotland

10

11 **ABSTRACT**

12 Over the past twenty years microplastic pollution has been recorded in all major marine habitats,
13 and is now considered to be of high environmental concern. Correspondingly, the number of
14 reports of microplastic ingestion by marine species is increasing. Despite this, there are still
15 relatively few studies which address the uptake and retention of microplastic in wild populations.
16 Langoustine, *Nephrops norvegicus*, sampled from the Clyde Sea Area, have previously been seen to
17 contain large aggregations of microplastic fibres. The large proportion of contaminated
18 individuals and size of the microplastic aggregations observed suggests that *Nephrops* are at high
19 risk of microplastic ingestion. In this study the levels of ingested microplastic in populations of *N.*
20 *norvegicus* from the Clyde Sea Area, North Minch and North Sea are examined. Animals in the
21 near-shore, Clyde Sea population showed both a higher percentage of microplastic containing
22 individuals and much greater weights of microplastic retained in the gut. *N. norvegicus* revealed
23 that only a small percentage of individuals from the North Sea and Minch contained microplastic,
24 predominantly single strands. An expanded sample from the Clyde Sea Area was examined to
25 identify the factors influencing microplastic retention. This revealed that males, larger individuals,
26 and animals that had recently moulted contained lower levels of microplastic. The presence of
27 identified food items in the gut was not seen to correlate with microplastic loads. Observations of
28 microplastic in the shed stomach lining of recently moulted individuals and the lack of
29 aggregations in wild-caught individuals suggests that ecdysis is the primary route of microplastic
30 loss by *N. norvegicus*. Therefore the large aggregations observed in wild-caught animals are
31 believed to build up over extended periods as a result of the complex gut structure of *N.*
32 *norvegicus*.

33

34 **Keywords:** microplastic; pollution; monitoring; Decapoda;

35

36 **Capsule:** Analysis of microplastic aggregation by wild *Nephrops norvegicus* from three populations determined that
37 location and moult stage have the largest effect on aggregation.

38

39 1. Introduction

40 The current scientific focus on the distribution and fate of microplastic pollution has led to
41 numerous studies of its effects on marine communities. Due to the resistance of polymers to
42 degradation and their relative buoyancy, plastics are able to persist for long periods in the marine
43 environment, and be carried far from their source (Barnes et al., 2009). As a result, microplastics
44 have been reported in environments far from anthropogenic activities. (Barnes et al., 2009; Van
45 Cauwenberghe et al., 2013).

46 Many of these observations of marine microplastic have shown a high degree of heterogeneity in
47 microplastic distribution (Ryan et al., 2009). The greatest densities of microplastic debris have
48 been reported from the centres of gyres (Moore et al., 2001), lagoons (Vianello et al., 2013), and in
49 coastal sediments (Claessens et al., 2011). These apparent at risk areas are the result of a number
50 of environmental factors, such as wind direction, currents, and bathymetry (Dixon and Dixon,
51 1983; Moreira et al., 2016; Shaw and Mapes, 1979). Proximity to sources of microplastic pollution
52 have also been seen to have a significant impact on local abundance (Reddy et al., 2006).

53 Despite the recent increase in available literature, much of the evidence on plastic uptake by
54 animals relates to the ingestion of macroplastic by large marine vertebrates, such as birds (van
55 Franeker et al., 2004) and turtles (Lutz, 1990). Sampling of fish collected in trawls has also shown
56 that a range of fishes also take up both macro- and microplastics from the environment (Boerger
57 et al., 2010; Lusher et al., 2013). Unfortunately due to the large ranges over which these species
58 forage, and the uncertainty over the length of time plastic is retained in the gut, it is difficult to
59 draw a conclusion as to the relationship between environmental and ingested plastics.

60 Many of the observations of plastic ingestion by invertebrates stem from laboratory
61 investigations. In this way, uptake of microplastics has been observed in blue mussel, *Mytilus*
62 *edulis* (Browne et al., 2008; Farrell and Nelson, 2013; von Moos et al., 2012), shore crab, *Carcinus*
63 *maenas* (Farrell and Nelson, 2013), sandhoppers, *Talitrus saltator* (Ugolini et al., 2013), lugworm,
64 *Arenicola marina* (Browne et al., 2013), and echinoderms (Graham and Thompson, 2009). Shore
65 crabs have also been seen to take in plastic microspheres through the gills during normal
66 respiration (Watts et al., 2014), and trophic links have been indicated by the transfer of
67 microbeads from mussels (Farrell and Nelson, 2013).

68 Fewer studies examine the uptake of microplastic by wild-caught invertebrates; however, the
69 level of uptake observed appears to support the findings of laboratory investigations. Large
70 numbers of contaminated individuals have been recorded amongst crustaceans; for example 63%
71 of brown shrimp, *Crangon crangon*, sampled from the English Channel and southern North Sea
72 were seen to contain microplastics (Devriese et al., 2015), as were 82% of langoustine, *Nephrops*
73 *norvegicus*, from the Clyde Sea Area (Murray and Cowie, 2011). Lower levels were observed

74 amongst gooseneck barnacles, *Lepas* spp., with 33.5% of 385 individuals sampled from the North
75 Pacific Sub-tropical Gyre seen to contain microplastic (Goldstein and Goodwin, 2013).

76 *N. norvegicus* is a species of great importance to the UK fishing industry. In 2014 it accounted for a
77 fifth of the weight of shellfish landings by the UK fleet and a third of the value, at 30 thousand
78 tonnes and £99 million. This substantial sum made it the second most important fishery in the UK
79 in 2014. Murray and Cowie's examination of 120 *N. norvegicus* from the Clyde Sea indicated that
80 large aggregations of microplastic are found in a significant proportion of the population. Unlike
81 the vertebrates previously seen to take up plastics in the wild, *N. norvegicus* feed within a small
82 area around their burrows; thus the level of contamination in wild caught animals is potentially
83 indicative of the amount of microplastic in the surrounding environment.

84 The Clyde Sea Area is an enclosed waterbody in close proximity to numerous potential
85 microplastic sources, a combination of factors which suggests a high abundance of environmental
86 microplastics. However, geographically separated populations of *N. norvegicus* are exposed to
87 very different bathymetric conditions and anthropogenic influences. The variation in distance
88 from sources of litter suggests that the average intake of microplastic by *N. norvegicus* populations
89 in other locations may be much lower.

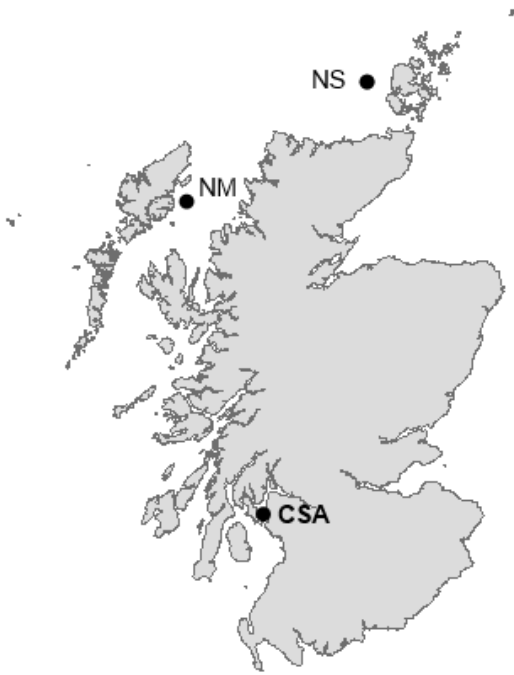
90 This study examines the occurrence of microplastic in *N. norvegicus* in Scottish waters. The work
91 aims to determine whether the high levels of microplastic observed by Murray and Cowie (2011)
92 are representative of those in other populations. Analysis of the environmental and biological
93 factors related to microplastic levels in the three studied populations is used to identify the
94 factors responsible for the aggregation of ingested microplastic.

95

96 **2. Materials and Methods**

97 *2.1. Microplastic in Scottish Nephrops norvegicus*

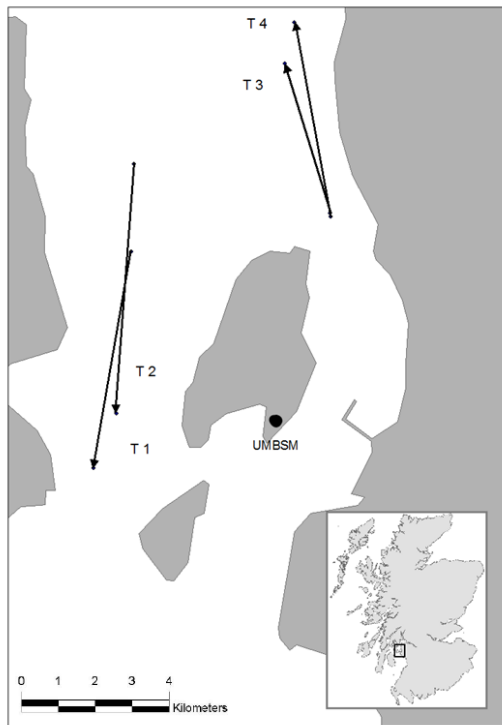
98 *N. norvegicus* were collected from three sites in North and West Scotland; the Clyde Sea Area
99 (CSA), the North Minch (NM), and the North Sea (NS) (Fig. 1). In the CSA four trawls were taken at
100 Skelmorlie Bank and in the Main Channel at depths between 58 and 110 metres in May, June and
101 August (Fig. 2). Sampling was carried out using otter trawls rigged with 50mm mesh. To reduce
102 the potential for uptake of fibres from the sampling net was reduced by only carrying out short
103 trawls. Individuals were frozen immediately on landing to prevent digestion of the gut content.



104

105 **Fig. 1.** Trawl Locations in the North Sea (NS) -3°49.07'E, 59°03.39'N, North Minch (NM) -6°09.13'E,
 106 58°08.57'N, and Clyde Sea Area (CSA) -4.9751E, 55.7892N

107



108

109 **Fig. 2.** Showing trawl locations in the Clyde Sea Area in relation to the University Marine Biological Station
 110 Millport (UMBSM). T1: 16/06/2011 -4.8903E, 55.7998N ~ -4.9093E, 55.8463N, T2: 16/06/2011 -4.9751E,
 111 55.7892N ~ -4.9872E, 55.7368N, T3: 08/07/2011 -4.8905E, 55.8005N ~ -4.9127E, 55.8362N, T4:
 112 11/08/2011 -4.9755E, 55.8105N ~ -4.9131E, 55.8472N

113

114 Animals were defrosted prior to dissection, and their sex, moult stage, and carapace length
115 recorded. Moult stage was determined by testing the hardness of the thorax, directly behind the
116 eyes. Intermoult individuals could be identified by their hard carapaces, while recently moulted
117 individuals have a jelly-like carapace, and those of animals immediately prior to and post moult is
118 papery (Farmer, 1973). The carapace of the individual was then removed and the muscle of the
119 thorax and tail separated to allow the removal of the stomach and gastric tract, which was
120 preserved in 80% ethanol (Murray and Cowie, 2011).

121 The content of each gut was examined under a stereo microscope to determine the volume and
122 identity of natural prey and presence of potential microplastics. Gut contents were examined in
123 subsets of approximately 0.5 ml (a level spatula) to ensure that the detectability of plastic in the
124 gut contents was not impacted by the volume of food. Plastic materials were categorised as either
125 pre-production pellets, fragments, films or fibres. Aggregations of fibres were grouped into the
126 following subcategories; up to five strands, strands and a loose ball of fibres, and a tight ball of
127 multiple fibres (Murray and Cowie, 2011). A Mettler MX5 balance (Mettler-Toledo international
128 Inc., Columbus, USA) was then used to record the weight of plastic recovered from each individual
129 to five decimal places. Prior to weighing, any algae tangled among the plastic fibres were removed
130 and the samples air dried for 48 hours. Each sample was weighed three times and a mean taken.

131

132 *2.2 Identification of Microplastic*

133 FT-IR spectrometry was used to identify a sub-set of 100 suspected plastic items. Tangled fibres
134 were separated for individual analysis and all samples were rinsed in distilled water and allowed
135 to air dry to ensure the cleanest possible spectrum. Samples were analysed using a Shimadzu
136 8400s spectrometer and the resulting spectra were compared to those of a range of known
137 polymer standards to confirm their identity. The percentage of successfully identified plastics was
138 used to calculate the actual number of microplastic items recovered.

139

140 *2.3. Duration of Microplastic Retention*

141 Previous examination of 120 individuals from the CSA indicated lower plastic contamination in
142 recently moulted individuals (Murray and Cowie, 2011). To establish whether microplastics are
143 lost during ecdysis a two month feeding trial was carried out. Ten recently moulted female *N.*
144 *norvegicus* were placed into individual tanks and fed with a daily ration of 0.5 g of squid mantle,
145 seeded with five 0.5mm strands of polypropylene (PP) rope. After the first month, bilateral eye
146 ablation was used to induce moult (Fingerman, 1987). Feeding with microplastic seeded squid

147 was continued until the individual had achieved ecdysis. The shed gut lining was recovered and
148 the moulted individual frozen for gut content analysis as described above.

149

150 *2.4. Statistical Analysis*

151 Analysis of the factors affecting microplastic accumulation by *N. norvegicus* was carried out using
152 Minitab 15. The relative frequencies of microplastic containing animals in the three populations
153 were examined using Chi-squared analysis.

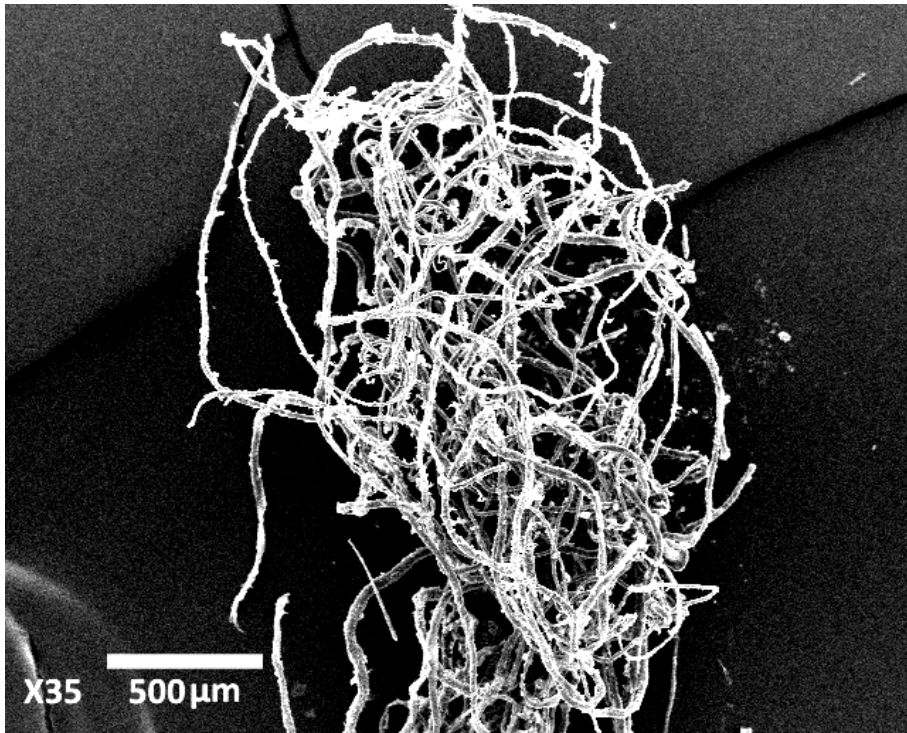
154 The data from the CSA sample was used to determine both the factors affecting microplastic
155 ingestion, and those affecting the weight of retained microplastic. The statistical software R,
156 version 3.0.2, was used to relate the microplastic data to carapace length, sex, moult stage, trawl
157 number, sampling site, and the presence and type of food. Factors associated with the likelihood
158 of plastic occurrence in *N. norvegicus* were determined by fitting a binary logistic model (BLM).
159 The factors responsible for the weight of retained plastic were examined using a generalised
160 linear model (GLM).

161

162 **3. Results**

163 Trawl samples returned 1450 animals for dissection and analysis. Those from the North Minch
164 (150 animals) and North Sea (300 animals) were all male, whilst the larger sample of individuals
165 from the CSA (1000 animals) was separated into 50% males and females. Of the total 1450
166 individuals, 975 (67%) were seen to contain microplastic, predominantly microfibres.

167 The fibres recovered were a range of colours and thicknesses, the exact proportions of which
168 could not be determined due to the highly tangled aggregations. Of the samples yielding
169 sufficiently clear spectra for analysis 94% were confirmed as plastics. Nylon and polypropylene
170 were the most frequently observed polymers, and made up 37.2%, 29.8% and 12.8% of the
171 analysed plastic, respectively. Smaller amounts of polyethylene - mainly from ingested films - and
172 PVC were also recovered.



173

174 **Fig. 3.** Aggregation of plastic fibres recovered from the foregut of a female *Nephrops norvegicus* from the
 175 Clyde Sea Area

176

177 *3.1. Local Variation in Microplastic Uptake*

178 Variation was seen in the proportion of individuals at each site which contained microplastics
 179 (Table 1). Chi-squared analysis of the number of contaminated individuals at each site indicated a
 180 significant difference between the three locations ($P < 0.001$, $X^2 = 572.756$, $df = 10$). This disparity
 181 is driven by individuals sampled from the Clyde.

182 **Table 1**

183 The occurrence and retention of microplastics by Scottish *Nephrops norvegicus* stocks

Site	Sample Size	Proportion of Sample seen to Contain Microplastics	Maximum Microplastic Weight (mg)	Average Microplastic Weight (mg)
Clyde Sea Area	1000	84.10%	0.09	-
North Minch	150	43.00%	0.01	>0.01
North Sea	300	28.70%	0.80	0.40

184

185

186 The most commonly isolated plastics were fragmented fibres. Other plastics found were mainly
 187 films, although one pre-production nib was isolated. In the offshore populations, contaminated
 188 individuals contained a maximum of 5 fibres, whilst 41% of the CSA were seen to have aggregated
 189 tangled “Balls” of multiple fibres and algae (Fig. 3).

190

191 3.2. Factors Affecting Microplastic in the Gut of *N. norvegicus* from the Clyde Sea

192 Trawl depths in the CSA ranged from 74 – 115m in the Main Channel and 60 – 75m in the Fairlie
 193 Channel; sediment analysis at the two sites revealed average grain sizes of 0.18mm and 0.166mm
 194 respectively. The carapace length of individuals in the CSA sample ranged from 19.8 to 59.1mm,
 195 and was found to be normally distributed when examined using Kolmogorov-Smirnov analysis ($P <$
 196 0.010). The proportion of individuals at each moult phase differed between males and females,
 197 possibly the result of reduced moult frequency in mature females (Farmer, 1973). The
 198 examination of identifiable prey items indicated a diet dominated by bivalve molluscs and
 199 crustaceans, with *N. norvegicus* carapace regularly observed. These two categories made up
 200 74.1% of the identifiable gut contents, with the rest being comprised of fish bones, echinoderms
 201 and polychaetes. Variation in the aggregation and weight of plastic was observed in individuals
 202 recovered from the four trawls, with lower weights of plastic seen later in the year (Table 2).

203

204 **Table 2**

205 The occurrence and retention of microplastics by *Nephrops norvegicus* in the Clyde Sea Area

Trawl	Site	Date	Total Animals	Upto five fibres	Between 5 and a loose ball of fibres	Ball of fibres	Individuals seen to contain films	Average of Plastic Weight (mg)
1	Main Channel	16/06/2011	383	19.32%	21.40%	51.69%	3.39%	0.66
2	Fairlie Channel	16/06/2011	184	32.06%	22.28%	36.95%	3.80%	0.47
3	Main Channel	08/07/2011	275	40.36%	19.27%	18.18%	3.27%	0.20
4	Fairlie Channel	11/08/2011	158	33.54%	6.96%	19.62%	12.65%	0.28

206

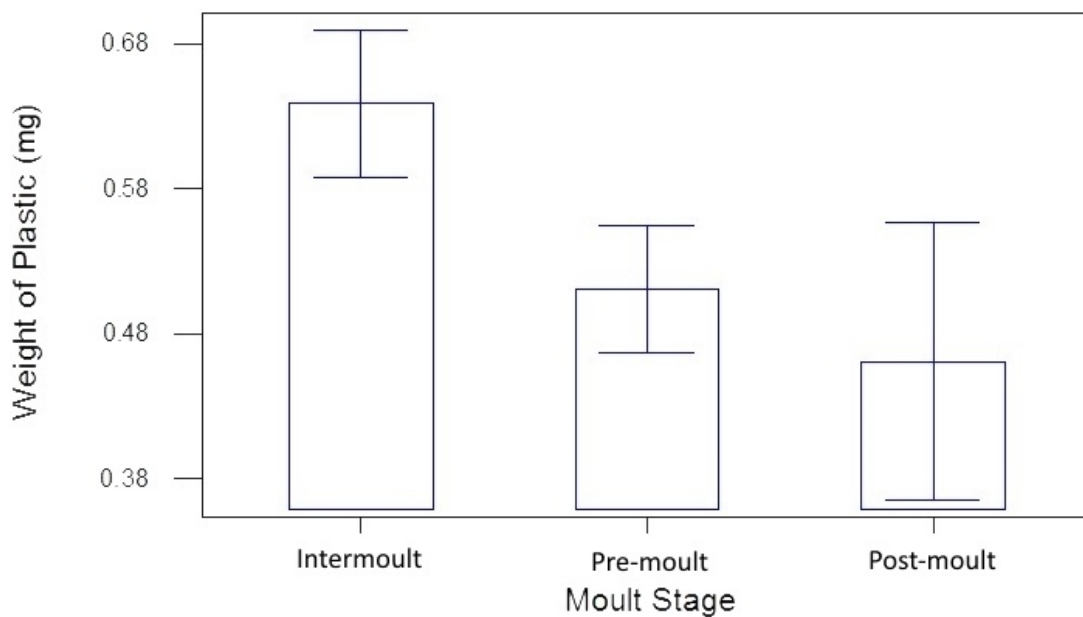
207 The results of the BLM identified moult stage, date of trawl, and carapace length as having a
 208 significant impact on the likelihood of plastic contamination in *N. norvegicus*. Recently moulted
 209 (“jelly” carapace) individuals were seen to be less likely to contain plastics than those at
 210 intermoult (“hard” carapace) ($z = -6.112$, $P < 0.001$). Carapace length was also seen to effect the
 211 likelihood of plastic presence, with smaller individuals more likely to contain microplastics ($z = -$
 212 1.829 , $P < 0.05$); however, the observed relationship was of lower significance than that of moult
 213 stage and trawl date.

214

215 While there was a significant difference in the occurrence and aggregation of plastic recovered
216 from *N. norvegicus* from different geographical areas, there was no difference observed between
217 the trawl locations within the CSA. The only non-biotic factor observed to have a significant
218 impact on whether plastic was present within the gut was the season in which the animals were
219 collected; with lower likelihood of plastic contamination in tows carried out in June; trawl three
220 ($z = -3.675$, $P < 0.001$), and August; trawl 4 ($z = 4.3$, $P < 0.001$). This variation is believed to be due to
221 a reduction in the number of recently moulted individuals later in the year.

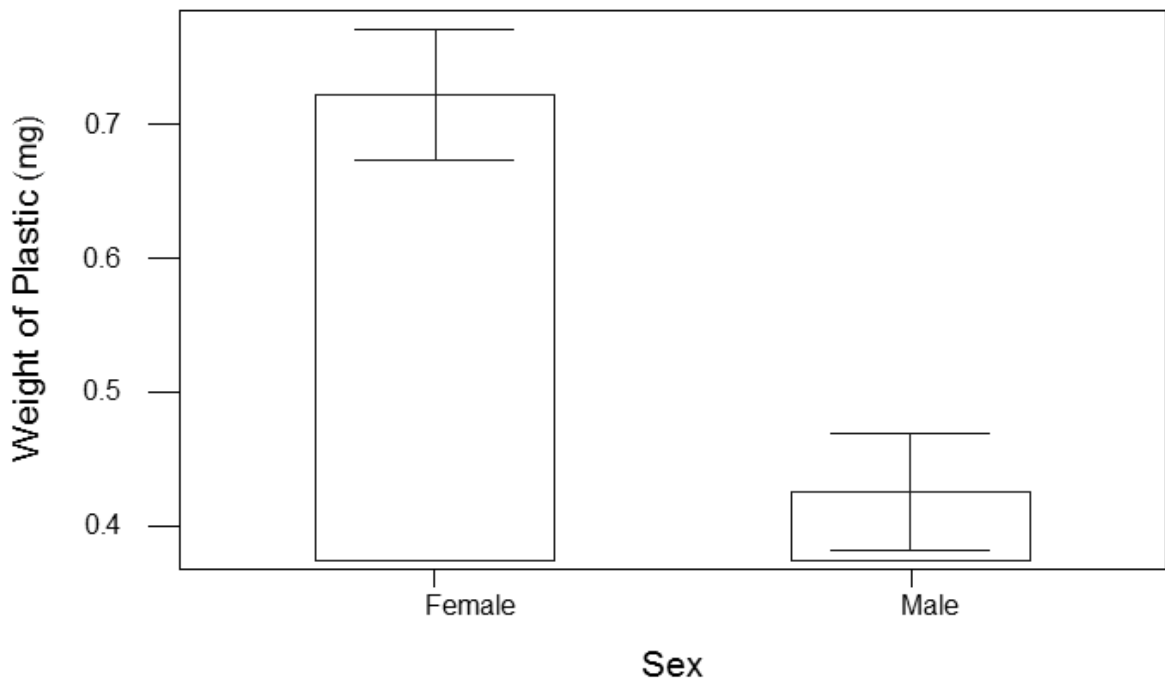
222 The results of the GLM analysis of the factors associated with variation in the weight of
223 microplastic returned a similar response to that of the BLM of plastic occurrence. The results
224 indicated that the moult stage of the individual was significantly related to the weight of plastic
225 present ($P < 0.001$), this was driven by lower weights of plastic in recently moulted individuals
226 (Fig. 4). Females were seen to retain greater weights of plastic than males ($t = 4.245$, $P < 0.001$)
227 (Fig.5). A significant negative relationship was also observed between the proportion of gut
228 occupied by food and the weight of recovered plastic ($P < 0.001$); individuals recorded as having no
229 food in the foregut were observed to have the highest microplastic load. Sampling trawl had the
230 highest influence over the amount of plastic retained ($P < 0.001$), this was driven by a low average
231 plastic weight recovered in animals from trawls three and four.

232



233

234 **Fig. 4.** The weight of plastic (mg) recorded in *Nephrops norvegicus* at each moult stage



235

236 **Fig. 5.** The weight of plastic (mg) recorded in male and female *Nephrops norvegicus*

237

238 3.3. Plastic Lost at Moult

239 Seven of the 10 animals subjected to eye ablation moulted within the two month experimental
 240 period. During the feeding period the animals did not consume the whole food ration each day,
 241 and so the total number of fibres ingested could not be definitively stated; however, when the gut
 242 linings were examined under stereo microscope, five were seen to contain microplastics. Stomach
 243 content analysis carried out on all post moult individuals revealed no remaining plastics in the
 244 foregut, whereas plastic aggregations were observed in the three un-moulted individuals.

245

246 4. Discussion

247 Despite the number of studies into the distribution of microplastics, there are few that look at
 248 their ingestion in benthic habitats (Reddy et al., 2006; Van Cauwenberghe et al., 2013). Our results
 249 demonstrate microplastic uptake by *N. norvegicus* from each of the sampled locations. The most
 250 commonly observed plastics in all three populations were fibres, indicating either that these are
 251 the most abundant plastics at all sites or that fibres are more easily ingested. In previous studies
 252 of microplastic in the marine environment, fibres have been the dominant plastic category .

253 These fibres varied in colour, thickness and degree of wear, and are believed to originate from a
 254 range of sources. FT-IR analysis of single micro-fibres proved highly laborious, occasionally

255 resulting in unclear, 'noisy' results – an issue also stated by Gallagher et al. (2015), however this
256 did reveal 96% success rate in visual identification of plastics. This revealed a mix of polymers.
257 Many of the observed fibres may have entered the CSA from the River Clyde and are potentially
258 the released from clothes washing as outlined in Browne et al. (2011). Trawl nets may release
259 plastics into the CSA both from regular use and the breakdown of lost gear; however, the number
260 of blue and orange fibres observed was comparatively low.

261

262 4.1. Local Variation in Microplastic Uptake

263 The frequency of microplastic occurrence and level of aggregation observed in the CSA support
264 that previously recorded by Murray and Cowie (2011) from a smaller sample size; however, it is
265 apparent from the results that *N. norvegicus* from the North Minch and North Sea have
266 substantially lower microplastic loads. The disparity in microplastic uptake is believed to be
267 caused by the CSA's relative proximity to microplastic sources, resulting in locally raised
268 concentrations of environmental microplastics. As such, areas close to high levels of human
269 activity - such as estuaries and enclosed water bodies - may be thought of as high risk, and
270 animals living there as having a greater likelihood of microplastic uptake. Within the Clyde sea
271 there were significant differences in microplastic load between sample trawls; however, previous
272 examination of the average abundance of microplastic in the sediments of the North Channel and
273 Fairlie Channel – 45.5 and 42.2 plastic items per kilogram respectively – showed no significant
274 variation (Welden, unpublished data). It is believed that the variation observed can be attributed
275 to biotic differences.

276 The current route by which microplastics enter the food chain is unclear; however, many species
277 of invertebrates (Devriese et al., 2015; Ugolini et al., 2013; Van Cauwenberghe and Janssen, 2014)
278 and fishes (Boerger et al., 2010; Lusher et al., 2013; Lusher et al., 2015) have shown some degree
279 of contamination. In the marine environment, interactions between animals and microplastic may
280 occur in a number of ways; when examining plastic contaminated fish Lusher et al (2013) suggest
281 that plastic is taken up accidentally during feeding, whereas Boerger et al (2010) indicate that
282 microplastics may be actively consumed due to their resemblance to planktonic prey. Planktonic
283 crustaceans have also been seen to actively ingest plastics, although some species appeared able
284 to discriminate against larger polystyrene beads (Bern, 1990).

285 *N. norvegicus* act as both scavengers and carnivores, and may take up plastic during feeding or
286 burrowing activities. Other crustaceans, such as crabs, have comparable feeding methods, and
287 may be at a similar risk of microplastic loading. The shore crab *C. maenas* has been seen to take in
288 microplastic spheres from contaminated mussel, *M. edulis* (Farrell and Nelson, 2013; Watts et al.,

289 2014), and through the gills during respiration (Watts et al., 2014), as well as fibres from
290 prepared mussel/gelatine blocks (Watts et al., 2015).

291 In a study of the impact of feeding mode, filterers such as bivalves were found to ingest the
292 highest levels of microspheres (Setälä et al., 2015); if this is also true of fibres, molluscs in the CSA
293 would experience a higher frequency of microplastic ingestion than that observed here. However,
294 this relies on microplastic being obtained directly from the environment and does not take into
295 account potential bioaccumulation. *N. norvegicus* are opportunistic scavengers, consuming a
296 range of prey species. Bioaccumulation would rely on animals consuming multiple small prey
297 animals whole, consuming their plastic load in the process. The observations of gut contents
298 reported here revealed a high percentage of larger animals, such as mollusc and crustaceans,
299 which were partially consumed. The potential for bioaccumulation is currently obscured by
300 numerous sources of uncertainty; further research into the trophic transfer of microplastic and its
301 retention by a range of species is required before the true risk can be established.

302

303 4.2. Biological Factors Influencing Microplastic Retention

304 The volume of microplastic recovered from *N. norvegicus* in the CSA reveals that aggregations are
305 held in the foregut for extended periods of time. The statistical analysis of the factors responsible
306 for high microplastic weights indicated that sex, size, and moult stage have the greatest influence
307 on aggregation.

308 The negative relationship between body size and microplastic loads may be the result of the gut
309 morphology of *N. norvegicus*. The digestive tract of crustaceans is relatively complex that
310 compared to other invertebrates. The gastric mill is a set of chitinous plates found in the foregut,
311 at the entrance to the hindgut (Farmer, 1975). The shape of these plates and the narrowing at the
312 entrance to the hindgut may prevent microplastic from being egested with natural food-stuff.
313 Previous work examining the morphology of the gastric mill in relation to carapace length has
314 shown that the gaps in the mill increase with growth (Welden et al., 2015). The increased size of
315 the gaps between plates of the mill may allow a greater amount of microplastic to be lost by
316 egestion in larger individuals.

317 This link between size and microplastic loss may also explain the relationship seen between sex
318 and microplastic aggregation. Female *N. norvegicus* grow at a slower rate than males due to a
319 decreased moult frequency. As a result they have smaller gastric mills which would prevent the
320 egestion of microplastics that could be passed by larger, male conspecifics.

321 The presence of fibres in the discarded gut lining of moulted *N. norvegicus* indicates that
322 microplastic can be lost at ecdysis. This supports the outputs of both the BLM of microplastic

323 occurrence and the GLM of factors influencing the weight of microplastic retained in the gut.
324 Consequently, the authors believe moulting to be the lead cause of microplastic loss in
325 langoustine.

326 Of the biological factors linked to the retention of microplastic it is likely that the size of the
327 gastric mill has the greatest influence on the retention of microplastics; particularly in larger size
328 classes. The significantly lower weight of microplastic in recently moulted individuals from the
329 Clyde indicate that *N. norvegicus* rid themselves of large plastic aggregations at moult, this was
330 supported by the observed plastic in the shed guts of animals subjected to eye ablation. These two
331 factors are believed to have the greatest effect on the weight of retained microplastic. The
332 discrepancy in microplastic observed between male and female langoustine is believed to be the
333 result of increased moult frequency in male langoustine and, as male langoustine are generally
334 larger, increased size in the gastric mill.

335

336 4.3. Potential Impacts of Microplastic Retention

337 The large aggregations of microplastic fibres observed in the CSA population indicate that *N.*
338 *norvegicus* in this area are at increased risk of the biological impacts of plastic ingestion. Previous
339 studies have shown a number of effects of ingested plastic on an animal's fitness. These include
340 false satiation, previously described in seabirds (Ryan, 1988) and turtles (Lutz, 1990; McCauley
341 and Bjorndal, 1999), and nutrient dilution - preventing the assimilation of ingested foods
342 (McCauley and Bjorndal, 1999).

343 Although different in their mechanics, both of these conditions cause a reduced nutritional state
344 and have been seen to result in starvation. For example, in the lugworm, *Arenicola marina*, plastic
345 ingestion negatively affected feeding rate, leading to reduced body mass (Besseling et al., 2012). In
346 addition to their impact on the body condition, plastics also carry hydrophobic contaminants
347 (Teuten et al., 2007; Teuten et al., 2009). Regular ingestion and retention may result in pollutants
348 transferring from plastic to the organism (Besseling et al., 2012).

349 The results presented above describe a negative relationship between microplastic weight and
350 stomach fullness. This may indicate reduced feeding as a result of false satiation. Using the relative
351 proportions of the identified polymers we calculated a mean specific gravity for the plastics in *N.*
352 *norvegicus* in the CSA. From this it was possible to calculate an approximate mean volume of
353 0.68mm³ of aggregated plastic per contaminated individual. The calculated volume of the largest
354 recorded aggregation was 9.40mm³. This volume may appear low on first examination; however
355 the size of the observed aggregations were increased by trapped natural materials. An individual
356 of 20mm carapace length is expected to have a gut volume of 0.806cm³ (Welden et al., 2015), and

357 the combined plastic and algae aggregations observed took up to ten percent of the foregut.
358 Whilst *N. norvegicus* are highly tolerant to starvation (Mente, 2010), long periods of retention may
359 cause reduced (or even negative) growth.

360 Due to their smaller size and reduced moult rate at maturity, female *N. norvegicus* would retain
361 plastic for up to twice as long as males, making them more likely to contain high plastic loads.
362 Reduced body mass, is known to lower fecundity in a number of crustacean species (Beyers and
363 Goosen, 1987; Hines, 1991; Lizárraga-Cubedo et al., 2003), including *N. norvegicus* (Abellô and
364 Sardá, 1982). In European Lobster, *Homarus gammarus*, smaller individuals have also been shown
365 to have smaller eggs (Tully et al., 2001). In this way, sub-lethal microplastic loads may have
366 impacts at the population level.

367 *N. norvegicus* is a species of high economic importance in Europe. The impacts of microplastic
368 ingestion on fitness and fecundity may impact the viability of nearshore fisheries. As a result of
369 this uncertainty, the authors consider further examination of the impact of microplastic on the
370 fitness of *N. norvegicus* to be of high importance.

371

372 **5. Conclusion**

373 It is clear from the results presented that *N. norvegicus* from nearshore habitats exhibit
374 significantly higher microplastic abundance in them than those located in areas further from
375 anthropogenic inputs. In addition to the much lower percentage of individuals seen to contain
376 plastic, the large aggregations recorded in Clyde Sea animals were not observed in those from the
377 North Sea and North Minch.

378 As well as the effect of location on plastic uptake, the individuals in the CSA sample indicate that
379 size, sex and moult stage significantly influence microplastic loads. The ability of *N. norvegicus* to
380 routinely expel microplastic aggregations along with the gut lining at moult reduces the negative
381 effect of their complex gut morphology to some extent; however, the possibility of a 12 month
382 microplastic exposure period suggests a high probability of associated negative impacts.

383

384 **References**

- 385 Abelló, P., Sardá, F. (1982) The Fecundity of the Norway Lobster *Nephrops Norvegicus* (L.) Off the
386 Catalan and Portuguese Coasts. *Crustaceana* 43, 13-20.
- 387 Barnes, D.K.A., Galgani, F., Thompson, R.C., Barlaz, M. (2009) Accumulation and fragmentation of
388 plastic debris in global environments. *Philosophical Transactions of the Royal Society B: Biological*
389 *Sciences* 364, 1985-1998.
- 390 Bern, L. (1990) Size-related discrimination of nutritive and inert particles by freshwater
391 zooplankton. *Journal of Plankton Research* 12, 1059-1067.
- 392 Besseling, E., Wegner, A., Foekema, E., Van Den Heuvel-Greve, M., Koelmans, A.A. (2012) Effects of
393 microplastic on fitness and PCB bioaccumulation by the lugworm *Arenicola marina* (L.).
394 *Environmental science & technology*.
- 395 Beyers, C.J.D.B., Goosen, P.C. (1987) Variations in fecundity and size at sexual maturity of female
396 rock lobster *Jasus lalandii* in the Benguela ecosystem. *South African Journal of Marine Science* 5,
397 513-521.
- 398 Boerger, C.M., Lattin, G.L., Moore, S.L., Moore, C.J. (2010) Plastic ingestion by planktivorous fishes
399 in the North Pacific Central Gyre. *Marine pollution bulletin* 60, 2275-2278.
- 400 Browne, M.A., Crump, P., Niven, S.J., Teuten, E., Tonkin, A., Galloway, T., Thompson, R. (2011)
401 Accumulation of Microplastic on Shorelines Worldwide: Sources and Sinks. *Environmental science*
402 *& technology* 45, 9175-9179.
- 403 Browne, M.A., Dissanayake, A., Galloway, T.S., Lowe, D.M., Thompson, R.C. (2008) Ingested
404 Microscopic Plastic Translocates to the Circulatory System of the Mussel, *Mytilus edulis* (L.).
405 *Environmental science & technology* 42, 5026-5031.
- 406 Browne, M.A., Niven, S.J., Galloway, T.S., Rowland, S.J., Thompson, R.C. (2013) Microplastic moves
407 pollutants and additives to worms, reducing functions linked to health and biodiversity. *Curr Biol*
408 23, 2388-2392.
- 409 Claessens, M., Meester, S.D., Landuyt, L.V., Clerck, K.D., Janssen, C.R. (2011) Occurrence and
410 distribution of microplastics in marine sediments along the Belgian coast. *Marine pollution*
411 *bulletin* 62, 2199-2204.
- 412 Devriese, L.I., van der Meulen, M.D., Maes, T., Bekaert, K., Paul-Pont, I., Frère, L., Robbens, J.,
413 Vethaak, A.D. (2015) Microplastic contamination in brown shrimp (*Crangon crangon*, Linnaeus
414 1758) from coastal waters of the Southern North Sea and Channel area. *Marine pollution bulletin*
415 98, 179-187.
- 416 Dixon, T.J., Dixon, T.R. (1983) Marine litter distribution and composition in the North Sea. *Marine*
417 *pollution bulletin* 14, 145-148.
- 418 Farmer, A. (1975) Synopsis of biological data on the Norway lobster *Nephrops norvegicus*
419 (Linnaeus, 1758). *FAO Fisheries Synopses (FAO)*. no. 112.
- 420 Farmer, A.S. (1973) Age and growth in *Nephrops norvegicus* (Decapoda: Nephropidae). *Marine*
421 *Biology* 23, 315-325.
- 422 Farrell, P., Nelson, K. (2013) Trophic level transfer of microplastic: *Mytilus edulis* (L.) to *Carcinus*
423 *maenas* (L.). *Environmental Pollution* 177, 1-3.

- 424 Fingerman, M. (1987) The Endocrine Mechanisms of Crustaceans. *Journal of Crustacean Biology* 7,
425 1-24.
- 426 Gallagher, A., Rees, A., Rowe, R., Stevens, J., Wright, P. (2015) Microplastics in the Solent estuarine
427 complex, UK: an initial assessment. *Marine pollution bulletin*.
- 428 Goldstein, M.C., Goodwin, D.S. (2013) Gooseneck barnacles (*Lepas* spp.) ingest microplastic debris
429 in the North Pacific Subtropical Gyre. *PeerJ* 1, 184.
- 430 Graham, E.R., Thompson, J.T. (2009) Deposit- and suspension-feeding sea cucumbers
431 (Echinodermata) ingest plastic fragments. *Journal of experimental marine biology and ecology*
432 368, 22-29.
- 433 Hines, A.H. (1991) Fecundity and Reproductive Output in Nine Species of Cancer crabs (Crustacea,
434 Brachyura, Cancridae). *Canadian Journal of Fisheries and Aquatic Sciences* 48, 267-275.
- 435 Lizárraga-Cubedo, H.A., Tuck, I., Bailey, N., Pierce, G.J., Kinnear, J.A.M. (2003) Comparisons of size
436 at maturity and fecundity of two Scottish populations of the European lobster, *Homarus*
437 *gammarus*. *Fisheries Research* 65, 137-152.
- 438 Lusher, A., McHugh, M., Thompson, R. (2013) Occurrence of microplastics in the gastrointestinal
439 tract of pelagic and demersal fish from the English Channel. *Marine pollution bulletin* 67, 94-99.
- 440 Lusher, A.L., O'Donnell, C., Officer, R., O'Connor, I. (2015) Microplastic interactions with North
441 Atlantic mesopelagic fish. *ICES Journal of Marine Science: Journal du Conseil*.
- 442 Lutz, P.L., (1990) Studies on the ingestion of plastic and latex by sea turtles, in: Shomura, R.S.,
443 Godfrey, M.L. (Eds.), *Second International Conference on Marine Debris 2-7 April 1989. NOAA:*
444 *Panama City, 2-7 April 1989, pp. 719-735.*
- 445 McCauley, S.J., Bjorndal, K.A. (1999) Conservation Implications of Dietary Dilution from Debris
446 Ingestion: Sublethal Effects in Post-Hatchling Loggerhead Sea Turtles Implicaciones para la
447 Conservación, Dilución de Dietas por Ingestión de Basura: Efectos Subletales en Crías de la
448 Tortuga Marina *Caretta caretta*. *Conservation Biology* 13, 925-929.
- 449 Mente, E. (2010) Survival, food consumption and growth of Norway lobster (*Nephrops norvegicus*)
450 kept in laboratory conditions. *Integrative Zoology* 5, 256-263.
- 451 Moore, C.J., Moore, S.L., Leecaster, M.K., Weisberg, S.B. (2001) A Comparison of Plastic and
452 Plankton in the North Pacific Central Gyre. *Marine pollution bulletin* 42, 1297-1300.
- 453 Moreira, F.T., Prantoni, A.L., Martini, B., de Abreu, M.A., Stoiev, S.B., Turra, A. Small-scale temporal
454 and spatial variability in the abundance of plastic pellets on sandy beaches: Methodological
455 considerations for estimating the input of microplastics. *Marine pollution bulletin*.
- 456 Moreira, F.T., Prantoni, A.L., Martini, B., de Abreu, M.A., Stoiev, S.B., Turra, A. (2016) Small-scale
457 temporal and spatial variability in the abundance of plastic pellets on sandy beaches:
458 Methodological considerations for estimating the input of microplastics. *Marine pollution bulletin*
459 102, 114-121.
- 460 Murray, F., Cowie, P.R. (2011) Plastic contamination in the decapod crustacean *Nephrops*
461 *norvegicus* (Linnaeus, 1758). *Marine pollution bulletin* 62, 1207-1217.
- 462 Reddy, M.S., Shaik, B., Adimurthy, S., Ramachandraiah, G. (2006) Description of the small plastics
463 fragments in marine sediments along the Alang-Sosiya ship-breaking yard, India. *Estuarine,*
464 *Coastal and Shelf Science* 68, 656-660.

- 465 Ryan, P.G. (1988) Effects of ingested plastic on seabird feeding: Evidence from chickens. Marine
466 pollution bulletin 19, 125-128.
- 467 Ryan, P.G., Moore, C.J., van Franeker, J.A., Moloney, C.L. (2009) Monitoring the abundance of plastic
468 debris in the marine environment. Philosophical Transactions of the Royal Society B: Biological
469 Sciences 364, 1999-2012.
- 470 Setälä, O., Norkko, J., Lehtiniemi, M. (2015) Feeding type affects microplastic ingestion in a coastal
471 invertebrate community. Marine pollution bulletin.
- 472 Shaw, D.G., Mapes, G.A. (1979) Surface circulation and the distribution of pelagic tar and plastic.
473 Marine pollution bulletin 10, 160-162.
- 474 Teuten, E.L., Rowland, S.J., Galloway, T.S., Thompson, R.C. (2007) Potential for Plastics to
475 Transport Hydrophobic Contaminants. Environmental science & technology 41, 7759-7764.
- 476 Teuten, E.L., Saquing, J.M., Knappe, D.R.U., Barlaz, M.A., Jonsson, S., Björn, A., Rowland, S.J.,
477 Thompson, R.C., Galloway, T.S., Yamashita, R., Ochi, D., Watanuki, Y., Moore, C., Viet, P.H., Tana, T.S.,
478 Prudente, M., Boonyatumanond, R., Zakaria, M.P., Akkhang, K., Ogata, Y., Hirai, H., Iwasa, S.,
479 Mizukawa, K., Hagino, Y., Imamura, A., Saha, M., Takada, H. (2009) Transport and release of
480 chemicals from plastics to the environment and to wildlife. Philosophical Transactions of the
481 Royal Society B: Biological Sciences 364, 2027-2045.
- 482 Tully, O., Roantree, V., Robinson, M. (2001) Maturity, fecundity and reproductive potential of the
483 European lobster (*Homarus gammarus*) in Ireland. Journal of the Marine Biological Association of
484 the United Kingdom 81, 61-68.
- 485 Ugolini, A., Ungherese, G., Ciofini, M., Lapucci, A., Camaiti, M. (2013) Microplastic debris in
486 sandhoppers. Estuarine, Coastal and Shelf Science 129, 19-22.
- 487 Van Cauwenberghe, L., Janssen, C.R. (2014) Microplastics in bivalves cultured for human
488 consumption. Environmental Pollution 193, 65-70.
- 489 Van Cauwenberghe, L., Vanreusel, A., Mees, J., Janssen, C.R. (2013) Microplastic pollution in deep-
490 sea sediments. Environmental Pollution 182, 495-499.
- 491 van Franeker, J.A., Meijboom, A., de Jong, M.L., (2004) Marine litter monitoring by Northern
492 Fulmars in the Netherlands 1982-2003., Alterra-rapport 1093. Alterra, Wageningen.
- 493 Vianello, A., Boldrin, A., Guerriero, P., Moschino, V., Rella, R., Sturaro, A., Da Ros, L. (2013)
494 Microplastic particles in sediments of Lagoon of Venice, Italy: First observations on occurrence,
495 spatial patterns and identification. Estuarine, Coastal and Shelf Science 130, 54-61.
- 496 von Moos, N., Burkhardt-Holm, P., Köhler, A. (2012) Uptake and Effects of Microplastics on Cells
497 and Tissue of the Blue Mussel *Mytilus edulis* L. after an Experimental Exposure. Environmental
498 science & technology 46, 11327-11335.
- 499 Watts, A.J., Lewis, C., Goodhead, R.M., Beckett, S.J., Moger, J., Tyler, C.R., Galloway, T.S. (2014)
500 Uptake and retention of microplastics by the shore crab *Carcinus maenas*. Environmental science
501 & technology 48, 8823-8830.
- 502 Watts, A.J., Urbina, M.A., Corr, S., Lewis, C., Galloway, T.S. (2015) Ingestion of Plastic Microfibers by
503 the Crab *Carcinus maenas* and Its Effect on Food Consumption and Energy Balance.
504 Environmental science & technology 49, 14597-14604.
- 505 Welden, N.A., Taylor, A.C., Cowie, P.R. (2015) Growth and gut morphology of the lobster. Journal of
506 Crustacean Biology 35, 20-25.