

Recent Changes in Patterns of Mammal Infection with Highly Pathogenic Avian Influenza A(H5N1) Virus Worldwide

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We reviewed information about mammals naturally infected by highly pathogenic avian influenza A virus subtype H5N1 during 2 periods: the current panzootic (2020–2023) and previous waves of infection (2003–2019). In the current panzootic, 26 countries have reported ≥ 48 mammal species infected by H5N1 virus; in some cases, the virus has affected thousands of individual animals. The geographic area and the number of species affected by the current event are considerably larger than in previous waves of infection. The most plausible source of mammal infection in both periods appears to be close contact with infected birds, including their ingestion. Some studies, especially in the current panzootic, suggest that mammal-to-mammal transmission might be responsible for some infections; some mutations found could help this avian pathogen replicate in mammals. H5N1 virus may be changing and adapting to infect mammals. Continuous surveillance is essential to mitigate the risk for a global pandemic.

Since last century, highly pathogenic avian influenza (HPAI) viruses have caused diverse waves of infection (1). However, the ongoing panzootic event (2020–2023) caused by HPAI A(H5N1) virus could become one of the most important in terms of economic losses, geographic areas affected, and numbers of species and individual animals infected (1–4). This pathogen appears to be emerging in several regions of the world

(e.g., South America); it has caused death in domestic and wild birds but also in mammals (2,5,6). This trend is of great concern because it may indicate a change in the dynamics of this pathogen (i.e., an increase in their range of hosts and the severity of the disease) (3).

H5N1 has affected several mammal species since 2003 (6,7), thus raising concern because H5N1 mammalian adaptation could represent a risk not only for diverse wild mammals but also for human health (8–10). Unfortunately, information about this topic, especially related to the current panzootic (2020–2023), is dispersed and available often only in gray literature (e.g., databases and official government websites). This fact complicates access and evaluation for many stakeholders working on the front lines (e.g., wildlife managers, conservationists, and public health authorities at regional and local levels).

For this article, we compiled and analyzed information from scientific literature about mammal species, including humans, naturally affected by the current panzootic event and compared those findings with the outcomes of previous waves of H5N1 infection. We focus particularly on the species infected, their habitat, phylogeny, and trophic level, and the sources of infection, virus mutations, clinical signs, and necropsy findings associated with this virus. We also address potential risks for biodiversity and human health.

Methods

We compiled scientific information on mammals infected by H5N1 virus through October 2023. We considered only scientific information on mammal species infected naturally (i.e., experimental studies were not included). We performed 2 systematic searches in Scopus and Google Scholar, first using the terms “H5N1 AND mammal”; this search was divided into 2 periods (1996–2019 and 2020–2023) (Appendix Figure

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DOI: <https://doi.org/10.3201/eid3003.231098>

1, 2, <https://wwwnc.cdc.gov/EID/article/30/3/23-1098-App1.pdf>). We then performed an additional search with no time restriction using the following key terms: “H5N1 OR HPAI OR Highly Pathogenic Avian Influenza AND mammal OR unusual host.” This additional search contributed no new articles on the study topic (Appendix Figure 3). We also adopted a snowball approach, examining all the references in the articles we found in our searches. We included review articles only if they contributed new information about mammal species infected naturally with H5N1; we excluded articles based on serologic surveys because of the difficulty in determining when infection occurred, which can introduce uncertainty into the diagnosis (11).

To obtain additional information on the current panzootic event, we also searched the following official databases: World Organisation for Animal Health (6), the US Department of Agriculture’s Animal and Plant Health Service (12), and the United Kingdom’s Animal and Plant Health Agency (13). To obtain information about humans affected by this pathogen we used information provided by the World Health Organization (14). We constructed a map with the countries with reports of mammal infections (Figure 1) and the phylogeny of mammal species affected by H5N1 (Figures 2, 3) by using iTOL version 5, following Letunic and Bork (15), from DNA sequence data available in Upham et al. (16). We retrieved the conservation statuses of infected mammals from International Union for Conservation of Nature Red List of Threatened Species (17) and information on their diets from that database and MammalBase (18).

Results and Discussion

Scientific Information Available

We found 59 scientific articles on mammals infected naturally by H5N1 virus, 23 from previous waves of infection (up to 2019) and 36 from the current panzootic event (Appendix Figure 1, 2). The articles reporting mammals infected naturally in previous waves were published during 2004–2018, whereas those addressing the current panzootic were published during 2021–2023. The current panzootic has thus generated more articles in 3 years than all the previous waves of infection (published over a 15-year period). This fact suggests increased general interest in emerging pathogens affecting biodiversity and mammals (wild and farmed) and also that the current panzootic event is causing greater concern and having a greater effect than previous ones (considering the geographic regions and mammal species affected) (4).

Geographic Localization of Information and Mammal Species Affected

During previous waves of infection, 10 countries reported mammals (not including humans) naturally infected by H5N1 (5 countries in Asia, 3 in Europe, and 2 in Africa) (Figure 1, panel A; Appendix Table). In the current event, 26 countries have reported information on mammals (not including humans) infected by this virus; most information is from Europe (17 countries), followed by South America (5 countries), North America (2 countries), and Asia (2 countries) (Figure 1, panel B; Appendix Table). To the best of our knowledge, for the current outbreak, no information is available on mammals from other parts of the world, which can probably be explained by a lack of testing or reporting of cases. Our review suggests that H5N1 virus is expanding its geographic range to new continents such as North and South America (Figure 1). This fact is of concern because when an emerging pathogen reaches naive populations, the consequences for biodiversity can be catastrophic, especially for threatened species (19).

We found that previous waves of infection affected several mammals around the world (7,20); for example, tigers (*Panthera tigris*), leopards (*Panthera pardus*), domestic cats (*Felis catus*), domestic dogs (*Canis lupus familiaris*), Owston’s palm civet (*Chrotogale owstoni*), stone martens (*Martes foina*), plateau pikas (*Ochotona curzoniae*), minks (*Neovison vison*), and raccoon dogs (*Nyctereutes procyonoides*) (Appendix Table). All the mammal species affected were terrestrial or semiaquatic species (Figure 2, panel A). Most mammals infected during previous waves (75%; n = 9) belong to the order Carnivora, whereas the remainder correspond to the Lagomorpha, Artiodactyla, and Perissodactyla orders (Figure 2, panel B). Infected mammal species included top predators (e.g., tigers and leopards) and some mesopredators (e.g., minks) (Appendix Table). Most species infected in previous waves were carnivores (n = 6) and omnivores (n = 4), followed by herbivores (n = 2) (Figure 2, panel C; Appendix Table).

So far, in the current panzootic, ≥ 48 mammal species from disparate regions of the world have been reported as naturally infected by H5N1 (Appendix Table). Most of those species (n = 35) are terrestrial or semiaquatic mammals (Figure 3, panel A; Appendix Table), but 13 species of marine mammals also were affected, resulting in massive deaths (up to thousands of individual animals) in geographic regions such as Peru, Chile, and Argentina (Figure 3, panel A; Appendix Table). Of the total number of mammals infected, 81% (n = 39) belong to the order

Carnivora, and the remainder correspond to Didelphimorphia, Rodentia, and Cetartiodactyla (Figure 3, panel B). Infected mammal species include top predators (e.g., mountain lion [*Puma concolor*]) and several mesopredators (e.g., red fox [*Vulpes vulpes*]) (Appendix Table). Most mammal species infected are carnivores (n = 34), followed by omnivores (n = 13) and herbivores (n = 1); some of those species (n = 13) also are considered facultative scavengers (i.e., they include in their diet a considerable quantity of carrion; in our case to be a facultative scavenger

carrion should be named in the diet) (Figure 3, panel C; Appendix Table).

The species infected in the 2 events show similarities. Most species belong to the order Carnivora and are top or mesopredators with a carnivorous diet; some species also are facultative scavengers. However, in the current panzootic event, the diverse marine mammals affected have suffered massive deaths (e.g., American sea lion [*Otaria flavescens*]) (Appendix Table). Marine mammals have been affected by other influenza viruses such as H10N7 (21), but the species

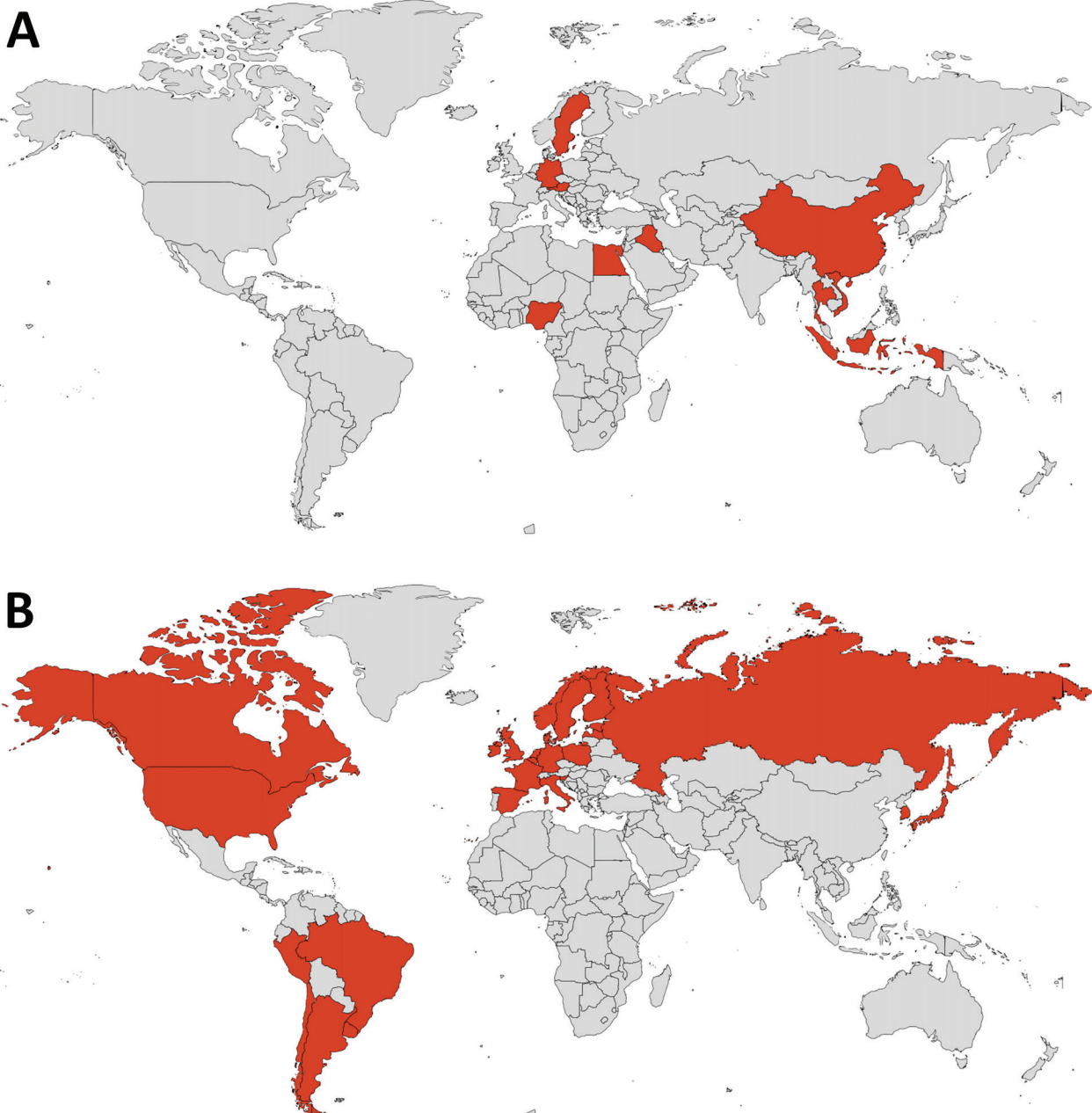


Figure 1. Geographic location of mammal species affected by highly pathogenic influenza virus A(H5N1) in previous waves of infection, 2003–2019 (A), and in the current panzootic, 2020–2023 (B).

affected and the number of dead individual animals attributable to the current event is of great concern (22,23); for example, the proportion of American sea lions that died in Peru represents 5% of their population there (22).

The current panzootic is ongoing, and the number of species being infected naturally is increasing (40 new mammal species have been reported as infected by this pathogen during the current panzootic), so the effect on mammal species may continue to worsen with time. This effect could just be attributable to the current high H5N1 infection rates throughout the world, which means the virus is reaching more areas and mammal species living in these places (i.e., high environmental circulation of this pathogen) (8). However, the dynamics of the virus may also be changing (3), in which case its infectivity in unusual species such as mammals is probably increasing (8). During the final review process of this article, 2 additional species were reported to be infected by this virus in the United States: the Abert's squirrel (*Sciurus aberti*) and the polar bear (*Ursus maritimus*) (newly infected species are not shown in figures or the Appendix Table) (6).

Source of Infection

Although the source of infection in mammals is often unknown, most scientific information available during previous and the current H5N1 event suggests that the most plausible source of infection is close contact with infected birds, including their ingestion, which may occur through predation of sick individual animals or scavenging on carcasses. For instance, in the year 2004, a total of 147 tigers and 2 leopards housed in zoos in Thailand became infected and died after consuming infected chicken carcasses (24,25). In China, this infection source was also associated with the death of a tiger in 2013 (26) and a lion in 2016 (27). In the current panzootic, the first case of H5N1 infection in minks in Spain was probably caused by contact with infected birds (perhaps gulls) (9). Ingestion of infected bird carcasses was probably the route of infection of red foxes in the Netherlands, Finland, and Japan during 2020–2022 (28–31), American sea lions in Peru in 2023 (22), diverse mesocarnivores in Canada during 2021–2022 (32) and otters (*Lutra lutra*) and a lynx (*Lynx lynx*) in Finland in 2021–2022 (31). Of concern, studies in infected tigers, farmed minks, and social species such as American sea lions, raise an alarm that mammal-to-mammal transmission may have occurred (9,22,24,33), but further research is needed to confirm this possibility.

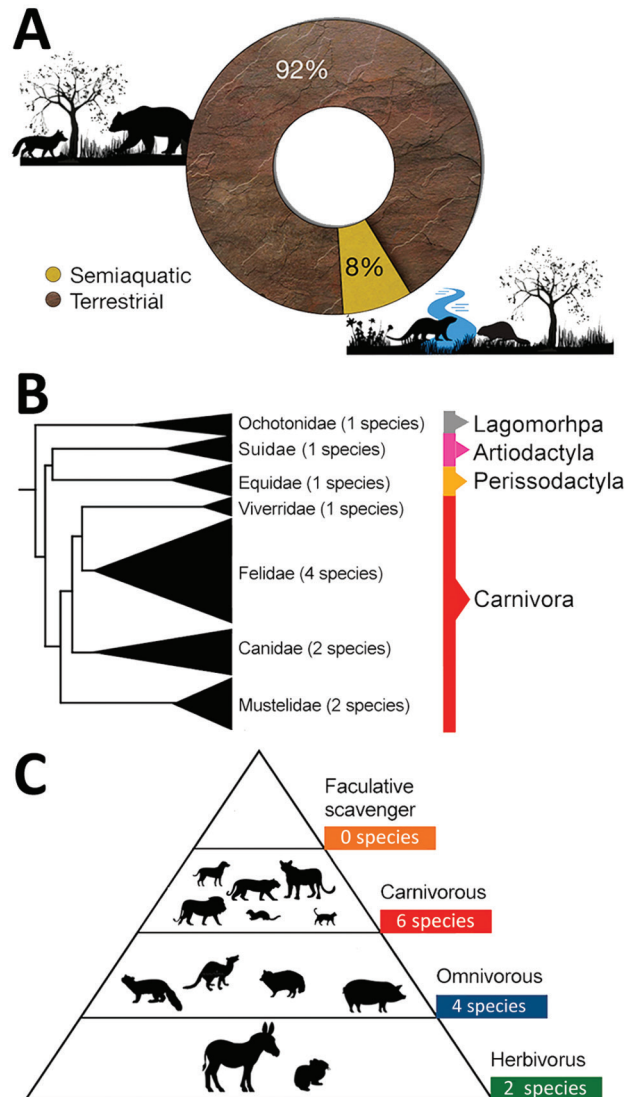


Figure 2. Characteristics of mammal species affected worldwide by highly pathogenic influenza virus A (H5N1) in previous waves of infection (2003–2019). A) Habitat of mammal species affected by H5N1. B) Phylogeny of mammal species affected (tree constructed using iTOL version 5 following Letunic and Bork [15], from DNA sequence data available in Upham et al. [16]). C) Trophic level (facultative scavenger, carnivore, omnivore, or herbivore) of mammalian species affected worldwide by H5N1.

If mammal-to-mammal transmission occurs during the current H5N1 panzootic, such transmission could imply that the virus mutated to enable virus replication in mammal tissues (9). Some researchers have reported mutations compatible with adaptation to mammal replication (9,25,33,34), which is concerning and requires attention. However, evaluating whether those mutations happen in wild birds before mammal infections or arise de novo in mammals after infection is important.

Mutations Found

Through sequencing of the H5N1 viruses infecting mammals, some relevant mutations such as E627K in polymerase basic protein 2 (PB2) (PB2-E627K) and D701N in polymerase basic protein 2 (PB2) (PB2-D701N) have been found in previous waves and in the current panzootic (Appendix Table). Those mutations are commonly associated with virulence and efficiency in the replication of this pathogen in

mammals (31,33,35). For instance, during 2004–2005, in Thailand, the isolated H5N1 viruses that infected tigers, a domestic cat, a domestic dog, and a leopard contained the PB2-E627K mutation (25,35,36). In the current panzootic, red foxes from the Netherlands also showed the mammalian adaptation of PB2-E627K (28). In viruses collected from red foxes, an otter, and a lynx in Finland in 2021–2022, the PB2-E627K and PB2-D701N mutations were identified (the latter mutation was reported in 1 red fox and 1 lynx in Finland) (31). Similarly, in the current panzootic, red foxes, otters, and polecats (*Mustela putorius*) in the Netherlands, and red foxes in Canada, and the United States had the PB2-E627K mutation (8,32,37). The PB2-E627K and PB2-D701N mutations were also detected in harbor seals (*Phoca vitulina*) in the United States (34), and the latter mutation was found in South American sea lions in Peru (33), and in a red fox in Canada (32). In both previous and current events, other mutations meriting further research were also found in diverse mammal species, including terrestrial, semiaquatic, and marine mammals (Appendix Table).

Mutations that facilitate replication of the virus in mammal hosts (e.g., enhancing polymerase activity in mammal cells), such as PB2-E627K and PB2-D701N, could be of concern (8,31,33). Potential mutations must be continuously scrutinized to detect whether the H5N1 virus is adapting to mammal-to-mammal transmission. This approach is important for wildlife conservation because if such transmission occurs, the consequences for threatened mammal species could be severe (e.g., threatened South American sea lion deaths in Peru [22]). In addition, mutations must be monitored for changes that may favor transmission to and between humans, which would increase the risk for a pandemic.

Clinical Signs of H5N1 in Mammals

The most common clinical signs reported in infected mammals, both in previous waves and the current H5N1 panzootic, are neurologic and respiratory. For instance, in 2005, an infected Owston’s civet in Vietnam showed loss of appetite and neurologic signs such as convulsions and paralysis; the same clinical signs were reported in a stone marten in Germany in 2006 (38,39). Similarly, hundreds of infected tigers in a zoo in Thailand showed respiratory and neurologic signs before they died (24). In the current panzootic event, infected minks from Spain manifested loss of appetite, hyper salivation, depression, bloody snout, and neurologic signs such as ataxia and tremors (9). American sea lions in Peru and harbor seals in the

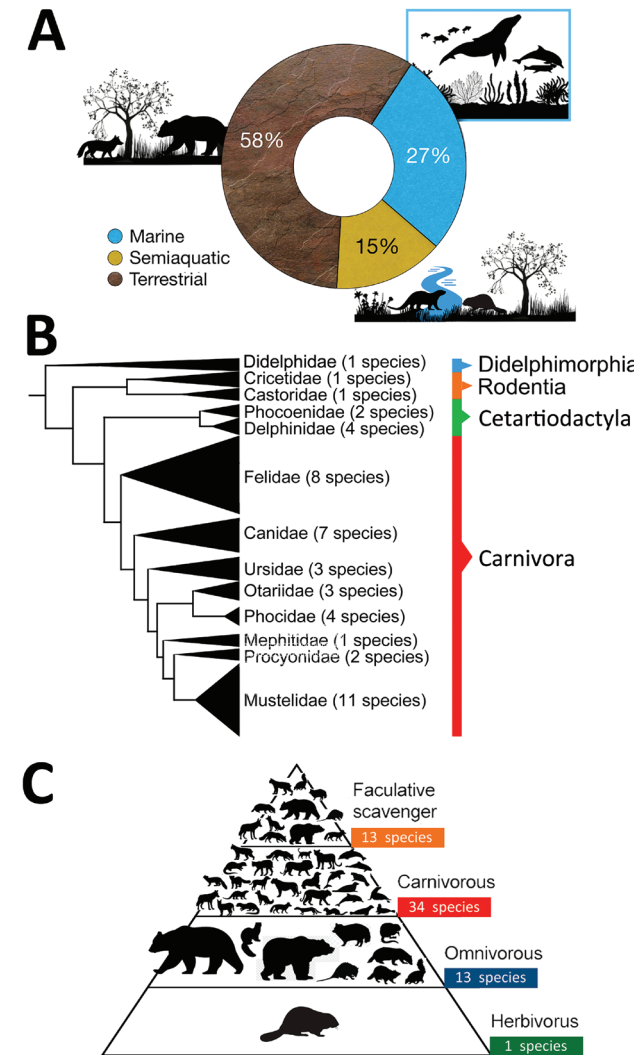


Figure 3. Characteristics of mammal species affected worldwide by highly pathogenic influenza virus A (H5N1) the current panzootic (2020–2023). A) Habitat of mammal species affected by H5N1. B) Phylogeny of mammal species affected (tree constructed using iTOL version 5 following Letunic and Bork [15], from DNA sequence data available in Upham et al. [16]). C) Trophic level (facultative scavenger, carnivore, omnivore, or herbivore) of mammal species affected worldwide by H5N1. Some of the omnivorous and carnivorous mammals included in the pyramid (n = 13) also consume carrion; thus, they are also considered to be facultative scavengers and are incorporated in the upper part of the pyramid.

United States showed respiratory signs (dyspnea and whitish secretions in nares) and neurologic signs (tremors and convulsions) (22,34). Red foxes, an otter, a polecat, and a badger (*Meles meles*) in the Netherlands had neurologic signs such as convulsions and head shaking (8,30). In Finland, an infected otter was also reported to have a set of neurologic signs (31). Finally, in the United States and Canada, several mammals manifested neurologic and respiratory signs (32,37). Those findings suggest that H5N1 virus has neurotropism in mammals, as reported in birds (6,28), causing severe disease and pathologic lesions (e.g., encephalitis); brain samples should be included in wildlife surveillance programs for reliable detection of the H5N1 virus in mammals (8).

Although neurologic and respiratory signs are commonly reported in mammals infected with H5N1, some species and individual animals show subclinical disease. For instance, infected pigs (*Sus scrofa domestica*) from Indonesia, Nigeria, and China had no signs of influenza but tested positive for H5N1 (40–42). Similarly, in Austria, infected domestic cats display asymptomatic infections (43). Subclinical infections are concerning because they are not easily detected; infected individual animals may be transmitting the virus to other species and even humans, representing a risk to the ecosystem and human health (40,41).

Necropsy Findings

In previous waves of infection and the current H5N1 panzootic, the most frequently reported anatomopathologic lesions in infected mammals were pneumonia and encephalitis. Those kinds of lesions (e.g., congestion of brain, meningoencephalitis, hemorrhagic lungs, and pleural effusion) were reported in dead tigers in Thailand and China during 2004–2014 (24,26,44), in a lion in China in 2016 (27), and in cats and dogs infected naturally in Thailand in 2004 (45,46). In the current panzootic, for instance, red foxes from the Netherlands had collapsed lungs with a marbled red aspect; histopathologic analyses showed a subacute to chronic purulent granulomatous broncho-interstitial pneumonia and nonsuppurative encephalitis with perivascular cuffing (28). Red foxes, polecats, otters, and a badger in the Netherlands also showed nonsuppurative meningitis, encephalitis, or meningoencephalitis, all with differences in severity (8). American sea lions in Peru had congestive brains compatible with encephalitis (22). A porpoise (*Phocoena phocoena*) in Sweden manifested meningoencephalitis (47). Similar findings, meningoencephalitis and pneumonia, were also found in mammals in Finland, the United States, and Canada (31,32,37).

Those findings suggest that respiratory and neurologic lesions are the most common pathologies of necropsied mammals infected with H5N1 in both previous waves of infection and the current panzootic. The lesions largely explain the neurologic and respiratory signs observed in mammals affected by this virus. Complete necropsies of infected mammals may help determine whether those anatomopathologic findings are frequent and pathognomonic for this disease in every species and most individual animals, as preliminary results suggest.

Risks for Biodiversity

The current panzootic is affecting a larger number of species around the world than previous waves of H5N1 infection, and some are of conservation concern. Previous waves affected 2 endangered and 2 vulnerable species (Appendix Table). The current panzootic has so far affected 4 near threatened, 4 endangered, 3 vulnerable, and 1 critically endangered species (Appendix Table); this emerging pathogen may affect species of conservation concern, exacerbating their situation.

In general, most mortality events associated with the current panzootic appear to affect few individual animals and in only certain areas; thus far, large populations have not been affected in the way wild birds have been affected (4,6). However, this virus is suspected of producing massive deaths in some marine mammals; for example, >20,000 South American sea lions were reported to have died suddenly, and many individual animals tested positive for H5N1 (6,22,23). This fact raises concern as to the potential effect of this virus on the demography of some threatened mammal populations. This emerging pathogen represents a new species invading and impacting new environments and species and could therefore constitute a new threat for diverse species currently threatened by human action (e.g., land use change, contamination, and habitat loss) (19,48).

Potential Risks for Human Health

During 2003–2023, a total of 878 humans tested positive for the H5N1 virus, and 458 deaths were reported, indicating a lethality of $\approx 52\%$ (14). During 2003–2019, most human cases came from Asia and Africa, particularly from China ($n = 53$), Egypt ($n = 359$), and Indonesia ($n = 200$). From 2020 through July 2023, human cases of H5N1 infection occurred in diverse countries, such as Laos (1 case), India (1 case), United Kingdom (4 cases), China (2 cases), the United States (1 case), Vietnam (1 case), Spain (2 cases), Ecuador (1 case), Chile (1 case), and Cambodia (2 cases) (14).

Those recent cases resulted in ≥ 3 deaths (14). Of note, this zoonotic virus has produced human cases in new geographic areas, such as South America.

The spillover to humans has been associated with close contact between humans and infected animals, particularly poultry; this kind of contact is relatively common in some geographic regions (even close contact between dead mammals and humans, as in Peru [22]). So far, no evidence indicates human-to-human transmission, and the risk for a pandemic event still seems low (8). However, one of the most severe influenza viruses to have affected humans (i.e., Spanish influenza [1918–1919]) developed from an avian influenza virus that adapted to humans (49), a fact that should be considered when assessing the spillover risk.

Mutations in the virus found in diverse mammal species, especially in the current panzootic, are of great concern. For instance, the T271A mutation reported in minks in Spain is also present in the H1N1 that produced a pandemic in 2009 (9). Similarly, the PB2-E627K mutation found in this virus in diverse geographic areas could indicate an adaptation for replication in mammals (28,31). Moreover, some infected species, such as minks, may act as a mixing vessel for interspecies transmission between birds, mammals, and humans (9). Mutations and infections with H5N1 in potential mixing-vessel species (e.g., minks and wild and domestic pigs) should be followed closely because of the potential risk to human health.

Final Considerations

Given the magnitude of the current H5N1 panzootic, continuous surveillance is necessary to identify any increase in risk to biodiversity and human health. It is therefore essential that all affected countries share all their available information (e.g., genomic data of the H5N1 virus, species, and number of individual animals affected). We urge that all findings be shared quickly. International collaboration must be intensified to obtain rapid results; some less-developed regions have technologic and logistic barriers that hinder the production and analysis of information on the impact of this virus, and they may need help. There is a need for strong collaborative work between countries and institutions in preparation for any spillover that may lead to a mammalian panzootic or human pandemic.

It is fundamental that we rethink the interface between humans, domestic animals, and wild animals to prevent the emergence of dangerous pathogens that affect biodiversity and human health (48). Governments must assume responsibility for protecting biodiversity and human health from diseases caused

by human activities, particularly diseases originating from intensive production (50), such as this H5N1 avian influenza virus. If we hope to conserve biodiversity and protect human health, we must change the way we produce our food (poultry farming, in this specific case) and how we interact with and affect wildlife.

Financial support was provided by Consejo Nacional de Investigaciones Científicas y Técnicas, Agencia Nacional de Promoción Científica y Tecnológica (grant no. PICT-2021-TI-00039), Universidad Nacional del Comahue (project 04/B227, grant to S.A.L.), and Aves Argentinas (grant to P.P.).

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References

- Shi J, Zeng X, Cui P, Yan C, Chen H. Alarming situation of emerging H5 and H7 avian influenza and effective control strategies. *Emerg Microbes Infect.* 2023;12:2155072. <https://doi.org/10.1080/22221751.2022.2155072>
- Wille M, Barr IG. Resurgence of avian influenza virus. *Science.* 2022;376:459–60. <https://doi.org/10.1126/science.abo1232>
- Harvey JA, Mullinax JM, Runge MC, Prosser DJ. The changing dynamics of highly pathogenic avian influenza H5N1: next steps for management and science in North America. *Biol Conserv.* 2023;282:110041. <https://doi.org/10.1016/j.biocon.2023.110041>
- Adlhoch C, Fusaro A, Gonzales JL, Kuiken T, Mirinaviciute G, Niqueux É, et al.; European Food Safety Authority, European Centre for Disease Prevention and Control, European Union Reference Laboratory for Avian Influenza. Avian influenza overview March–April 2023. *EFSA J.* 2023;21:e08039.
- Gamarra-Toledo V, Plaza PI, Angulo F, Gutiérrez R, García-Tello O, Saravia-Guevara P, et al. Highly pathogenic avian influenza (HPAI) strongly impacts wild birds in Peru. *Biol Conserv.* 2023;286:110272. <https://doi.org/10.1016/j.biocon.2023.110272>
- World Organization for Animal Health. WAHIS: World Animal Health Information System. 2023 [cited 2023 Oct 30]. <https://wahis.woah.org>
- Reperant LA, Rimmelzwaan GF, Kuiken T. Avian influenza viruses in mammals. *Rev Sci Tech.* 2009;28:137–59. <https://doi.org/10.20506/rst.28.1.1876>
- Vreman S, Kik M, Germeraad E, Heutink R, Harders F, Spierenburg M, et al. Zoonotic mutation of highly pathogenic avian influenza H5N1 virus identified in the brain of multiple wild carnivore species. *Pathogens.* 2023;12:168. <https://doi.org/10.3390/pathogens12020168>

9. Agüero M, Monne I, Sánchez A, Zecchin B, Fusaro A, Ruano MJ, et al. Highly pathogenic avian influenza A(H5N1) virus infection in farmed minks, Spain, October 2022. *Euro Surveill.* 2023;28:2300001. <https://doi.org/10.2807/1560-7917.ES.2023.28.3.2300001>
10. Kupferschmidt K. Bird flu spread between mink is a 'warning bell'. *Science.* 2023;379:316–7. <https://doi.org/10.1126/science.adg8342>
11. Horimoto T, Maeda K, Murakami S, Kiso M, Iwatsuki-Horimoto K, Sashika M, et al. Highly pathogenic avian influenza virus infection in feral raccoons, Japan. *Emerg Infect Dis.* 2011;17:714–7. <https://doi.org/10.3201/eid1704.101604>
12. US Department of Agriculture. 2022–2023 Detections of highly pathogenic avian influenza in mammals [cited 2023 Oct 30]. <https://www.aphis.usda.gov/aphis/ourfocus/animalhealth/animal-disease-information/avian/avian-influenza/hpai-2022/2022-hpai-mammals>
13. United Kingdom Animal and Plant Health Agency. Confirmed findings of influenza of avian origin in non-avian wildlife [cited 2023 Oct 30]. <https://www.gov.uk/government/publications/bird-flu-avian-influenza-findings-in-non-avian-wildlife/confirmed-findings-of-avian-origin-in-non-avian-wildlife>
14. World Health Organization. Cumulative number of confirmed human cases for avian influenza A(H5N1) reported to WHO, 2003–2023, 3 October 2023. 2023 [cited 2023 Oct 30]. [https://www.who.int/publications/m/item/cumulative-number-of-confirmed-human-cases-for-avian-influenza-a\(h5n1\)-reported-to-who--2003-2023--3-october-2023](https://www.who.int/publications/m/item/cumulative-number-of-confirmed-human-cases-for-avian-influenza-a(h5n1)-reported-to-who--2003-2023--3-october-2023)
15. Letunic I, Bork P. Interactive Tree Of Life (iTOL) v5: an online tool for phylogenetic tree display and annotation. *Nucleic Acids Res.* 2021;49(W1):W293–6. <https://doi.org/10.1093/nar/gkab301>
16. Upham NS, Esselstyn JA, Jetz W. Inferring the mammal tree: species-level sets of phylogenies for questions in ecology, evolution, and conservation. *PLoS Biol.* 2019;17:e3000494. <https://doi.org/10.1371/journal.pbio.3000494>
17. IUCN. IUCN red list of threatened species. 2023 [cited 2023 Oct 30]. <https://www.iucnredlist.org>
18. MammalBase. Database of recent mammals [cited 2023 Oct 30]. <https://www.mammalbase.net/mb>
19. Dobson A, Foufopoulos J. Emerging infectious pathogens of wildlife. *Philos Trans R Soc Lond B Biol Sci.* 2001;356:1001–12. <https://doi.org/10.1098/rstb.2001.0900>
20. Root J, Shriner S. Avian influenza A virus associations in wild, terrestrial mammals: a review of potential synanthropic vectors to poultry facilities. *Viruses.* 2020;12:1352. <https://doi.org/10.3390/v12121352>
21. Bodewes R, Bestebroer TM, van der Vries E, Verhagen JH, Herfst S, Koopmans MP, et al. Avian influenza A(H10N7) virus-associated mass deaths among harbor seals. *Emerg Infect Dis.* 2015;21:720–2. <https://doi.org/10.3201/eid2104.141675>
22. Gamarra-Toledo V, Plaza P, Inga G, Gutiérrez R, García-Tello O, Valdivia-Ramírez L, et al. Mass mortality of sea lions caused by highly pathogenic influenza virus (H5N1) in South America. *Emerg Infect Dis.* 2023;29:2553–6. <https://doi.org/10.3201/eid2912.230192>
23. OFFLU Ad-Hoc Group on HPAI H5 in Wildlife of South America and Antarctica. Southward expansion of high pathogenicity avian influenza H5 in wildlife in South America: estimated impact on wildlife populations, and risk of incursion into Antarctica. 2023 [cited 2023 Oct 30]. <https://www.offlu.org/wp-content/uploads/2023/08/OFFLU-statement-HPAI-wildlife-South-America-20230823.pdf>
24. Thanawongnuwech R, Amonsin A, Tantilertcharoen R, Damrongwatanapokin S, Theamboonlers A, Payungporn S, et al. Probable tiger-to-tiger transmission of avian influenza H5N1. *Emerg Infect Dis.* 2005;11:699–701. <https://doi.org/10.3201/eid1105.050007>
25. Keawcharoen J, Oraveerakul K, Kuiken T, Fouchier RA, Amonsin A, Payungporn S, et al. Avian influenza H5N1 in tigers and leopards. *Emerg Infect Dis.* 2004;10:2189–91. <https://doi.org/10.3201/eid1012.040759>
26. He S, Shi J, Qi X, Huang G, Chen H, Lu C. Lethal infection by a novel reassortant H5N1 avian influenza A virus in a zoo-housed tiger. *Microbes Infect.* 2015;17:54–61. <https://doi.org/10.1016/j.micinf.2014.10.004>
27. Chen Q, Wang H, Zhao L, Ma L, Wang R, Lei Y, et al. First documented case of avian influenza (H5N1) virus infection in a lion. *Emerg Microbes Infect.* 2016;5:e125. <https://doi.org/10.1038/emi.2016.127>
28. Bordes L, Vreman S, Heutink R, Roose M, Venema S, Pritz-Verschuren SBE, et al. Highly pathogenic avian influenza H5N1 virus infections in wild red foxes (*Vulpes vulpes*) show neurotropism and adaptive virus mutations. *Microbiol Spectr.* 2023;11:e0286722. <https://doi.org/10.1128/spectrum.02867-22>
29. Hiono T, Kobayashi D, Kobayashi A, Suzuki T, Satake Y, Harada R, et al. Virological, pathological, and glycovirological investigations of an Ezo red fox and a tanuki naturally infected with H5N1 high pathogenicity avian influenza viruses in Hokkaido, Japan. *Virology.* 2023;578:35–44. <https://doi.org/10.1016/j.virol.2022.11.008>
30. Rijks JM, Hesselink H, Lollinga P, Wesselman R, Prins P, Weesendorp E, et al. Highly pathogenic avian influenza A (H5N1) virus in wild red foxes, the Netherlands, 2021. *Emerg Infect Dis.* 2021;27:2960–2. <https://doi.org/10.3201/eid2711.211281>
31. Tammiranta N, Isomursu M, Fusaro A, Nylund M, Nokireki T, Giussani E, et al. Highly pathogenic avian influenza A (H5N1) virus infections in wild carnivores connected to mass mortalities of pheasants in Finland [cited 2023 Oct 30]. *Infect Genet Evol.* 2023;111:105423.
32. Alkie TN, Cox S, Embury-Hyatt C, Stevens B, Pople N, Pybus MJ, et al. Characterization of neurotropic HPAI H5N1 viruses with novel genome constellations and mammalian adaptive mutations in free-living mesocarnivores in Canada. *Emerg Microbes Infect.* 2023;12:2186608. <https://doi.org/10.1080/22221751.2023.2186608>
33. Leguia M, García-Glaessner A, Muñoz-Saavedra B, Juárez D, Barrera P, Calvo-Mac C, et al. Highly pathogenic avian influenza A (H5N1) in marine mammals and seabirds in Peru. *Nat Commun.* 2023;14:5489. <https://doi.org/10.1038/s41467-023-41182-0>
34. Puryear W, Sawatzki K, Hill N, Foss A, Stone JJ, Doughty L, et al. Highly pathogenic avian influenza A(H5N1) virus outbreak in New England seals, United States. *Emerg Infect Dis.* 2023;29:786–91. <https://doi.org/10.3201/eid2904.221538>
35. Amonsin A, Payungporn S, Theamboonlers A, Thanawongnuwech R, Suradhat S, Pariyothorn N, et al. Genetic characterization of H5N1 influenza A viruses isolated from zoo tigers in Thailand. *Virology.* 2006;344:480–91. <https://doi.org/10.1016/j.virol.2005.08.032>
36. Amonsin A, Songserm T, Chutinimitkul S, Jam-On R, Sae-Heng N, Pariyothorn N, et al. Genetic analysis of influenza A virus (H5N1) derived from domestic cat and dog in Thailand. *Arch Virol.* 2007;152:1925–33. <https://doi.org/10.1007/s00705-007-1010-5>
37. Elsmo EJ, Wunschmann A, Beckmen KB, Broughton-Neiswanger LB, Buckles EL, Ellis J, et al.

- Pathology of natural infection with highly pathogenic avian influenza virus (H5N1) clade 2.3.4.4b in wild terrestrial mammals in the United States in 2022. *Emerg Infect Dis.* 2023;29:2451–60. <https://doi.org/10.3201/eid2912.230464>
38. Robertson SI, Bell DJ, Smith GJD, Nicholls JM, Chan KH, Nguyen DT, et al. Avian influenza H5N1 in viverrids: implications for wildlife health and conservation. *Proc Biol Sci.* 2006;273:1729–32. <https://doi.org/10.1098/rspb.2006.3549>
 39. Klopffleisch R, Wolf PU, Wolf C, Harder T, Starick E, Niebuhr M, et al. Encephalitis in a stone marten (*Martes foina*) after natural infection with highly pathogenic avian influenza virus subtype H5N1. *J Comp Pathol.* 2007; 137:155–9. <https://doi.org/10.1016/j.jcpa.2007.06.001>
 40. Nidom CA, Takano R, Yamada S, Sakai-Tagawa Y, Daulay S, Aswadi D, et al. Influenza A (H5N1) viruses from pigs, Indonesia. *Emerg Infect Dis.* 2010;16:1515–23. <https://doi.org/10.3201/eid1610.100508>
 41. Meseko C, Globig A, Ijomanta J, Joannis T, Nwosuh C, Shamaki D, et al. Evidence of exposure of domestic pigs to highly pathogenic avian influenza H5N1 in Nigeria. *Sci Rep.* 2018;8:5900. <https://doi.org/10.1038/s41598-018-24371-6>
 42. He L, Zhao G, Zhong L, Liu Q, Duan Z, Gu M, et al. Isolation and characterization of two H5N1 influenza viruses from swine in Jiangsu Province of China. *Arch Virol.* 2013;158:2531–41. <https://doi.org/10.1007/s00705-013-1771-y>
 43. Leschnik M, Weikel J, Möstl K, Revilla-Fernández S, Wodak E, Bagó Z, et al. Subclinical infection with avian influenza A (H5N1) virus in cats. *Emerg Infect Dis.* 2007;13:243–7. <https://doi.org/10.3201/eid1302.060608>
 44. Hu T, Zhao H, Zhang Y, Zhang W, Kong Q, Zhang Z, et al. Fatal influenza A (H5N1) virus infection in zoo-housed tigers in Yunnan Province, China. *Sci Rep.* 2016;6:25845. <https://doi.org/10.1038/srep25845>
 45. Songserm T, Amonsin A, Jam-on R, Sae-Heng N, Meemak N, Pariyothorn N, et al. Avian influenza H5N1 in naturally infected domestic cat. *Emerg Infect Dis.* 2006;12:681–3. <https://doi.org/10.3201/eid1204.051396>
 46. Songserm T, Amonsin A, Jam-on R, Sae-Heng N, Pariyothorn N, Payungporn S, et al. Fatal avian influenza A H5N1 in a dog. *Emerg Infect Dis.* 2006;12:1744–7. <https://doi.org/10.3201/eid1211.060542>
 47. Thorsson E, Zohari S, Roos A, Banihashem F, Bröjer C, Neimanis A. Highly pathogenic avian influenza A(H5N1) virus in a harbor porpoise, Sweden. *Emerg Infect Dis.* 2023;29:852–5. <https://doi.org/10.3201/eid2904.221426>
 48. Daszak P, Cunningham AA, Hyatt AD. Emerging infectious diseases of wildlife – threats to biodiversity and human health. *Science.* 2000;287:443–9. <https://doi.org/10.1126/science.287.5452.443>
 49. Taubenberger JK, Reid AH, Lourens RM, Wang R, Jin G, Fanning TG. Characterization of the 1918 influenza virus polymerase genes. *Nature.* 2005;437:889–93. <https://doi.org/10.1038/nature04230>
 50. Kuiken T, Cromie R. Protect wildlife from livestock diseases. *Science.* 2022;378:5.

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EID Podcast Rat Hepatitis E Virus in Norway Rats, Ontario, Canada, 2018–2021



Reports of acute hepatitis caused by rat hepatitis E virus (HEV) raise concerns regarding the potential risk for rat HEV transmission to people and hepatitis E as an emerging infectious disease worldwide. During 2018–2021, researchers tested liver samples from 372 Norway rats from southern Ontario, Canada to investigate presence of hepatitis E virus infection. Overall, 21 (5.6%) rats tested positive for the virus.

In this EID podcast, Dr. Sarah Robinson, a postdoctoral researcher at the University of Guelph, discusses hepatitis E virus in Norway rats in Ontario, Canada.

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Recent Changes in Patterns of Mammal Infection with Highly Pathogenic Avian Influenza A(H5N1) Virus Worldwide

Appendix

Appendix Table. Mammals infected by H5N1 or H5NX in previous infection waves and current panzootic, conservation status, order, family, diet consumed, number of individuals affected, mutations found and country of origin

Order	Family	Species	Conservation status**	Diet***	Individuals affected	Mutations found****	Country	References
PREVIOUS INFECTION WAVES (2003–2019)								
Terrestrial and semi-aquatic mammals								
Carnivora	Canidae	Raccoon dog (<i>Nyctereutes procyonoides</i>)	LC	O	At least 2	T/S156A, PB1-P13, PB1-Y436	China	(1)
	Canidae	Dog (<i>Canis lupus familiaris</i>)	NC	C	At least 1	PB2-E627K, M2-Ser31Asn, M2-Leu261e	Thailand	(2,3)
	Mustelidae	Mink (<i>Neovison vison</i>)	LC	C	At least 371	M2-S31N, NS1-G149A, NS1-D92E, HA-E513G	China, Sweden	(4–6)
	Mustelidae	Stone marten (<i>Martes foina</i>)	LC	O	At least 1	—	Germany	(7)
	Viverridae	Owston's civet (<i>Chrotogale owstoni</i>)	EN	O	At least 3	—	Vietnam	(8)
	Felidae	Tiger (<i>Panthera tigris</i>)	EN	C	At least 150	PB2-E627K, 31(N; Asparagine), HA-Asp94Asn, HA-Ser133Ala, HA-Thr156Ala, HA-Thr188Ile, HA-Lys189Arg, NA-His254Tyr, PB2-Asp701Asn, M1-Asn30Asp, M1-Thr215Ala, M2-Ser31Asn, NS1-Leu98Phe, NS1-Ile101Met, HA-Ser155Asn, HA-Gln222Leu, HA-Gly224Ser, PB2-Lys627Glu, NS1-Pro42Ser, NS1-Asp87Glu	Thailand, China	(9–13)
	Felidae	Lion (<i>Panthera leo</i>)	VU	C	At least 1	—	China	(14)

Order	Family	Species	Conservation status**	Diet***	Individuals affected	Mutations found****	Country	References
	Felidae	Leopard (<i>Panthera pardus</i>)	VU	C	At least 2	PB2-E627K	Thailand	(11)
	Felidae	Cat (<i>Felis catus</i>)	NC	C	At least 13	PB2-E627K, M2-Ser31Asn, M2-Leu261Ie	Austria, Thailand, Germany, Iraq	(2,15–18)
Lagomorpha	Ochotonidae	Plateau pikas (<i>Ochotona curzoniae</i>)	LC	H	At least 5	—	China	(19)
Artiodactyla	Suidae	Pigs (<i>Sus scrofa domestica</i>)	NC	O	At least 70	Ala134Ser-viral receptor	China, Nigeria, Indonesia	(20–22)
Perissodactyla	Equidae	Donkey (<i>Equus asinus</i>)	NC	H	At least 3	HA-98(Y to N), HA-193(K to R), HA-216(E to K), HA-221(P to S)	Egypt	(23)
CURRENT PANZOOTIC (2020–2023)								
Terrestrial and semi-aquatic mammals								
Carnivora	Canidae	Red fox (<i>Vulpes vulpes</i>)	LC	C/FS	At least 290	PB2-E627K, NP-G485R, PB2-A152T, PB2-T521I, PB1-M644V, NP-A336T, NA-L22S, NS-D209N, PB2-D701N, HA-S137A, HA-T160A, HA-A185E, HA-D195T, HA-V198I, HA-E268G, HA-V210A, PB2-V292I, PB1-D622G, PB1-N375S, PA-P28L, PA-A36V, HA-N101S, HA-V214A, NA-A395E, NA-S248N, NA-H155Y, NA-N366S, M1-N30D, M1-T215A, M2-S82N, NS1-V2261, NS1-P42S, NS1-I106M	Netherlands, Sweden, Finland, Estonia, Ireland, Norway, Belgium, UK, France, Italy, Japan, Germany, USA, Canada, Latvia, Denmark,	(24–46)
	Canidae	Arctic fox (<i>Vulpes lagopus</i>)	LC	C/FS	At least 2048	—	Finland	(27,43,45)
	Canidae	Grey fox (<i>Urocyon cinereoargenteus</i>)	LC	C/FS	At least 1	—	USA	(36)
	Canidae	Raccoon dog (<i>Nyctereutes procyonoides</i>)	LC	C	At least 5	—	Japan, Finland	(27,28,33,37,39–41,43–45)
	Canidae	South American bush dog (<i>Speothos venaticus venaticus</i>)	NT	C	At least 1	—	UK	(41,44,45)
	Canidae	Coyote (<i>Canis latrans</i>)	LC	C/FS	At least 1	—	USA	(24,27,28,30,36,37,40,41,44,45)
	Canidae	Dog (<i>Canis lupus familiaris</i>)	NC	C	At least 1	—	Canada	(27,41,44,45)

Order	Family	Species	Conservation status**	Diet***	Individuals affected	Mutations found****	Country	References
	Mustelidae	Otter (<i>Lutra lutra</i>)	NT	C	At least 7	PB2-E627K	Finland,	(25–29,31,37,39–41,44–46)
	Mustelidae	Polecat (<i>Mustela putorius</i>)	LC	C	At least 7	PB2-E627K, PB2-T271A	Netherlands, UK Netherlands, Slovenia, Belgium	(26–29,37–41,44,45)
	Mustelidae	Badger (<i>Meles meles</i>)	LC	O	At least 1	—	Netherlands	(28,29,37,38,40,41,44,45)
	Mustelidae	Fisher cat (<i>Pekania pennanti</i>)	LC	O/FS	At least 4	—	USA, Canada	(24,27,28,30,36,37,40,41,44,45)
	Mustelidae	Mink (<i>Neovison vison</i>)	LC	C	>50000	HA-A185E, HA-D195T, HA-V198I, HA-E268G, HA-V210A, HA-S137A, HA-T160, PB2-T271A, PB1-388R, PB1-317V, PB1-F2–30L, NA-74S, NS2-13G, PA-56T, NA-163L	Canada, Spain, Finland	(24,27,28,35,37,40,41,43–45,47)
	Mustelidae	Stone marten (<i>Martes foina</i>)	LC	O	At least 1	—	Netherlands	(41,44,45)
	Mustelidae	Pine marten (<i>Martes martes</i>)	LC	O/FS	At least 1	—	Germany	(27,45)
	Mustelidae	Marine otter (<i>Lontra felina</i>)	EN	C	At least 2	—	Chile	(27,41,44,45; Pardo Roa et al.†)
	Mustelidae	Huillin (<i>Lontra provocax</i>)*	EN	C	At least 1	—	Chile	(27,44,45)
	Mustelidae	American marten (<i>Martes americana</i>)	LC	O/FS	At least 1	—	USA	(27,30)
	Mustelidae	North American river otter (<i>Lontra canadensis</i>)	LC	C	At least 2	—	USA, Canada	(24,27,30,41,44,45)
	Procyonidae	Raccoon (<i>Procyon lotor</i>)	LC	O	At least 18	PB2-E627K	USA, Canada	(24,27,28,30,36,37,40,41,44,45)
	Procyonidae	South American coati (<i>Nasua nasua</i>)	LC	O	At least 19	—	Uruguay, Germany	(27,44,45)
	Felidae	Bobcat (<i>Lynx rufus</i>)	LC	C/FS	At least 6	—	USA	(24,27,28,30,36,37,40,41,44,45)
	Felidae	Lynx (<i>Lynx lynx</i>)	LC	C	At least 1	PB2-E627K, PB2-D701N	Finland	(26,28,37,39–41,44–46)
	Felidae	Tiger (<i>Panthera tigris</i>)	EN	C	At least 1	—	USA	(24,27,28,30,41,44,45)

Order	Family	Species	Conservation status**	Diet***	Individuals affected	Mutations found****	Country	References
	Felidae	Mountain lion (<i>Puma concolor</i>)	LC	C	At least 21	—	USA	(24,27,28,30,41,44,45)
	Felidae	Lion (<i>Panthera leo</i>)*	VU	C	At least 1	—	Peru	(27,45)
	Felidae	Caracal (<i>Caracal caracal</i>)	LC	C	At least 1	—	Poland	(44,45)
	Felidae	Cat (<i>Felis catus</i>)	NC	C	At least 35	PB2-E627K, NS2-E26G, PB2-K526R	USA, France, Canada, Korea (Republic of), Poland	(27,28,41,44,45,48–52)
	Felidae	Leopard (<i>Panthera pardus orientalis</i>)	CE	C	At least 1	---	USA	(24,27,28,30,40,41,44,45)
	Mephitidae	Skunks (<i>Mephitis mephitis</i>)	LC	O/FS	At least 90	HA-S137A, HA-T160A, HA-A185E, HA-D195T, HA-V198I, HA-E268G, HA-V210A	USA, Canada	(24,27,28,30,35–37,39–41,44,45)
	Ursidae	Asiatic black bear (<i>Ursus thibetanus</i>)	VU	O/FS	At least 1	—	France	(41,44,45)
	Ursidae	American black bear (<i>Ursus americanus</i>)	LC	O/FS	At least 7	PB2-D701N	USA, Canada	(24,27,28,30,37,40,41,44,45,53)
	Ursidae	Grizzly bear (<i>Ursus arctos</i>)	LC	O/FS	At least 4	—	USA	(24,27,28,30,40,41,44,45,54)
Didelphimorphia	Didelphidae	Virginia opossum (<i>Didelphis virginiana</i>)	LC	O/FS	At least 4	—	USA	(24,27,28,30,36,37,40,41,44,45)
Rodentia	Cricetidae	Muskrat (<i>Ondatra zibethicus</i>)	LC	O	At least 1	—	USA	(24)
	Castoridae	Beaver (<i>Castor canadensis</i>)	LC	H	At least 1	—	USA	(24)
Marine mammals Carnivora	Phocidae	Grey seal (<i>Halichoerus grypus</i>)	LC	C	At least 40	PA-A70V, PA-V379M, HA-P152S, NP-M448V/1, NP-D455N, NA-M231, NS1-R67Q, NS1-S87T, PB2-E627K	USA, UK, Canada, Poland, Netherlands, Germany	(24,27,28,30,31,37,40,41,44,45,55,56)
	Phocidae	Harbor seal (<i>Phoca vitulina</i>)	LC	C	At least 90	PB2-S12I, PB2-E627K, PB2-D701N, PB1-P135, PB1-R211K, PB1-M523L, PB1-V527I, PB1-1728V, PB1-R480K, PB1-N375T, PB1-F2-A56V	USA, Canada, UK, Denmark, Germany	(24,27,28,30,31,37,40,41,44,45,56)
	Phocidae	Caspian seal (<i>Pusa caspica</i>)	EN	C	—	—	Russia	(41,44,45)

Order	Family	Species	Conservation status**	Diet***	Individuals affected	Mutations found****	Country	References
	Phocidae	Southern elephant seal (<i>Mirounga leonina</i>)*	LC	C	At least 2	—	Argentina	(27,45)
	Otariidae	Northern fur seal (<i>Callorhinus ursinus</i>)	VU	C	At least 1	—	Russia	(27,45)
	Otariidae	South American fur seal (<i>Arctocephalus australis</i>)	LC	C	At least 35	—	Peru, Argentina, Uruguay, Brazil	(27,41,44,45)
	Otariidae	American sea lions (<i>Otaria flavescens</i>)	LC	C	>10000	PB2-D701N, PB1-S515A, PA-R57Q, PA-T85V, PA- M861, HA-H355R, NP- Y289F, NA-A81I, NS1- D26K, PB1-L378M, PB2- Q591K	Peru, Chile, Argentina, Uruguay, Brazil	(27,28,41,44,45,57- 59; Pardo Roa et al.†)
Cetacea	Phocoenidae	Burmeister's porpoise (<i>Phocoena spinipinnis</i>)	NT	C	At least 1	—	Chile	(41,44,45)
	Phocoenidae	Porpoise (<i>Phocoena phocoena</i>)	LC	C	At least 3	—	Sweden, UK, Canada	(24,28,31,37,40,41, 44,45,60)
	Delphinidae	Chilean dolphin (<i>Cephalorhynchus eutropia</i>)	NT	C	At least 1	—	Chile	(41,44,45)
	Delphinidae	White-sided dolphin (<i>Lagenorhynchus acutus</i>)	LC	C	At least 1	—	Canada	(24,28,40,41,44,45)
	Delphinidae	Common dolphin (<i>Delphinus delphis</i>)	LC	C	At least 3	PB1-L378M, PA-T85V, NA-A81I, NA-S339P, M1- N87T, NS1-D26K, NS1- E60V	Peru, UK,	(28,31,41,44,45,57)
	Delphinidae	Bottlenose dolphin (<i>Tursiops truncatus</i>)	LC	C	At least 2	NA-S246N, HA-T192I	USA, Peru	(24,27,28,30,37,40, 41,44,45; Murawski et al.‡)

* H5NX (H5 untipped).

**Conservation status based on The IUCN Red List of Threatened species: LC (Least concern); NT (Near Threatened); VU (Vulnerable); EN (Endangered); CR (Critically Endangered), NC (Non-classified).

*** Diets based on MammalBase-Database of recent mammals and the IUCN Red List of Threatened species: C (Carnivorous); FS (facultative scavenger); O (Omnivorous); H (Herbivorous).

**** To view the complete list and details of mutations found, refer to the cited bibliography.

†Pardo Roa et al., unpub. data, <https://doi.org/10.1101/2023.06.30.547205>.

‡Murawski et al., unpub data, <https://www.researchsquare.com/article/rs-3065313/latest>.

References

1. Qi X, Li X, Rider P, Fan W, Gu H, Xu L, et al. Molecular characterization of highly pathogenic H5N1 avian influenza A viruses isolated from raccoon dogs in China. PLoS One. 2009;4:e4682. [PubMed https://doi.org/10.1371/journal.pone.0004682](https://doi.org/10.1371/journal.pone.0004682)
2. Amonsin A, Songserm T, Chutinimitkul S, Jam-On R, Sae-Heng N, Pariyothorn N, et al. Genetic analysis of influenza A virus (H5N1) derived from domestic cat and dog in Thailand. Arch Virol. 2007;152:1925–33. [PubMed https://doi.org/10.1007/s00705-007-1010-5](https://doi.org/10.1007/s00705-007-1010-5)
3. Songserm T, Amonsin A, Jam-on R, Sae-Heng N, Pariyothorn N, Payungporn S, et al. Fatal avian influenza A H5N1 in a dog. Emerg Infect Dis. 2006;12:1744–7. [PubMed https://doi.org/10.3201/eid1211.060542](https://doi.org/10.3201/eid1211.060542)
4. Jiang W, Wang S, Zhang C, Li J, Hou G, Peng C, et al. Characterization of H5N1 highly pathogenic mink influenza viruses in eastern China. Vet Microbiol. 2017;201:225–30. [PubMed https://doi.org/10.1016/j.vetmic.2017.01.028](https://doi.org/10.1016/j.vetmic.2017.01.028)
5. Reperant LA, Rimmelzwaan GF, Kuiken T. Avian influenza viruses in mammals. Rev Sci Tech. 2009;28:137–59. [PubMed https://doi.org/10.20506/rst.28.1.1876](https://doi.org/10.20506/rst.28.1.1876)
6. Kiss I, Gyarmati P, Zohari S, Ramsay KW, Metreveli G, Weiss E, et al. Molecular characterization of highly pathogenic H5N1 avian influenza viruses isolated in Sweden in 2006. Virol J. 2008;5:113. [PubMed https://doi.org/10.1186/1743-422X-5-113](https://doi.org/10.1186/1743-422X-5-113)
7. Klopfleisch R, Wolf PU, Wolf C, Harder T, Starick E, Niebuhr M, et al. Encephalitis in a stone marten (*Martes foina*) after natural infection with highly pathogenic avian influenza virus subtype H5N1. J Comp Pathol. 2007;137:155–9. [PubMed https://doi.org/10.1016/j.jcpa.2007.06.001](https://doi.org/10.1016/j.jcpa.2007.06.001)
8. Robertson SI, Bell DJ, Smith GJD, Nicholls JM, Chan KH, Nguyen DT, et al. Avian influenza H5N1 in viverrids: implications for wildlife health and conservation. Proc Biol Sci. 2006;273:1729–32. [PubMed https://doi.org/10.1098/rspb.2006.3549](https://doi.org/10.1098/rspb.2006.3549)
9. Amonsin A, Payungporn S, Theamboonlers A, Thanawongnuwech R, Suradhat S, Pariyothorn N, et al. Genetic characterization of H5N1 influenza A viruses isolated from zoo tigers in Thailand. Virology. 2006;344:480–91. [PubMed https://doi.org/10.1016/j.virol.2005.08.032](https://doi.org/10.1016/j.virol.2005.08.032)
10. Hu T, Zhao H, Zhang Y, Zhang W, Kong Q, Zhang Z, et al. Fatal influenza A (H5N1) virus infection in zoo-housed tigers in Yunnan Province, China. Sci Rep. 2016;6:25845. [PubMed https://doi.org/10.1038/srep25845](https://doi.org/10.1038/srep25845)

11. Keawcharoen J, Oraveerakul K, Kuiken T, Fouchier RA, Amonsin A, Payungporn S, et al. Avian influenza H5N1 in tigers and leopards. *Emerg Infect Dis*. 2004;10:2189–91. [PubMed https://doi.org/10.3201/eid1012.040759](https://doi.org/10.3201/eid1012.040759)
12. He S, Shi J, Qi X, Huang G, Chen H, Lu C. Lethal infection by a novel reassortant H5N1 avian influenza A virus in a zoo-housed tiger. *Microbes Infect*. 2015;17:54–61. [PubMed https://doi.org/10.1016/j.micinf.2014.10.004](https://doi.org/10.1016/j.micinf.2014.10.004)
13. Thanawongnuwech R, Amonsin A, Tantilertcharoen R, Damrongwatanapokin S, Theamboonlers A, Payungporn S, et al. Probable tiger-to-tiger transmission of avian influenza H5N1. *Emerg Infect Dis*. 2005;11:699–701. [PubMed https://doi.org/10.3201/eid1105.050007](https://doi.org/10.3201/eid1105.050007)
14. Chen Q, Wang H, Zhao L, Ma L, Wang R, Lei Y, et al. First documented case of avian influenza (H5N1) virus infection in a lion. *Emerg Microbes Infect*. 2016;5:e125. [PubMed https://doi.org/10.1038/emi.2016.127](https://doi.org/10.1038/emi.2016.127)
15. Leschnik M, Weikel J, Möstl K, Revilla-Fernández S, Wodak E, Bagó Z, et al. Subclinical infection with avian influenza A (H5N1) virus in cats. *Emerg Infect Dis*. 2007;13:243–7. [PubMed https://doi.org/10.3201/eid1302.060608](https://doi.org/10.3201/eid1302.060608)
16. Songserm T, Amonsin A, Jam-on R, Sae-Heng N, Meemak N, Pariyothorn N, et al. Avian influenza H5N1 in naturally infected domestic cat. *Emerg Infect Dis*. 2006;12:681–3. [PubMed https://doi.org/10.3201/eid1204.051396](https://doi.org/10.3201/eid1204.051396)
17. Klopfleisch R, Wolf PU, Uhl W, Gerst S, Harder T, Starick E, et al. Distribution of lesions and antigen of highly pathogenic avian influenza virus A/Swan/Germany/R65/06 (H5N1) in domestic cats after presumptive infection by wild birds. *Vet Pathol*. 2007;44:261–8. [PubMed https://doi.org/10.1354/vp.44-3-261](https://doi.org/10.1354/vp.44-3-261)
18. Yingst SL, Saad MD, Felt SA. Qinghai-like H5N1 from domestic cats, northern Iraq. *Emerg Infect Dis*. 2006;12:1295–7. [PubMed https://doi.org/10.3201/eid1708.060264](https://doi.org/10.3201/eid1708.060264)
19. Zhou J, Sun W, Wang J, Guo J, Yin W, Wu N, et al. Characterization of the H5N1 highly pathogenic avian influenza virus derived from wild pikas in China. *J Virol*. 2009;83:8957–64. [PubMed https://doi.org/10.1128/JVI.00793-09](https://doi.org/10.1128/JVI.00793-09)
20. He L, Zhao G, Zhong L, Liu Q, Duan Z, Gu M, et al. Isolation and characterization of two H5N1 influenza viruses from swine in Jiangsu Province of China. *Arch Virol*. 2013;158:2531–41. [PubMed https://doi.org/10.1007/s00705-013-1771-y](https://doi.org/10.1007/s00705-013-1771-y)
21. Meseko C, Globig A, Ijomanta J, Joannis T, Nwosuh C, Shamaki D, et al. Evidence of exposure of domestic pigs to highly pathogenic avian influenza H5N1 in Nigeria. *Sci Rep*. 2018;8:5900. [PubMed https://doi.org/10.1038/s41598-018-24371-6](https://doi.org/10.1038/s41598-018-24371-6)

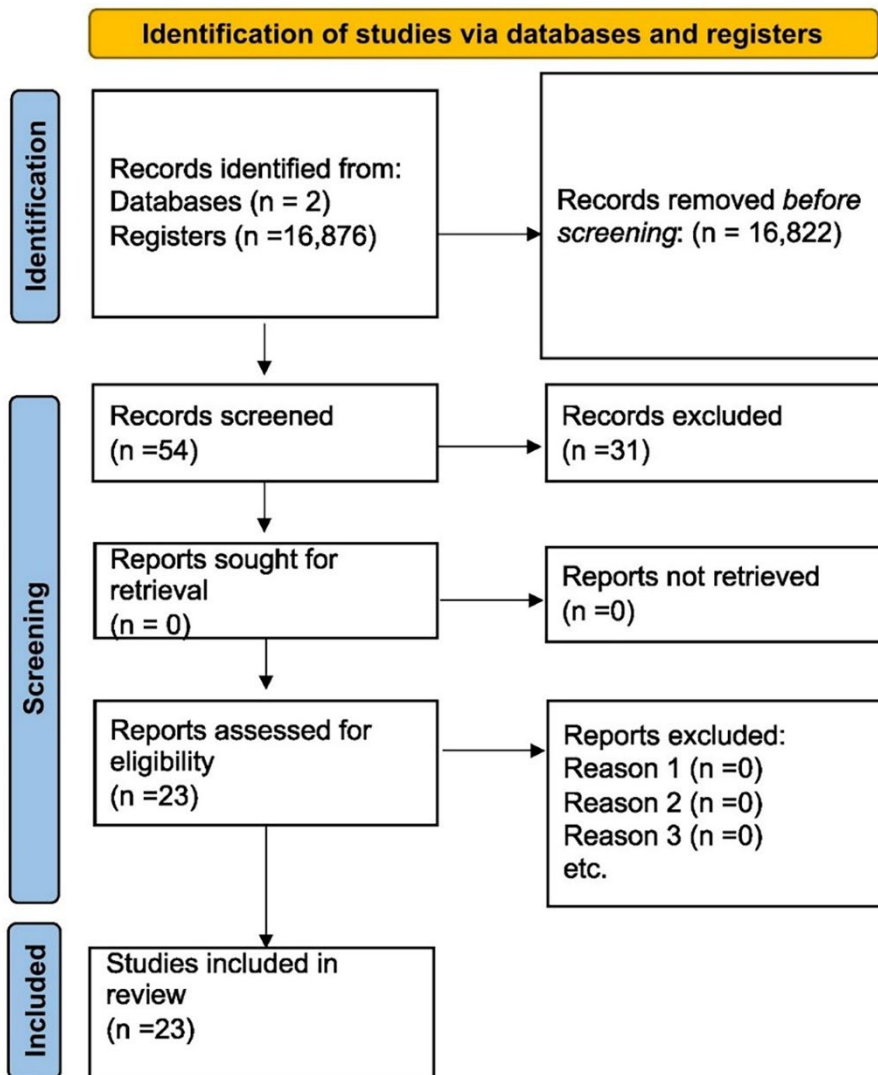
22. Nidom CA, Takano R, Yamada S, Sakai-Tagawa Y, Daulay S, Aswadi D, et al. Influenza A (H5N1) viruses from pigs, Indonesia. *Emerg Infect Dis*. 2010;16:1515–23. [PubMed](#)
<https://doi.org/10.3201/eid1610.100508>
23. Abdel-Moneim AS, Abdel-Ghany AE, Shany SA. Isolation and characterization of highly pathogenic avian influenza virus subtype H5N1 from donkeys. *J Biomed Sci*. 2010;17:25. [PubMed](#) <https://doi.org/10.1186/1423-0127-17-25>
24. Harvey JA, Mullinax JM, Runge MC, Prosser DJ. The changing dynamics of highly pathogenic avian influenza H5N1: next steps for management and science in North America. *Biol Conserv*. 2023;282:110041.
<https://doi.org/10.1016/j.biocon.2023.110041>
25. Adlhoch C, Fusaro A, Gonzales JL, Kuiken T, Marangon S, Niqueux É, et al.; European Food Safety Authority, European Centre for Disease Prevention, Control, European Union Reference Laboratory for Avian Influenza. Avian influenza overview September–December 2021. *EFSA J*. 2021;19:e07108. [PubMed](#)
26. Adlhoch C, Fusaro A, Gonzales JL, Kuiken T, Marangon S, Niqueux É, et al.; European Food Safety Authority; European Centre for Disease Prevention and Control; European Union Reference Laboratory for Avian Influenza. Avian influenza overview December 2021–March 2022. *EFSA J*. 2022;20:e07289. [PubMed](#)
27. World Organization for Animal Health. WAHIS: World Animal Health Information System. 2023 [cited 2023 Oct 30]. <https://wahis.woah.org/#/home>
28. Adlhoch C, Fusaro A, Gonzales JL, Kuiken T, Marangon S, Mirinaviciute G, et al.; European Food Safety Authority; European Centre for Disease Prevention and Control; European Union Reference Laboratory for Avian Influenza. Avian influenza overview December 2022–March 2023. *EFSA J*. 2023;21:e07917. [PubMed](#)
29. Vreman S, Kik M, Germeraad E, Heutink R, Harders F, Spierenburg M, et al. Zoonotic mutation of highly pathogenic avian influenza H5N1 virus identified in the brain of multiple wild carnivore species. *Pathogens*. 2023;12:168. [PubMed](#)
<https://doi.org/10.3390/pathogens12020168>
30. US Department of Agriculture. 2022–2023 Detections of highly pathogenic avian influenza in mammals [cited 2023 Oct 30].
<https://www.aphis.usda.gov/aphis/ourfocus/animalhealth/animal-disease-information/avian/avian-influenza/hpai-2022/2022-hpai-mammals>
31. United Kingdom Animal and Plant Health Agency. Confirmed findings of influenza of avian origin in non-avian wildlife [cited 2023 Oct 30].

- <https://www.gov.uk/government/publications/bird-flu-avian-influenza-findings-in-non-avian-wildlife/confirmed-findings-of-influenza-of-avian-origin-in-non-avian-wildlife>
32. Bordes L, Vreman S, Heutink R, Roose M, Venema S, Pritz-Verschuren SBE, et al. Highly pathogenic avian influenza H5N1 virus infections in wild red foxes (*Vulpes vulpes*) show neurotropism and adaptive virus mutations. *Microbiol Spectr*. 2023;11:e0286722. [PubMed https://doi.org/10.1128/spectrum.02867-22](https://doi.org/10.1128/spectrum.02867-22)
 33. Hiono T, Kobayashi D, Kobayashi A, Suzuki T, Satake Y, Harada R, et al. Virological, pathological, and glycovirological investigations of an Ezo red fox and a tanuki naturally infected with H5N1 high pathogenicity avian influenza viruses in Hokkaido, Japan. *Virology*. 2023;578:35–44. [PubMed https://doi.org/10.1016/j.virol.2022.11.008](https://doi.org/10.1016/j.virol.2022.11.008)
 34. Rijks JM, Hesselink H, Lollinga P, Wesselman R, Prins P, Weesendorp E, et al. Highly pathogenic avian influenza A (H5N1) virus in wild red foxes, the Netherlands, 2021. *Emerg Infect Dis*. 2021;27:2960–2. [PubMed https://doi.org/10.3201/eid2711.211281](https://doi.org/10.3201/eid2711.211281)
 35. Alkie TN, Cox S, Embury-Hyatt C, Stevens B, Pople N, Pybus MJ, et al. Characterization of neurotropic HPAI H5N1 viruses with novel genome constellations and mammalian adaptive mutations in free-living mesocarnivores in Canada. *Emerg Microbes Infect*. 2023;12:2186608. [PubMed https://doi.org/10.1080/22221751.2023.2186608](https://doi.org/10.1080/22221751.2023.2186608)
 36. Elsmo EJ, Wunschmann A, Beckmen KB, Broughton-Neiswanger LB, Buckles EL, Ellis J, et al. Pathology of natural infection with highly pathogenic avian influenza virus (H5N1) clade 2.3. 4.4 b in wild terrestrial mammals in the United States in 2022. *Emerg Infect Dis*. 2023;29:2451–60. [PubMed https://doi.org/10.3201/eid2912.230464](https://doi.org/10.3201/eid2912.230464)
 37. Adlhoch C, Fusaro A, Gonzales JL, Kuiken T, Marangon S, Niqueux É, et al.; European Food Safety Authority; European Centre for Disease Prevention and Control; European Union Reference Laboratory for Avian Influenza. Avian influenza overview June–September 2022. *EFSA J*. 2022;20:e07597. [PubMed https://doi.org/10.2903/j.efsa.2022.6511](https://doi.org/10.2903/j.efsa.2022.6511)
 38. Chestakova IV, van der Linden A, Bellido Martin B, Caliendo V, Vuong O, Thewessen S, et al. High number of HPAI H5 virus infections and antibodies in wild carnivores in the Netherlands, 2020–2022. *Emerg Microbes Infect*. 2023;12:2270068. [PubMed https://doi.org/10.1080/22221751.2023.2270068](https://doi.org/10.1080/22221751.2023.2270068)
 39. Adlhoch C, Fusaro A, Gonzales JL, Kuiken T, Marangon S, Niqueux É, et al.; European Food Safety Authority; European Centre for Disease Prevention and Control; European Union Reference Laboratory for Avian Influenza. Avian influenza overview March–June 2022. *EFSA J*. 2022;20:e07415. [PubMed https://doi.org/10.2903/j.efsa.2022.6415](https://doi.org/10.2903/j.efsa.2022.6415)

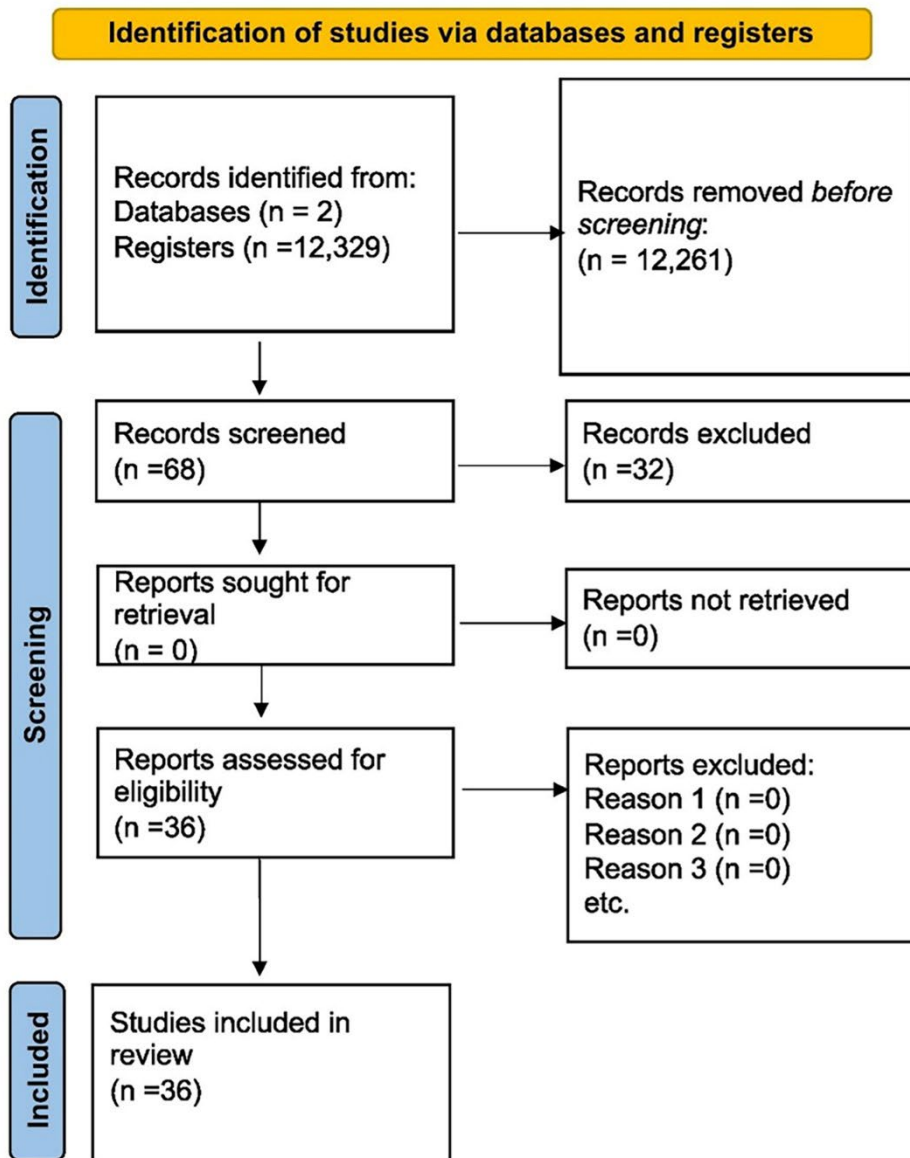
40. Adlhoch C, Fusaro A, Gonzales JL, Kuiken T, Marangon S, Niqueux É, et al.; European Food Safety Authority, European Centre for Disease Prevention and Control, European Union Reference Laboratory for Avian Influenza. Avian influenza overview September–December 2022. *EFSA J.* 2023;21:e07786. [PubMed](#)
41. Adlhoch C, Fusaro A, Gonzales JL, Kuiken T, Mirinavičiūtė G, Niqueux É, et al.; European Food Safety Authority, European Centre for Disease Prevention and Control, European Union Reference Laboratory for Avian Influenza. Avian influenza overview March–April 2023. *EFSA J.* 2023;21:e08039. [PubMed](#)
42. Cronk BD, Caserta LC, Laverack M, Gerdes RS, Hynes K, Hopf CR, et al. Infection and tissue distribution of highly pathogenic avian influenza A type H5N1 (clade 2.3.4.4b) in red fox kits (*Vulpes vulpes*). *Emerg Microbes Infect.* 2023;12:2249554. [PubMed](#)
<https://doi.org/10.1080/22221751.2023.2249554>
43. Lindh E, Lounela H, Ikonen N, Kantala T, Savolainen-Kopra C, Kauppinen A, et al. Highly pathogenic avian influenza A(H5N1) virus infection on multiple fur farms in the South and Central Ostrobothnia regions of Finland, July 2023. *Euro Surveill.* 2023;28:2300400. [PubMed](#) <https://doi.org/10.2807/1560-7917.ES.2023.28.31.2300400>
44. Adlhoch C, Fusaro A, Gonzales JL, Kuiken T, Melidou A, Mirinavičiūtė G, et al.; European Food Safety Authority, European Centre for Disease Prevention and Control, European Union Reference Laboratory for Avian Influenza. Avian influenza overview April–June 2023. *EFSA J.* 2023;21:e08191. [PubMed](#)
45. Adlhoch C, Fusaro A, Gonzales JL, Kuiken T, Mirinavičiūtė G, Niqueux É, et al.; European Food Safety Authority; European Centre for Disease Prevention and Control; European Union Reference Laboratory for Avian Influenza. Avian influenza overview June–September 2023. *EFSA J.* 2023;21:e08328. [PubMed](#)
46. Tammiranta N, Isomursu M, Fusaro A, Nylund M, Nokireki T, Giussani E, et al. Highly pathogenic avian influenza A (H5N1) virus infections in wild carnivores connected to mass mortalities of pheasants in Finland. *Infect Genet Evol.* 2023;111:105423. [PubMed](#)
<https://doi.org/10.1016/j.meegid.2023.105423>
47. Agüero M, Monne I, Sánchez A, Zecchin B, Fusaro A, Ruano MJ, et al. Highly pathogenic avian influenza A(H5N1) virus infection in farmed minks, Spain, October 2022. *Euro Surveill.* 2023;28:2300001. [PubMed](#) <https://doi.org/10.2807/1560-7917.ES.2023.28.3.2300001>

48. Rabalski L, Milewska A, Pohlmann A, Gackowska K, Lepionka T, Szczepaniak K, et al. Emergence and potential transmission route of avian influenza A (H5N1) virus in domestic cats in Poland, June 2023. *Euro Surveill.* 2023;28:2300390. [PubMed](#) <https://doi.org/10.2807/1560-7917.ES.2023.28.31.2300390>
49. Briand FX, Souchaud F, Pierre I, Beven V, Hirchaud E, Hérault F, et al. Highly pathogenic avian influenza A(H5N1) clade 2.3.4.4b virus in domestic Cat, France, 2022. *Emerg Infect Dis.* 2023;29:1696–8. [PubMed](#) <https://doi.org/10.3201/eid2908.230188>
50. Domańska-Blicharz K, Świętoń E, Świątalska A, Monne I, Fusaro A, Tarasiuk K, et al. Outbreak of highly pathogenic avian influenza A(H5N1) clade 2.3.4.4b virus in cats, Poland, June to July 2023. *Euro Surveill.* 2023;28:2300366. [PubMed](#) <https://doi.org/10.2807/1560-7917.ES.2023.28.31.2300366>
51. Szaluś-Jordanow O, Golke A, Dzieciatkowski T, Chrobak-Chmiel D, Rzewuska M, Czopowicz M, et al. A Fatal A/H5N1 avian influenza virus infection in a cat in Poland. *Microorganisms.* 2023;11:2263. [PubMed](#) <https://doi.org/10.3390/microorganisms11092263>
52. Sillman SJ, Drozd M, Loy D, Harris SP. Naturally occurring highly pathogenic avian influenza virus H5N1 clade 2.3.4.4b infection in three domestic cats in North America during 2023. *J Comp Pathol.* 2023;205:17–23. [PubMed](#) <https://doi.org/10.1016/j.jcpa.2023.07.001>
53. Jakobek BT, Berhane Y, Nadeau MS, Embury-Hyatt C, Lung O, Xu W, et al. Influenza A(H5N1) virus infections in 2 free-ranging black bears (*Ursus americanus*), Quebec, Canada. *Emerg Infect Dis.* 2023;29:2145–9. [PubMed](#) <https://doi.org/10.3201/eid2910.230548>
54. Kupferschmidt K. Bird flu spread between mink is a ‘warning bell’. *Science.* 2023;379:316–7. [PubMed](#) <https://doi.org/10.1126/science.adg8342>
55. Mirolo M, Pohlmann A, Ahrens AK, Kühl B, Rubio-Garcia A, Kramer K, et al. Highly pathogenic avian influenza A virus (HPAIV) H5N1 infection in two European grey seals (*Halichoerus grypus*) with encephalitis. *Emerg Microbes Infect.* 2023;12:e2257810. [PubMed](#) <https://doi.org/10.1080/22221751.2023.2257810>
56. Puryear W, Sawatzki K, Hill N, Foss A, Stone JJ, Doughty L, et al. Highly pathogenic avian influenza A(H5N1) virus outbreak in New England seals, United States. *Emerg Infect Dis.* 2023;29:786–91. [PubMed](#) <https://doi.org/10.3201/eid2904.221538>

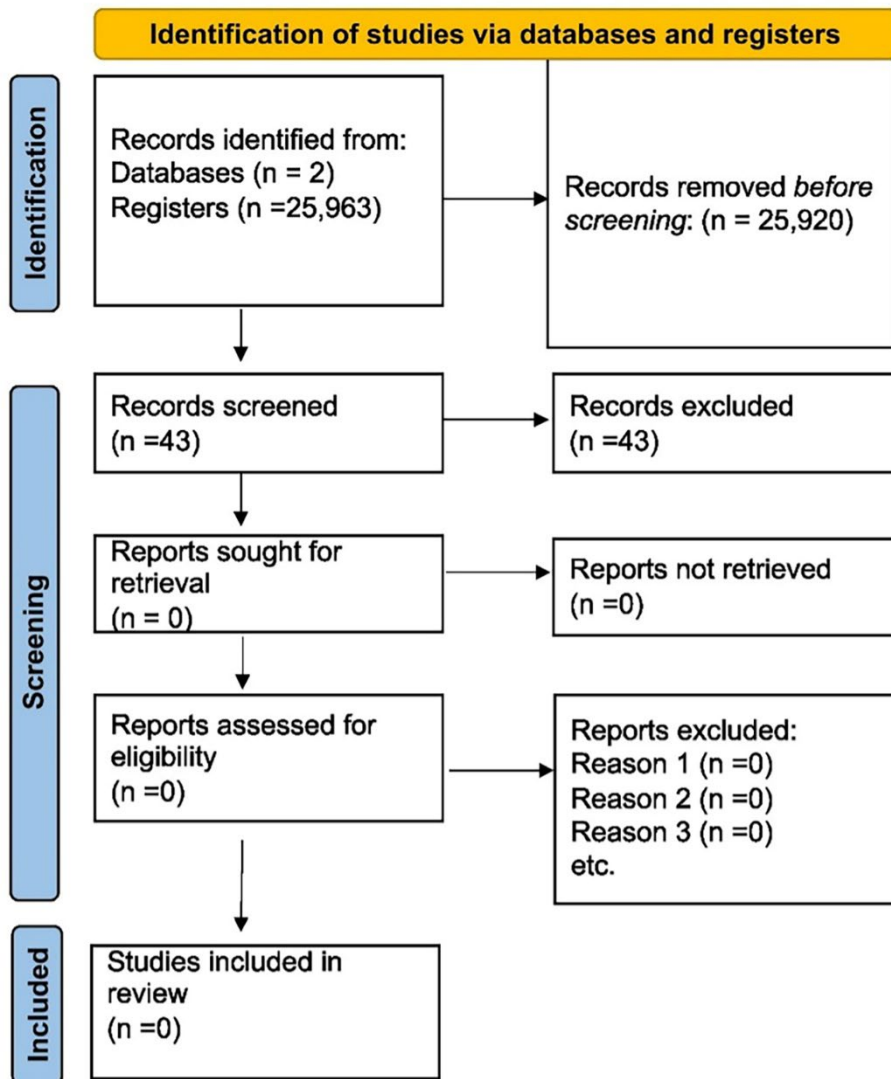
57. Leguia M, Garcia-Glaessner A, Muñoz-Saavedra B, Juarez D, Barrera P, Calvo-Mac C, et al. Highly pathogenic avian influenza A (H5N1) in marine mammals and seabirds in Peru. *Nat Commun.* 2023;14:5489. [PubMed https://doi.org/10.1038/s41467-023-41182-0](https://doi.org/10.1038/s41467-023-41182-0)
58. Gamarra-Toledo V, Plaza PI, Gutiérrez R, Inga-Diaz G, Saravia-Guevara P, Pereyra-Meza O, et al. Mass mortality of marine mammals caused by highly pathogenic avian influenza A(H5N1) virus. *Emerg Infect Dis.* 2023;29:2553–6. [PubMed https://doi.org/10.3201/eid2912.230192](https://doi.org/10.3201/eid2912.230192)
59. Ulloa M, Fernández A, Ariyama N, Colom-Rivero A, Rivera C, Nuñez P, et al. Mass mortality event in South American sea lions (*Otaria flavescens*) correlated to highly pathogenic avian influenza (HPAI) H5N1 outbreak in Chile. *Vet Q.* 2023;43:1–10. [PubMed https://doi.org/10.1080/01652176.2023.2265173](https://doi.org/10.1080/01652176.2023.2265173)
60. Thorsson E, Zohari S, Roos A, Banihashem F, Bröjer C, Neimanis A. Highly pathogenic avian influenza A(H5N1) virus in a harbor porpoise, Sweden. *Emerg Infect Dis.* 2023;29:852–5. [PubMed https://doi.org/10.3201/eid2904.221426](https://doi.org/10.3201/eid2904.221426)



Appendix Figure 1. Prisma flow diagram with the keywords “H5N1” and “Mammals” up to 2019 (Previous waves of H5N1 infection).



Appendix Figure 2. Prisma flow diagram with the keywords “H5N1” and “Mammals” 2020–2023 (Current panzootic 2020–2023).



Appendix Figure 3. Prisma flow diagram with the keywords “H5N1” OR HPAI OR Highly Pathogenic Avian Influenza AND “Mammals” OR Unusual Host.