

Novel Retinal Lesion in Ebola Survivors, Sierra Leone, 2016

Paul J. Steptoe, Janet T. Scott, Julia M. Baxter, Craig K. Parkes, Rahul Dwivedi, Gabriela Czanner, Matthew J. Vandy, Fayiah Momorie, Alimamy D. Fornah, Patrick Komba, Jade Richards, Foday Sahr, Nicholas A.V. Beare, Malcolm G. Semple

We conducted a case-control study in Freetown, Sierra Leone, to investigate ocular signs in Ebola virus disease (EVD) survivors. A total of 82 EVD survivors with ocular symptoms and 105 controls from asymptomatic civilian and military personnel and symptomatic eye clinic attendees underwent ophthalmic examination, including widefield retinal imaging. Snellen visual acuity was $\leq 6/7.5$ in 75.6% (97.5% CI 63%–85.7%) of EVD survivors and 75.5% (97.5% CI 59.1%–87.9%) of controls. Unilateral white cataracts were present in 7.4% (97.5% CI 2.4%–16.7%) of EVD survivors and no controls. Aqueous humor from 2 EVD survivors with cataract but no anterior chamber inflammation were PCR-negative for Zaire Ebola virus, permitting cataract surgery. A novel retinal lesion following the anatomic distribution of the optic nerve axons occurred in 14.6% (97.5% CI 7.1%–25.6%) of EVD survivors and no controls, suggesting neuronal transmission as a route of ocular entry.

The most recent Ebola virus disease (EVD) outbreak in West Africa is the largest outbreak in history. As of March 27, 2016, an estimated 3,956 persons in Sierra Leone had died from EVD, and 10,168 had survived (1). The scale of this epidemic has enabled the study of large numbers of survivors, facilitating the characterization of post-Ebola syndrome. Ocular symptoms have been reported, with incidence among survivors ranging from 14% to 60% (2–4). Evidence of acute uveitis on ophthalmic examination ranges from 18% to 58% (4–7). Classification of uveitis also varies and has been reported as 36%–62% anterior, 3% intermediate, 26%–36% posterior, and 18%–25% panuveitis (4,8). However, little is known regarding the

Author affiliations: University of Liverpool, Liverpool, UK (P.J. Steptoe, J.T. Scott, G. Czanner, N.A.V. Beare, M.G. Semple); Royal Liverpool Hospital, Liverpool (P.J. Steptoe, J.M. Baxter, C.K. Parkes, R. Dwivedi, N.A.V. Beare); National Institute for Health Research Health Protection Research Unit in Emerging and Zoonotic Infections, Liverpool (J.T. Scott, M.G. Semple); Connaught Hospital, Freetown, Sierra Leone (M.J. Vandy); 34th Military Hospital, Freetown (F. Momorie, A.D. Fornah, P. Komba, F. Sahr); Public Health England Laboratory, Makeni, Sierra Leone (J. Richards)

DOI: <https://dx.doi.org/10.3201/eid2307.161608>

medium- to long-term visual outcome of survivors or the rates of background uveitis and chorioretinal lesions within the local population.

Two published cases (9–11) and 2 case series (7,12) included fundus imaging, which attribute a range of retinal lesions to Ebola uveitis. Fourteen weeks after EVD discharge, a unilateral anterior hypertensive uveitis developed in 1 survivor and soon progressed into an aggressive anterior scleritis and intermediate uveitis. Viable Zaire Ebola virus (EBOV) was detected from the aqueous humor 9 weeks after the clearance of viremia (9). The duration of EBOV ocular persistence remains unknown, although repeated aqueous humor testing in the same survivor was negative for EBOV by quantitative reverse transcription PCR (qRT-PCR) 1 year later (10). Recurrences up to 13 months after EVD discharge have been reported, but confirmation of Ebola etiology through aqueous humor analysis was not conducted (7). Because of the unknown prevalence and duration of EBOV persistence in aqueous humor, survivors' access to cataract surgery is still restricted. Our study aimed to detect if any specific retinal signs can be attributed to past EVD in survivors, to describe the implications for visual acuity, and to assess for EBOV persistence in survivors with cataracts amenable to cataract surgery where no intraocular inflammation was present.

Methods

Study Design

We conducted a case-control prospective study comparing ophthalmic findings between EVD survivors and a control group during January–June 2016. Reporting of the findings is in accordance with guidelines set forth in the Strengthening the Reporting of Observational Studies in Epidemiology (STROBE) statement (13).

Study Population

We searched a database of EVD survivors from the 2014–2016 EVD epidemic who had attended the EVD survivors clinic at 34th Regiment Military Hospital in Freetown, Sierra Leone, for patients who had reported ophthalmic complaints at any of their follow-up appointments (2). Patients

were contacted by telephone and invited to attend the ophthalmology clinic for review. EVD survivors from other medical facilities in the region who had reported ophthalmic complaints also attended the clinic through word of mouth and electronic social media networking from other survivors. EVD survivor status was verified by the possession of a valid discharge certificate from an Ebola treatment center. Date of acute admission, date of discharge, and location of the Ebola treatment center were recorded from each discharge certificate.

Controls were recruited from ophthalmically symptomatic and asymptomatic local military personnel, their local family members, and symptomatic civilians. Survivors and controls were invited to participate in English or Krio, as preferred, with local ophthalmic nurses acting as interpreters. Consent was confirmed by fingerprint or signature.

Ocular Examination

Data were collected on first visit. The onset and nature of ocular complaint, and any systemic complaints were recorded on a standardized form before examination. Patients underwent visual acuity testing with either Snellen or Illiterate E-chart acuity methods. Snellen visual acuity was grouped into visual acuity ranges according to the International Classification of Diseases, Ninth Revision, Clinical Modification, and reported as patient's best eye vision.

Ocular anterior chamber assessment was conducted with a table-mounted slit lamp by 3 local ophthalmic clinical officers. The initial 35% of anterior chamber examinations were supervised and verified by an ophthalmologist from the United Kingdom. Patient examinations thereafter were conducted by local clinical officers alone with a telecommunication link for advice if required. Assessment of anterior chamber inflammation was graded according to the Standardization of Uveitis Nomenclature criteria (14). Intraocular pressures were measured by automated pneumatic tonometry (Canon TX-F; Melville, NY, USA); if out of reference range, this measure was repeated by using Goldmann applanation tonometry.

Widefield retinal images were obtained from patients with the use of a nonmydriatic Daytona Scanning Laser Ophthalmoscope (fundus camera; Optos, Dunfermline, UK). Optical coherence tomography was undertaken with the use of a Topcon DRI Triton swept source optical coherence tomography (Topcon Corporation, Tokyo, Japan). Posterior subcapsular and cortical cataract were graded from a comparison of standard images used in the Lens Opacities Classification System III (15) and applied to acquired fundus images. White cataracts were identified during patient examination, and fundus imaging was not possible. Presence of signs in the vitreous indicative of intermediate uveitis were also recorded from scanning laser ophthalmoscope imaging.

All clinical and artifactual signs present on scanning laser ophthalmoscopic imaging and corresponding autofluorescent imaging were recorded, grouped, and incorporated into an original classification form with associated standard images and descriptions (online Technical Appendix 1, <https://wwwnc.cdc.gov/EID/article/23/7/16-1608-Techapp1.pdf>). All images were graded for these features by 2 independent, masked ophthalmologists from the United Kingdom with specialist interests in medical retina. Certainty of positive findings were quantified as "yes, definitely," defined as >90% certainty, or "yes, questionably," defined as >50% certainty. Mutual agreements of definite or probable certainty were counted. Where discordance existed between findings, a third independent consultant ophthalmologist made final arbitration.

Paracentesis of the anterior chamber was performed at a slit lamp with a sterile 30-gauge needle while the clinician was wearing personal protective equipment. After informed consent was obtained, the procedure was conducted on 2 patients with white cataracts but no clinical signs of anterior chamber inflammation. At the time of sampling, the 2 survivors were 430 and 482 days postdischarge from their respective Ebola treatment centers. By using an anterior chamber tap procedure protocol (online Technical Appendix 2, <https://wwwnc.cdc.gov/EID/article/23/7/16-1608-Techapp2.pdf>), 0.1 mL of aqueous humor was obtained in both cases. Both specimens were delivered to the Public Health England laboratory (Makeni, Sierra Leone) for analysis for EBOV RNA on qRT-PCR assay. Testing was performed with the use of the standard institutional operating protocols by clinical laboratory technologists who were trained in the safe handling of infectious pathogens.

Statistical Methods

We reported results per patient and grouped by subject by using IBM SPSS version 22 (<http://www-01.ibm.com/support/docview.wss?uid=swg27038407>). Where data were missing, we reduced the denominator for each variable. We double-checked 10% of data entry and found 0% transcription errors. We calculated 97.5% CIs by using the exact binomial (Clopper-Pearson) method (16); no overlap between CIs indicates a statistically significant result. Fisher exact statistical value was calculated for significant results.

The study was approved by the Sierra Leone Ethics and Scientific Review Committee on January 29, 2016. In addition, the study was authorized by the Pharmacy Board of Sierra Leone.

Results

The numbers of patients recruited and examined at 34th Regiment Military Hospital were 82 EVD survivors (161 eyes; 2 missing retina images and 1 prosthetic eye) and 105 never-infected controls (208 eyes; 2 missing retinal images).

RESEARCH

Male-to-female ratio was 1:1.48 of EVD survivors and 1:0.64 of controls. Median age at time of ophthalmic examination was 28 years (interquartile range [IQR] 22–38 years) for EVD survivors and 41 years (IQR 30–48 years) for controls. Median time from Ebola treatment unit discharge to ophthalmic examination for survivors was 411 days (n = 70) (IQR 368–470 days). Ophthalmic examination findings were summarized for survivors and controls (Table).

We subclassified pigmented and nonpigmented retinal lesions into 10 discrete groups (online Technical Appendix 1) and noted frequency of each lesion type (Figure 1). We found no occurrences of the retinal lesion documented in a previous case report (9) in this EVD survivor cohort. Only the type 6 subcategory of retinal lesion was observed exclusively in EVD survivors, occurring in 12/82 (14.6% [97.5% CI 7.1%–25.6%]) EVD survivors and 0/105 controls (0%

Table. Ophthalmic examination findings in a case–control study of ocular signs in Ebola virus disease survivors, Sierra Leone, 2016*

Finding	Survivors		Controls	
	No.	% (97.5% CI)†	No.	% (97.5% CI)†
Best eye visual acuity‡				
Missing data	3	–	56	–
Normal	59	74.7 (62.1–84.9)	37	75.5 (59.1–87.9)
Near normal	18	22.8 (13.1–35.1)	8	16.3 (6.4–31.6)
Moderate	1	1.3 (0–7.8)	3	6.1 (1–18.6)
Severe	1	1.3 (0–7.8)	0	0 (0–8.6)
Profound	0	0 (0–5.5)	1	2 (0–12.3)
Near total	0	0 (0–5.5)	0	0 (0–8.6)
Total	0	0 (0–5.5)	0	0 (0–8.6)
Intraocular pressure, mmHg				
Missing data	35	–	74	–
Hypotonous (≤ 5)	5	10.6 (3–25)	0	0 (0–13.2)
Reduced (6–10)	5	10.6 (3–25)	3	9.7 (1.6–28.2)
Within normal range (11–21)	35	74.5 (57.6–87.3)	26	83.9 (63.8–95.4)
Elevated (22–29)	1	2.1 (0–12.8)	2	6.5 (0.5–23.7)
High (≥ 30)	1	2.1 (0–12.8)	0	0 (0–13.2)
Worst eye cup:disc ratio§				
Bilateral ungradable	1	–	0	–
Unilateral ungradable	11	–	8	–
Normal (0.1–0.6)	73	90 (80.1–96.2)	79	75.2 (64.5–84.1)
Moderate (0.7–0.8)	7	8.6 (3.1–18.3)	23	21.9 (13.5–32.3)
Advanced (≥ 0.9)	1	1.2 (0–7.6)	3	2.9 (0.5–9)
Cataract				
All cataract	19	23.2 (13.6–35.3)	18	17 (9.7–27)
White cataract	6	7.3 (2.3–16.5)	0	0 (0–4.1)
White cataract with hypotony, IOP ≤ 5 mm Hg¶	4	80 (23.6–99.7)	NA	NA
Active anterior uveitis				
Missing data	13	–	67	–
Anterior chamber cells present	5	7.3 (2–17.4)	4	10.5 (2.4–27)
Previous anterior uveitis				
Missing data	12	–	65	–
Signs of previous anterior uveitis#	7	10 (3.6–21)	0	0 (0–10.4)
Vitreous signs**				
Signs suggestive of active or past intermediate uveitis	8 (9.8)	9.8 (3.8–19.6)	14	13.3 (6.9–22.5)
Retinal signs**				
Retinal hemorrhages	0	0 (0–5.2)	2	1.9 (0.2–7.5)
Retinal neovascularization	0	0 (0–5.2)	1	1 (0–5.9)
Papilledema	0	0 (0–5.2)	0	0 (0–4.1)
Retinal vasculitis	0	0 (0–5.2)	4	3.8 (0.8–10.4)
Macula hole	0	0 (0–5.2)	1	1 (0–5.9)
Retinal tears	1	1.2 (0–7.5)	1	1 (0–5.9)
Retinal detachment	0	0 (0–5.2)	2	1.9 (0.2–7.5)
Asteroid hyalosis	0	0 (0–5.2)	1	1 (0–5.9)
Myelinated nerve fibers	0	0 (0–5.2)	1	1 (0–5.9)
Benign flecked retina	1	1.2 (0–7.5)	0	0 (0–4.1)
Geographic retinal darkening and variants	16	19.5 (10.7–31.2)	13	12.4 (6.2–21.4)
White without pressure	18	22 (12.6–34)	20	19 (11.2–29.2)

*IOP, intraocular pressure; NA, not available; –, not applicable.

†Calculated by using exact binomial Clopper-Pearson method.

‡Grading based on International Classification of Diseases, Ninth Revision, Clinical Modification (true Snellen fractions).

§When only 1 cup:disc ratio was gradable, only that ratio was used for analysis.

¶Missing data on 2 patients.

#Posterior synechiae and/or pigment on anterior lens capsule, keratic precipitates but no anterior chamber inflammation, or both.

**Graded based on widefield retinal image.

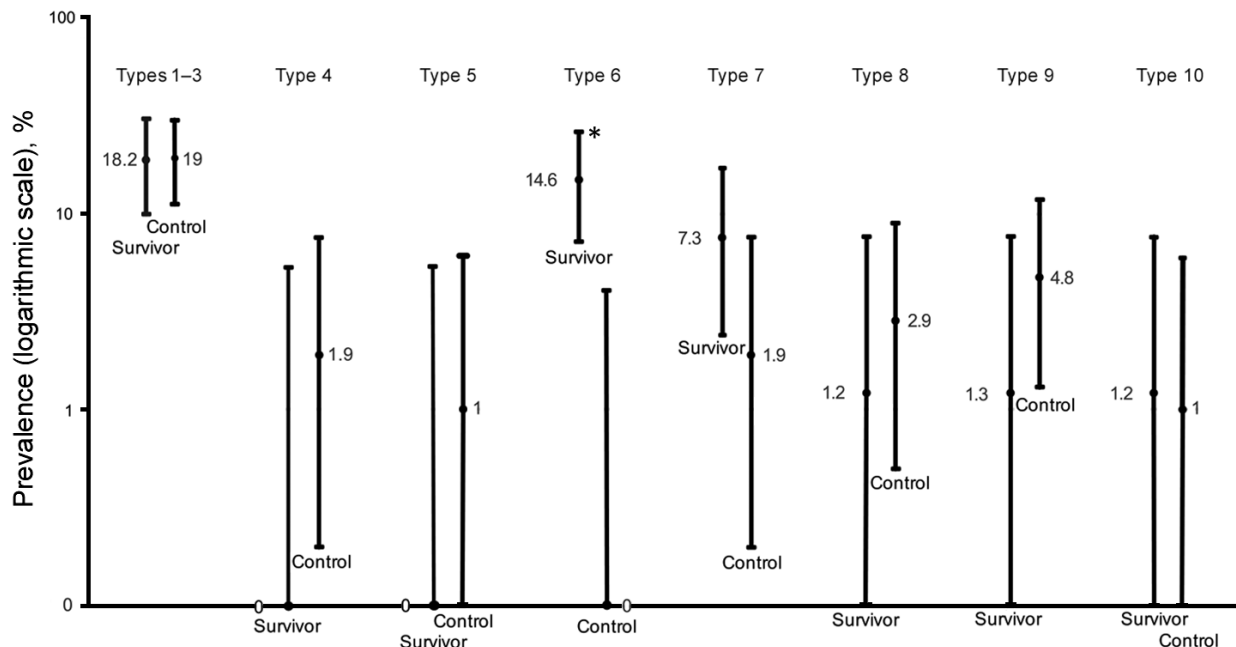


Figure 1. Prevalence of retinal scar lesion types in a case–control study of ocular signs in Ebola virus disease survivors, Sierra Leone, 2016. Type 1, uniform pigmented lesion; type 2, uniform pigmented lesion with gray halo; type 3, uniform pigmented lesion with lacunae; type 4, pigmented lesion with deep surrounding atrophy; type 5, previously described lesion attributed to Ebola (8); type 6, angulated lesions (peripapillary and/or peripheral); type 7, indistinct small pigmented lesions; type 8, irregularly pigmented vascular projection lesion; type 9, pigmented curvilinear peripheral bands; type 10, optic disc projection to macula lesion. Error bars indicate 97.5% CI. Asterisk indicates statistical significance ($p < 0.01$) based on Fisher exact statistic value (2.7×10^5).

[97.5% CI 0%–4.1%]) ($p < 0.01$). In 50% of EVD survivors, this type of lesion was observed bilaterally.

Two fundal distributions of type 6 lesions were evident: isolated or multifocal lesions in the peripheral retina or peripapillary lesions observed emanating from the optic disc (Figure 2). Each lesion shape was variable but often exhibited characteristic sharp angulations, resembling a diamond or wedge (Figure 3). Surrounding these lesions was a well-demarcated area of darkened retina in comparison with the adjacent retina. Presence of any retinal lesions of types 1–10, excluding type 6, were observed in 21/82 (25.6% [97.5% CI 15.5%–38%]) EVD survivors and 25/105 (23.8% [97.5% CI 15.1%–34.4%]) controls.

The aqueous humor of 2 EVD survivors with white cataract and no anterior chamber inflammation was negative for EBOV RNA on qRT-PCR assay. Postprocedure conjunctival swabs also were negative. The aqueous humor sampling procedure was uncomplicated and well-tolerated. No complications were reported on follow up.

Discussion

This case–control study identified a novel retinal sign that appears to be specific to EVD survivors. This sign occurred among a local population with a high rate of background chorioretinal disease. Uveitis after EVD has been reported (3,8), and a recent case report included a published fundus

image from a survivor with a chorioretinal lesion attributed to EVD (9). That patient went on to have panuveitis.

The retinal lesions specific to EVD survivors were located either adjacent to the optic disc or in the fundus periphery. In the 8 cases in which lesions appear adjacent to the optic disc, their curvilinear projections from the disc margin appear to align with the anatomic pathways of the retinal ganglion cell axons that constitute the optic nerve. This distribution suggests a neurotrophic spread into the eye from the optic nerve and along the retinal ganglion cell axons. The other possible mode of entry into the eye is hematologic. Although the retinal ganglion cell axons often have parallel curvatures around the retinal arcade vessels, the lesions clearly follow the nerve fiber distribution in the absence of major vessels (Figure 1, panels A and C). Furthermore, we have not found any signs suggestive of associated vascular involvement, such as vasculitis, vascular occlusions, retinal ischemia, or secondary neovascularization, to support a hematologic spread. Neurotrophic properties are increasingly being recognized in EBOV (18). West Nile virus disease, caused by a known neurotropic virus, is associated with retinal lesions that follow a similar pattern of distribution to the pattern we have observed in our study (19).

Each Ebola lesion shape is variable, but a characteristic angulated appearance often resembling a diamond or wedge shape appears unique (Figure 2). As far as we are

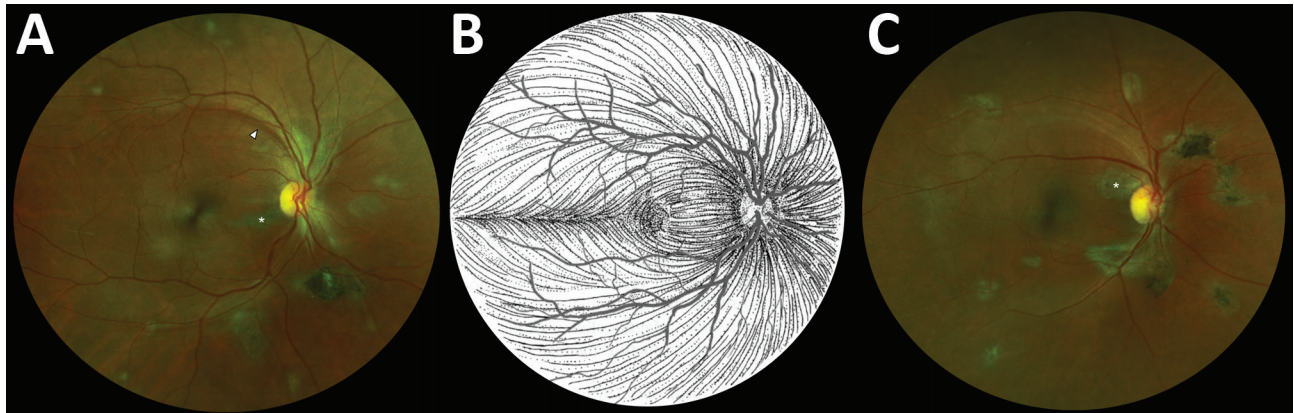


Figure 2. Composite scanning laser ophthalmoscope retinal images showing type 6 Ebola peripapillary and peripheral lesions, observed following the anatomic distribution of the ganglion cell axons (retinal nerve fiber layer), in a case-control study of ocular signs in Ebola virus disease survivors, Sierra Leone, 2016. A) Example 1, right eye. B) Illustration of the ganglion cell axon anatomic distribution. Courtesy of W.L.M. Alward. C) Example 2, right eye. Asterisks indicate curvilinear lesions distinct from the retinal vasculature. White arrowhead indicates retinal nerve fiber wedge defect.

aware, the appearance of these lesions is not characteristic of any other retinal disease. The reason for the sharp angulated appearance of these lesions might be explained by the tight triangular packing of the retinal cone mosaic (Figure 3, panel D) (17,20). The regular pattern of the photoreceptor triangular mosaic is disrupted by larger blue cones (17) and diminishes with eccentricity (20), which might explain the variability in shape. Optical coherence tomography indicates that these lesions are limited to the retina (Figure 3, panel B), and the resemblance of the lesion shape to the photoreceptor mosaic suggests that the ganglion cell axons act merely as a means of transportation to the photoreceptor end target.

Despite the proximity of the lesions to the optic nerve head, we observed no optic nerve head swelling or pallor in our study. This fact is in contrast to the 10% of optic nerve swelling reported in 1 abstract (5), although the time from acute infection to ophthalmic examination in that case was not stated. This difference might be attributable to varying durations since acute infection, allowing for any potential disc swelling to resolve in our cohort, for whom the median time since discharge was 411 days. Further optic nerve functional assessment, such as visual field analysis or color vision testing, has yet to be conducted.

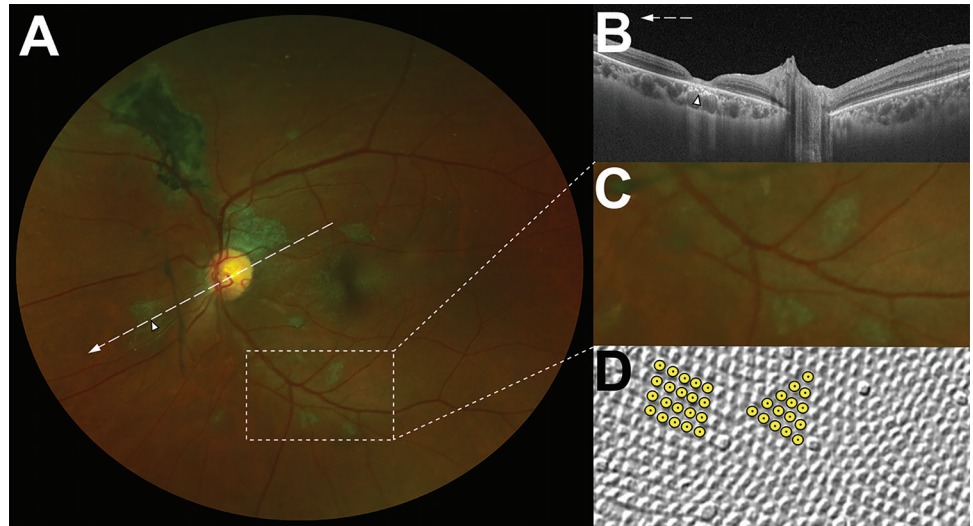
The Ebola retinal lesions did not affect visual acuity. Overall, no difference was observed in uncorrected visual acuity between EVD survivors and controls. The most common cause of visual impairment in EVD survivors was white cataract (7.3%), which was accompanied by hypotony (low intraocular pressure) in 80% of EVD survivors. Hypotony suggests inadequate aqueous humor production and can limit the visual potential of an eye through complications such as retinal folds at the macula (i.e., hypotensive maculopathy).

Concern exists about the safety of cataract surgery in EVD survivors in Sierra Leone because of the unknown duration of EBOV ocular persistence. A sample size of 2 negative aqueous humor samples in this study is too small to make any definitive conclusions but shows that EBOV does not necessarily persist in aqueous humor in those with cataract but no ongoing intraocular inflammation. This finding suggests that cataract surgery can be conducted safely, providing an opportunity to restore vision and remove the stigma of EVD survivor status associated with having a visible white cataract. At present, we would recommend that anterior chamber sampling with EBOV PCR and a negative result should precede cataract surgery. However, cataract surgery might be challenging and visual outcomes disappointing in cases of secondary hypotony, which occurred in 80% of EVD survivors.

Before this study, only 1 aqueous humor sample had been obtained in an EVD survivor (9), enabling the detection of viable EBOV in aqueous humor during acute uveitis 9 weeks after discharge from hospital (9). Virus persistence in aqueous humor has also been observed in uveitis after Marburg virus infection (21), becoming negative on being repeated at 10 weeks (22). In EVD and Marburg virus-associated uveitis, intraocular pressure was markedly elevated (9,21). Although Ebola-related acute uveitis has been reported to be associated with high intraocular pressure, we did not find any evidence of persistently high intraocular pressure in survivors with Ebola retinal lesions.

Uveitis accounts for 24% of blindness in Sierra Leone and is second only to cataracts as the leading cause (23). A proportion of those cataracts might be a consequence of intraocular inflammation, especially in younger patients. Given the high endemic rates of parasitic, viral, and fungal

Figure 3. Characteristic features of lesions observed in a case-control study of ocular signs in Ebola virus disease survivors, Sierra Leone, 2016. A) Composite scanning laser ophthalmoscope retinal image, left eye. Arrow indicates direction of the optical coherence tomography scan. B) Optical coherence tomography. White, long, dashed line indicates cross-sectional plane; white arrowhead indicates Ebola lesion limited to the retinal layers with an intact retinal pigment epithelium. C) Examples of straight-edged, sharp angulated lesions (magnified from panel A 1.5 \times). D) Example of tangential section through the human fovea with illustrative highlighting of a triangular photoreceptor matrix corresponding to Ebola lesional shape. Courtesy of Ahnelt et al. (17).



disease in the region, infectious uveitis is likely to have a higher prevalence than in Western populations (24). Nevertheless, the proportion of controls with chorioretinal lesions and retinal vasculitis was surprising. Pigmented and atrophic chorioretinal scars not in keeping with the Ebola retinal lesions were no more common in EVD survivors than controls, and it is important not to attribute these findings to EBOV infection in survivors documented in case series (7,12).

The leading cause of uveitis in Sierra Leone is onchocerciasis, but this disease is in decline because of the systematic distribution of ivermectin to affected areas (25,26). The rate of other uveitis-associated blindness appears to be increasing in Sierra Leone (23). This study was conducted in Freetown, where the incidence of onchocerciasis is lower than in rural regions, and other causes are probably responsible. Toxoplasmosis accounted for 43% of symptomatic cases of posterior uveitis in 1 study (27), and it was probably a common cause among the patients in our study, although no serologic testing for toxoplasmosis was available. HIV prevalence in persons ≥ 15 years of age in Sierra Leone was estimated to be 1.25% in 2015 (28). The Ebola outbreak disrupted the fragile health system, including HIV reporting mechanisms and AIDS response (29). This HIV rate is still relatively low compared with many other African nations. Further diagnostic investigation is required to attempt to attribute causation to the various chorioretinal lesions observed in this study. Geographic areas of retinal whitening (white without pressure) are thought to be normal variants (30,31). Areas of retinal darkening (dark without pressure) have previously been attributed to sickle cell disease (32).

Our study is subject to 1 limitation with regard to the control group, who were selected opportunistically with unmatched cases and controls, and differences in age and sex ratios between the groups. This fact reflects the difficulties and limitations of conducting research in the post-Ebola setting in Freetown in 2016. The study was conducted in a military hospital, which housed the Ebola treatment unit and the continuing EVD survivors clinic. The hospital also serves the local civilian community and a military barracks community. The use of a non-EVD control group, even without matching, allowed a comparison in the fundus findings between post-EVD and control groups. We found a higher prevalence of retinal disease in the symptomatic clinic-attending control group than in the asymptomatic population control group; both groups included some military members of staff and families. This comparison allows us to be more positive about the specificity of the Ebola retinal lesion. Given our aim to compare EVD with non-EVD fundus findings, an age- and sex-matched population control group probably would not change the study conclusions.

EVD survivors were identified by the possession of an Ebola treatment center discharge certificate. Forgery of these certificates has been known given the free access to healthcare it confers. IgG confirmation of previous EBOV infection is planned for ongoing follow-up studies. Our study provides information on the medium-term ocular sequelae of EVD survivors with a median time of 411 days since hospital discharge. Our study does not provide data on acute uveitis and ocular disease in the immediate aftermath of EVD as reported elsewhere (2,4,6).

Although we can reasonably conclude the retinal lesions described in our study are sequelae of EVD, no

pre-EVD retinal imaging was available to conclusively identify the timing of acquisition of the lesions. Our control group demonstrates the common retinal signs and pathologies that are present in the population before Ebola exposure.

We have documented a novel retinal abnormality in EVD survivors that appears to be specific to EVD, although the proportion in the cohort with the condition is small. The background prevalence of chorioretinal abnormalities, including scarring with pigmentation, in the population is high and should not be attributed to EVD. Although further studies with larger sample sizes are required, EBOV does not necessarily persist in the aqueous humor of those with cataracts and no ongoing intraocular inflammation. These initial results raise the possibility of safe cataract surgery for EVD survivors with no signs of ongoing intraocular inflammation.

Acknowledgments

We thank Wallace L.M. Alward for permission to use his retinal nerve fiber layer illustration; Optos PLC for their generous donation of the Daytona Ophthalmoscope, which continues to improve patient care for the people of Sierra Leone; Onlime SL Ltd. for supplying the clinics at 34th Regiment Military Hospital with internet access; Medisoft, for the offer of electronic patient record software (although not used in this study); and the administration at 34th Regiment Military Hospital for supporting and facilitating the study and for upgrading the eye clinic. Thank you to the patients and subjects for agreeing to participate in this study.

This work was funded by The Dowager Countess Eleanor Peel Trust, Bayer Global Ophthalmology Awards Programme, and Enhancing Research Activity in Epidemic Situations (ERAES) Wellcome Trust Programme. The sponsors of the study had no role in study design, data collection, data analysis, data interpretation, or writing of the manuscript. In addition, J.T.S. and M.G.S. are supported by the National Institute for Health Research Health Protection Research Unit in Emerging and Zoonotic Infections at the University of Liverpool.

P.J.S. had full access to all the data in the study and takes responsibility for the integrity of the data and the accuracy of the data analysis. P.J.S., J.T.S., N.A.V.B., and M.G.S. conceived and designed the study. P.J.S., J.T.S., and M.G.S. wrote the ethics submission. P.J.S., J.T.S., and M.G.S. wrote the specimen collection protocols. P.J.S., J.T.S., and M.G.S. wrote the consent forms, and F.M., A.D.F., and P.K. translated them into Krio. P.J.S. trained and initially supervised staff in ophthalmic examination and imaging. M.G.S. supervised consent and aqueous humor specimen collection/transport and maintained quality assurance over procedures. P.J.S. and F.M. undertook aqueous humor collection. P.J.S., F.M., A.D.F., and P.K. were responsible for data collection and data storage. P.J.S., M.G.S.,

and N.A.V.B. developed the classification of image abnormalities and the data analysis plan. P.J.S., C.K.P., R.D., J.M.B., and N.A.V.B. performed image grading and the data analysis. N.A.V.B. provided the final arbitration of image grading. P.J.S., J.T.S., and G.C. wrote the statistical analysis plan. J.R. performed aqueous humor laboratory analysis. P.J.S. and N.A.V.B. drafted the manuscript. P.J.S. and M.G.S. drafted the paper, and all other authors reviewed and approved the final version.

Dr. Steptoe is a clinical research fellow at the University of Liverpool and a specialist ophthalmology trainee in the Mersey region, UK. His research interests include tropical ophthalmology with an emphasis on ophthalmic infections and uveitis.

References

1. WHO. Ebola situation report 30th March 2016 [cited 2016 Jul 5]. <http://apps.who.int/ebola/current-situation/ebola-situation-report-30-march-2016>
2. Scott JT, Sesay FR, Massaquoi TA, Idriss BR, Sahr F, Semple MG. Post-Ebola Syndrome, Sierra Leone. *Emerg Infect Dis*. 2016;22:641–6. <http://dx.doi.org/10.3201/eid2204.151302>
3. Kibadi K, Mupapa K, Kuvula K, Massamba M, Ndaberey D, Muyembe-Tamfum JJ, et al. Late ophthalmologic manifestations in survivors of the 1995 Ebola virus epidemic in Kikwit, Democratic Republic of the Congo. *J Infect Dis*. 1999;179(Suppl 1):S13–4. <http://dx.doi.org/10.1086/514288>
4. Mattia JG, Vandy MJ, Chang JC, Platt DE, Dierberg K, Bausch DG, et al. Early clinical sequelae of Ebola virus disease in Sierra Leone: a cross-sectional study. *Lancet Infect Dis*. 2016;16:331–8. [http://dx.doi.org/10.1016/S1473-3099\(15\)00489-2](http://dx.doi.org/10.1016/S1473-3099(15)00489-2)
5. Bishop R, Eghrari A, Brady C, Ray V, Reilly C, Ferris F, et al. Expanding the spectrum of Ebola-associated eye disease: a summary of ocular findings in a large cohort of Ebola survivors. *Invest Ophthalmol Vis Sci*. 2016. ARVO E-abstract 4763 [cited 2016 Jul 5]. http://www.arvo.org/webs/am2016/sectionpdf/IM/Session_433.pdf
6. Tiffany A, Vetter P, Mattia J, Dayer J-A, Bartsch M, Kasztura M, et al. Ebola virus disease complications as experienced by survivors in Sierra Leone. *Clin Infect Dis*. 2016;62:1360–6. <http://dx.doi.org/10.1093/cid/ciw158>
7. Hereth-Hebert E, Bah MO, Etard JF, Sow MS, Resnikoff S, Fardeau C, et al.; Postebogui Study Group. Ocular complications in survivors of the Ebola outbreak in Guinea. *Am J Ophthalmol*. 2017;175:114–21. <http://dx.doi.org/10.1016/j.ajo.2016.12.005>
8. Hebert EH, Sow S, Etard Jean-François TA, Msellati P, Taverne B, Barry M, et al. A case series of 11 uveitis in patients who recovered from Ebola in Guinea. *Trop Med Int Health*. 2015;20(Suppl. 1):435.
9. Varkey JB, Shantha JG, Crozier I, Kraft CS, Lyon GM, Mehta AK, et al. Persistence of Ebola virus in ocular fluid during convalescence. *N Engl J Med*. 2015;372:2423–7. <http://dx.doi.org/10.1056/NEJMoa1500306>
10. Shantha JG, Crozier I, Varkey JB, Kraft CS, Lyon GM III, Mehta AK, et al. Long-term Management of Panuveitis and Iris Heterochromia in an Ebola Survivor. *Ophthalmology*. 2016;123:2626–2628.e2. <http://dx.doi.org/10.1016/j.ophtha.2016.07.013>
11. Chancellor JR, Padmanabhan SP, Greenough TC, Sacra R, Ellison RT III, Madoff LC, et al. Uveitis and systemic inflammatory markers in convalescent phase of Ebola virus disease. *Emerg Infect Dis*. 2016;22:295–7. <http://dx.doi.org/10.3201/eid2202.151416>

12. Shantha JG, Crozier I, Hayek BR, Bruce BB, Gargu C, Brown J, et al. Ophthalmic manifestations and causes of vision impairment in Ebola virus disease survivors in Monrovia, Liberia. *Ophthalmology*. 2017;124:170–7. <http://dx.doi.org/10.1016/j.ophtha.2016.10.011>
13. von Elm E, Altman DG, Egger M, Pocock SJ, Gøtzsche PC, Vandenbroucke JP; STROBE Initiative. The Strengthening of Reporting of Observational Studies in Epidemiology (STROBE) statement: guidelines for reporting observational studies. *Lancet*. 2007;370:1453–7. [http://dx.doi.org/10.1016/S0140-6736\(07\)61602-X](http://dx.doi.org/10.1016/S0140-6736(07)61602-X)
14. Trusko B, Thorne J, Jabs D, Belfort R, Dick A, Gangaputra S, et al.; Standardization of Uveitis Nomenclature (SUN) Project. The Standardization of Uveitis Nomenclature (SUN) Project. Development of a clinical evidence base utilizing informatics tools and techniques. *Methods Inf Med*. 2013;52:259–65, S1–6. <https://doi.org/10.3414/ME12-01-0063>
15. Chylack LT Jr, Wolfe JK, Singer DM, Leske MC, Bullimore MA, Bailey IL, et al.; The Longitudinal Study of Cataract Study Group. The Lens Opacities Classification System III. *Arch Ophthalmol*. 1993;111:831–6. <http://dx.doi.org/10.1001/archoph.1993.01090060119035>
16. Clopper CJ, Pearson ES. The use of confidence or fiducial limits illustrated in the case of the binomial. *Biometrika*. 1934;26:404–13. <http://dx.doi.org/10.1093/biomet/26.4.404>
17. Ahnelt PK, Kolb H, Pflug R. Identification of a subtype of cone photoreceptor, likely to be blue sensitive, in the human retina. *J Comp Neurol*. 1987;255:18–34. <http://dx.doi.org/10.1002/cne.902550103>
18. Billioux BJ, Smith B, Nath A. Neurological complications of Ebola virus infection. *Neurotherapeutics*. 2016;13:461–70. <http://dx.doi.org/10.1007/s13311-016-0457-z>
19. Pleyer U, Klauf V, Wilking H, Nentwich MM. [Tropical ophthalmology : Intraocular inflammation caused by “new” infectious pathogens and travel-related infections]. *Ophthalmologe*. 2016;113:35–46. <http://dx.doi.org/10.1007/s00347-015-0176-6>
20. Hugh Perry V. The distribution of cones in the primate retina. In: *advances in photoreception: proceedings of a symposium on Frontiers of Visual Science*. Washington: National Academies Press; 1990. p. 105–16.
21. Kuming BS, Kokoris N. Uveal involvement in Marburg virus disease. *Br J Ophthalmol*. 1977;61:265–6. <http://dx.doi.org/10.1136/bjo.61.4.265>
22. Gear JS, Cassel GA, Gear AJ, Trappler B, Clausen L, Meyers AM, et al. Outbreak of Marburg virus disease in Johannesburg. *BMJ*. 1975;4:489–93. <http://dx.doi.org/10.1136/bmj.4.5995.489>
23. Ronday MJ, Stilma JS, Barbe RF, Kijlstra A, Rothova A. Blindness from uveitis in a hospital population in Sierra Leone. *Br J Ophthalmol*. 1994;78:690–3. <http://dx.doi.org/10.1136/bjo.78.9.690>
24. Onchocerciasis and its control. Report of a WHO Expert Committee on Onchocerciasis Control. *World Health Organ Tech Rep Ser*. 1995;852:1–104.
25. Whitworth JAG, Gilbert CE, Mabey DM, Maude GH, Morgan D, Taylor DW. Effects of repeated doses of ivermectin on ocular onchocerciasis: community-based trial in Sierra Leone. *Lancet*. 1991;338:1100–3. [http://dx.doi.org/10.1016/0140-6736\(91\)91963-U](http://dx.doi.org/10.1016/0140-6736(91)91963-U)
26. Rothova A, Van der Lelij A, Stilma JS, Klaassen-Broekema N, Wilson WR, Barbe RF. Ocular involvement in patients with onchocerciasis after repeated treatment with ivermectin. *Am J Ophthalmol*. 1990;110:6–16. [http://dx.doi.org/10.1016/S0002-9394\(14\)76930-2](http://dx.doi.org/10.1016/S0002-9394(14)76930-2)
27. Ronday MJ, Stilma JS, Barbe RF, McElroy WJ, Luyendijk L, Kolk AH, et al. Aetiology of uveitis in Sierra Leone, west Africa. *Br J Ophthalmol*. 1996;80:956–61. <http://dx.doi.org/10.1136/bjo.80.11.956>
28. World Health Organization. HIV/AIDS [cited 2017 Mar 29]. <http://www.afro.who.int/en/sierra-leone/health-topics/4958-hiv-aids.html>
29. Parpia AS, Ndeffo-Mbah ML, Wenzel NS, Galvani AP. Effects of response to 2014–2015 Ebola outbreak on deaths from malaria, HIV/AIDS, and tuberculosis, West Africa. *Emerg Infect Dis*. 2016;22:433–41. <http://dx.doi.org/10.3201/eid2203.150977>
30. Rutnin U, Schepens CL. Fundus appearance in normal eyes. IV. Retinal breaks and other findings. *Am J Ophthalmol*. 1967;64:1063–78. [http://dx.doi.org/10.1016/0002-9394\(67\)93057-7](http://dx.doi.org/10.1016/0002-9394(67)93057-7)
31. Diaz RI, Sigler EJ, Randolph JC, Rafieetay MR, Calzada JI. Spectral domain optical coherence tomography characteristics of white-without-pressure. *Retina*. 2014;34:1020–1. <http://dx.doi.org/10.1097/IAE.0000000000000012>
32. Nagpal KC, Goldberg MF, Asdourian G, Goldbaum M, Huamonte F. Dark-without-pressure fundus lesions. *Br J Ophthalmol*. 1975;59:476–9. <http://dx.doi.org/10.1136/bjo.59.9.476>

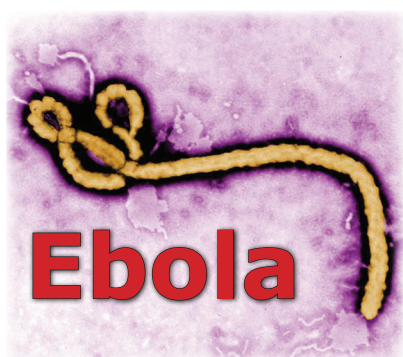
Address for correspondence: Paul J. Steptoe, Institute in the Park, Alder Hey Children’s Hospital, Eaton Rd, Liverpool, Merseyside, L12 2AP, UK; email: paul.steptoe@liverpool.ac.uk

EID SPOTLIGHT TOPIC

Ebola, previously known as Ebola hemorrhagic fever, is a rare and deadly disease caused by infection with one of the Ebola virus strains. Ebola can cause disease in humans and nonhuman primates (monkeys, gorillas, and chimpanzees).

Ebola is caused by infection with a virus of the family *Filoviridae*, genus *Ebolavirus*. There are five identified Ebola virus species, four of which are known to cause disease in humans. Ebola viruses are found in several African countries; they were first discovered in 1976 near the Ebola River in what is now the Democratic Republic of the Congo. Before the current outbreak, Ebola had appeared sporadically in Africa.

The natural reservoir host of Ebola virus remains unknown. However, on the basis of evidence and the nature of similar viruses, researchers believe that the virus is animal-borne and that bats are the most likely reservoir. Four of the five virus strains occur in an animal host native to Africa.



**EMERGING
INFECTIOUS DISEASES**

<http://wwwnc.cdc.gov/eid/page/ebola-spotlight>

Novel Retinal Lesion in Ebola Survivors, Sierra Leone, 2016

Technical Appendix 1

SL Retinal Image Grading Form

***Required**

1. Reviewer * *Mark only one oval.*

P S

C P

J B

R D

2. Enter Retinal Image Number *

.....

3. Select which eye * *Mark only one answer.*

Left *Skip to question 5.*

Right *Skip to question 4.*

Right Eye

4. Is there any view of the right fundus that can be classified? *
Mark only one answer.

Yes *Skip to question 7.*

No, probable lens opacity obscuring fundal view *Stop filling out this form.*

No, probably due to another cause other than cataract *Stop filling out this form.*

Left Eye

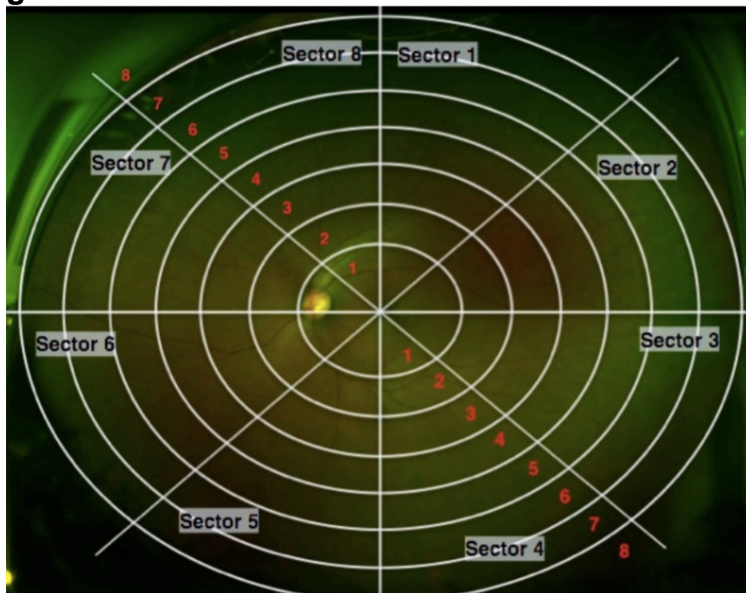
5. Is there any view of the left fundus that can be classified? *

Mark only one answer.

- Yes Skip to question 6.**
- No, probable lens opacity obscuring fundal view Stop filling out this form.**
- No, probably due to a cause other than cataract Stop filling out this form.**

Left Eye Grading Visible Fundus

If there are several images available look at them both as the combination of both images might show more areas of the retina than one image alone. In which case, grade the visible fundus from a combination of images available.



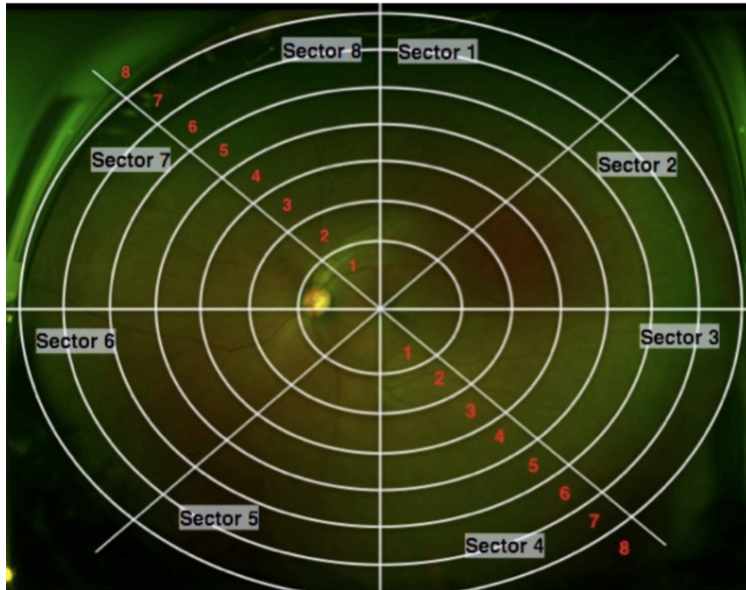
6. Grade the degree of visible retina * Mark only one oval per row.

	1	2	3	4	5	6	7	8
Sector 1	<input type="radio"/>	<input type="radio"/>	<input type="radio"/>	<input type="radio"/>	<input type="radio"/>	<input type="radio"/>	<input type="radio"/>	<input type="radio"/>
Sector 2	<input type="radio"/>	<input type="radio"/>	<input type="radio"/>	<input type="radio"/>	<input type="radio"/>	<input type="radio"/>	<input type="radio"/>	<input type="radio"/>
Sector 3	<input type="radio"/>	<input type="radio"/>	<input type="radio"/>	<input type="radio"/>	<input type="radio"/>	<input type="radio"/>	<input type="radio"/>	<input type="radio"/>
Sector 4	<input type="radio"/>	<input type="radio"/>	<input type="radio"/>	<input type="radio"/>	<input type="radio"/>	<input type="radio"/>	<input type="radio"/>	<input type="radio"/>
Sector 5	<input type="radio"/>	<input type="radio"/>	<input type="radio"/>	<input type="radio"/>	<input type="radio"/>	<input type="radio"/>	<input type="radio"/>	<input type="radio"/>
Sector 6	<input type="radio"/>	<input type="radio"/>	<input type="radio"/>	<input type="radio"/>	<input type="radio"/>	<input type="radio"/>	<input type="radio"/>	<input type="radio"/>
Sector 7	<input type="radio"/>	<input type="radio"/>	<input type="radio"/>	<input type="radio"/>	<input type="radio"/>	<input type="radio"/>	<input type="radio"/>	<input type="radio"/>
Sector 8	<input type="radio"/>	<input type="radio"/>	<input type="radio"/>	<input type="radio"/>	<input type="radio"/>	<input type="radio"/>	<input type="radio"/>	<input type="radio"/>

Skip to question 8.

Right Eye Grading Visible Fundus

If there are several images available look at them both as the combination of both images might show more areas of the retina than one image alone. In which case, grade the visible fundus from a combination of images available

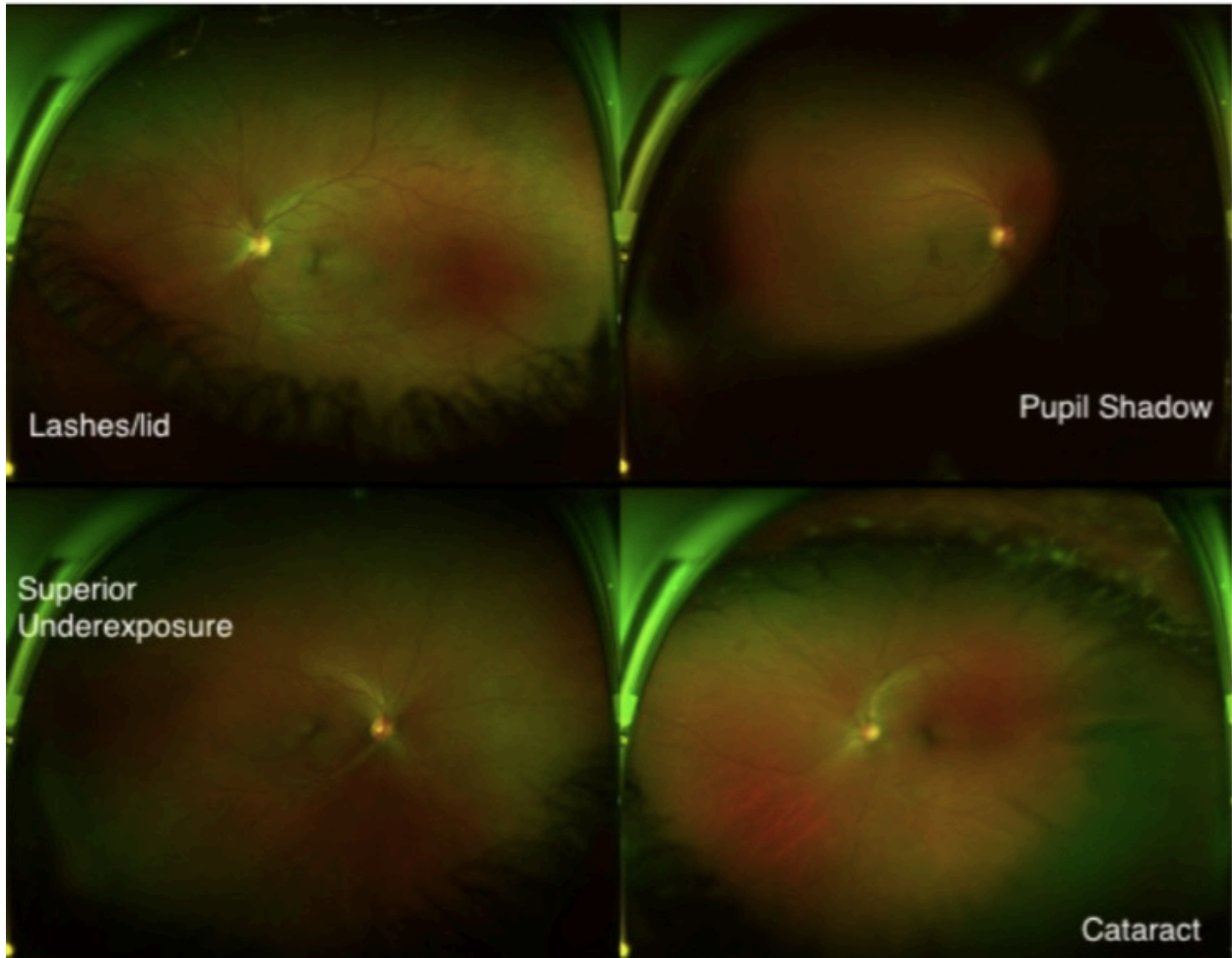


7. Grade the degree of visible retina * *Mark only one oval per row.*

	1	2	3	4	5	6	7	8
Sector 1	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>
Sector 2	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>
Sector 3	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>
Sector 4	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>
Sector 5	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>
Sector 6	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>
Sector 7	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>
Sector 8	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>

Skip to question 8.

Restricted Fundal View

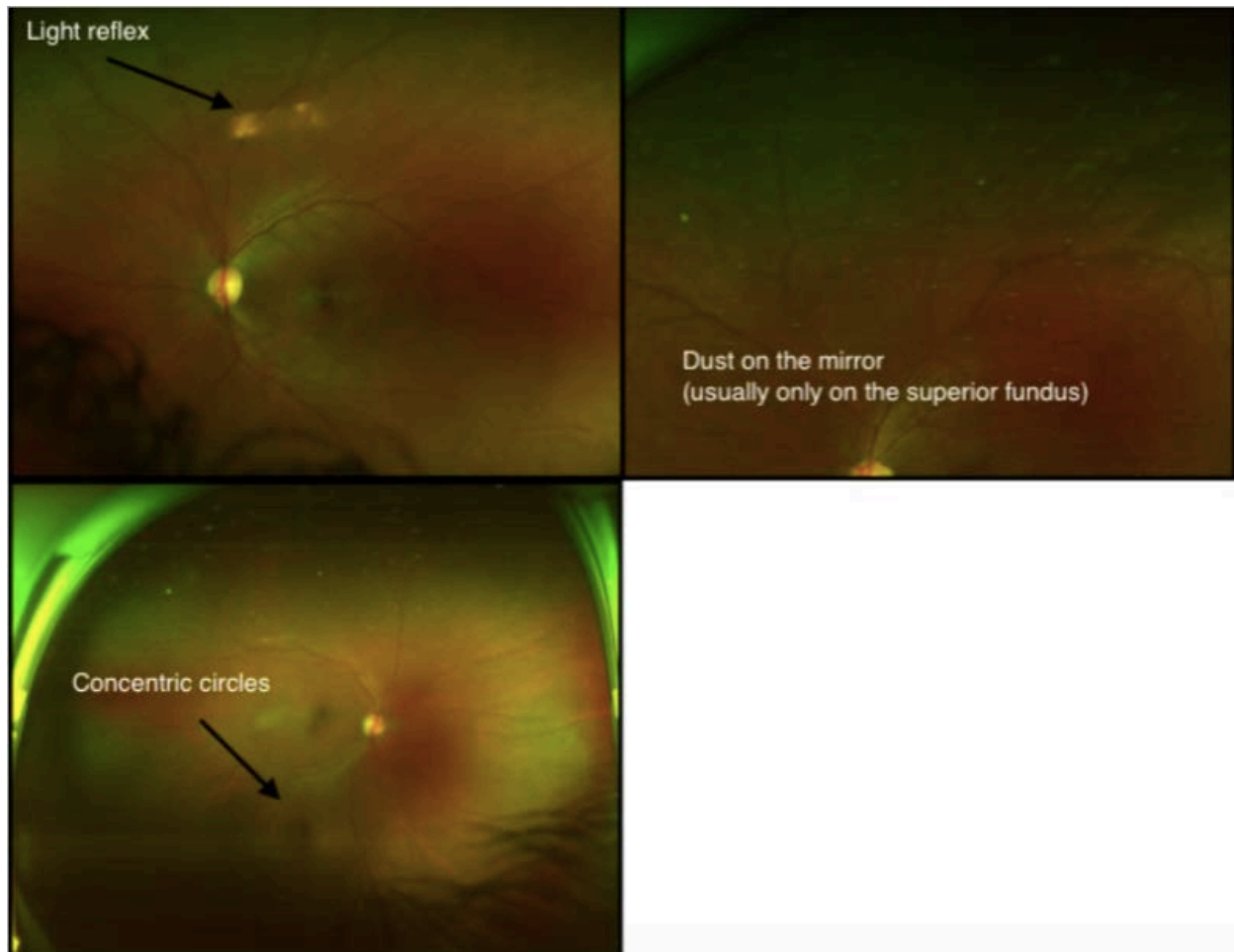


8. What are the contributing factors to the restricted fundal view?
Mark only one oval per row.

	0%	25%	50%	75%	100%
Cataract	<input type="radio"/>	<input type="radio"/>	<input type="radio"/>	<input type="radio"/>	<input type="radio"/>
Pupil Shadow	<input type="radio"/>	<input type="radio"/>	<input type="radio"/>	<input type="radio"/>	<input type="radio"/>
Eye lashes	<input type="radio"/>	<input type="radio"/>	<input type="radio"/>	<input type="radio"/>	<input type="radio"/>
Camera Underexposure	<input type="radio"/>	<input type="radio"/>	<input type="radio"/>	<input type="radio"/>	<input type="radio"/>

Skip to question 9.

Artifactual Findings



9. Are any of the following artifactual findings visible? *Tick all that apply.*

- Superficial camera dust
- Horizontal light reflex
- Concentric circles

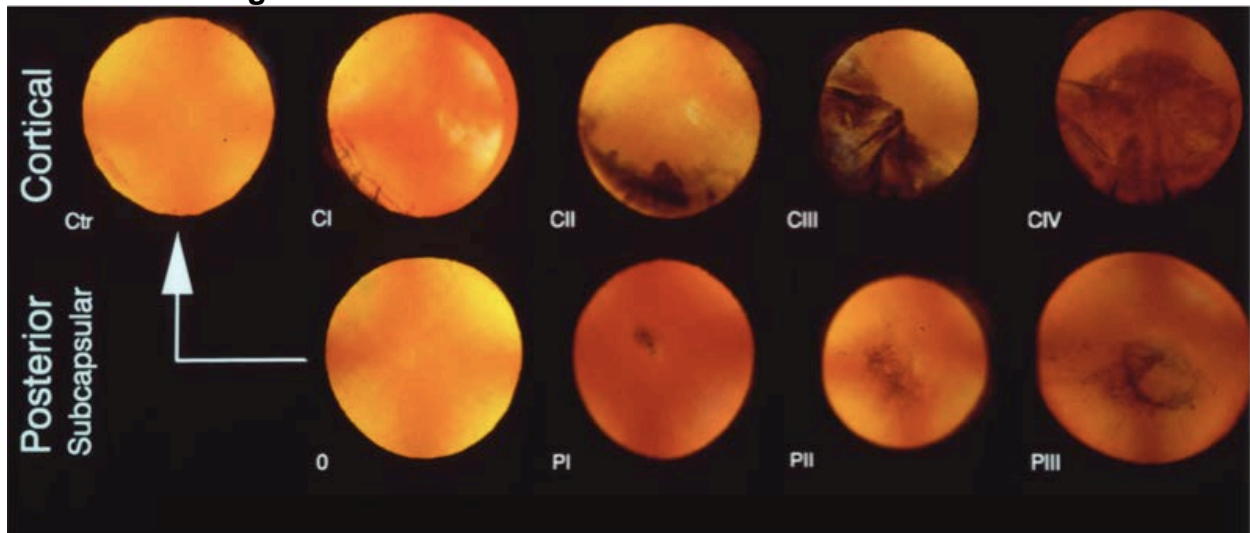
Cataract Assessment

Look carefully at the fundal image. Are there any shadows cast from media opacities, or any cortical cataract spokes?

Is there any evidence of cataract seen on the image? * *Mark only one oval.*

- Yes, Definitely *Skip to question 11.*
- Yes, Questionably *Skip to question 11.*
- No *Skip to question 12.*

Cataract Grading



11. What type of cataract can be seen and how much of the retina does it obscure comparing to the standard images above?

Mark only one oval per row.

	0	I	II	III	IV
Cortical	<input type="radio"/>	<input type="radio"/>	<input type="radio"/>	<input type="radio"/>	<input type="radio"/>
Posterior subcapsular cataract	<input type="radio"/>	<input type="radio"/>	<input type="radio"/>	<input type="radio"/>	<input type="radio"/>
Undefinable	<input type="radio"/>	<input type="radio"/>	<input type="radio"/>	<input type="radio"/>	<input type="radio"/>

Are there any findings to report?

12. Are there any abnormalities that can be seen on the image? *

Mark only one oval.

- Yes, definitely
- Yes, questionably
- No, This looks like a normal fundus *Skip to question 42.*

Vitreous (1)

13. Are there any finding that might be from the vitreous? *

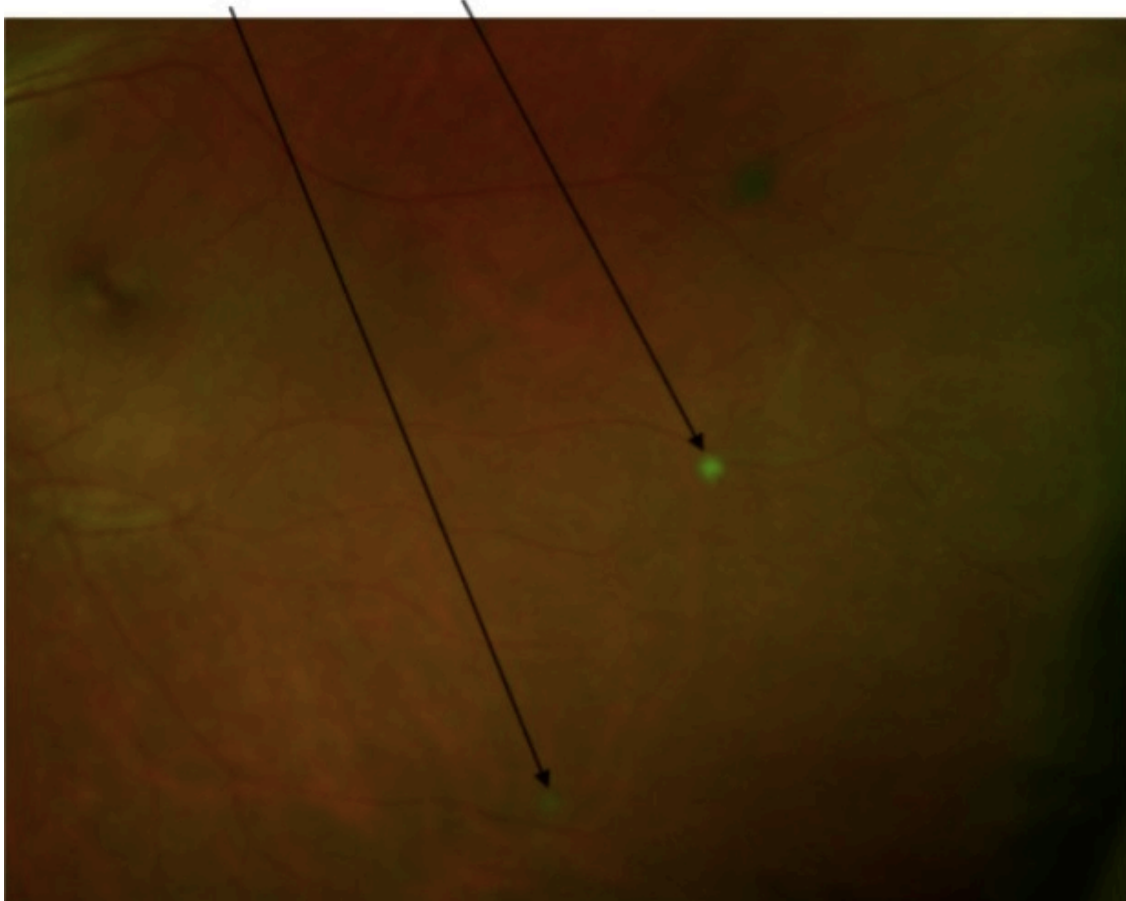
Mark only one oval.

- Yes
- No *Skip to question 21.*

Vitreous Assessment

Snowballs

Round circular vitreous opacities



14. Are there any snowballs visible? * Mark only one oval.

- Yes, definitely
- Yes, questionably
- No *Skip to question 16.*
- View insufficient to see *Skip to question 16.*

15. Count the number of snowballs seen
Mark only one oval.

1

2

3

4

5

6

7

8

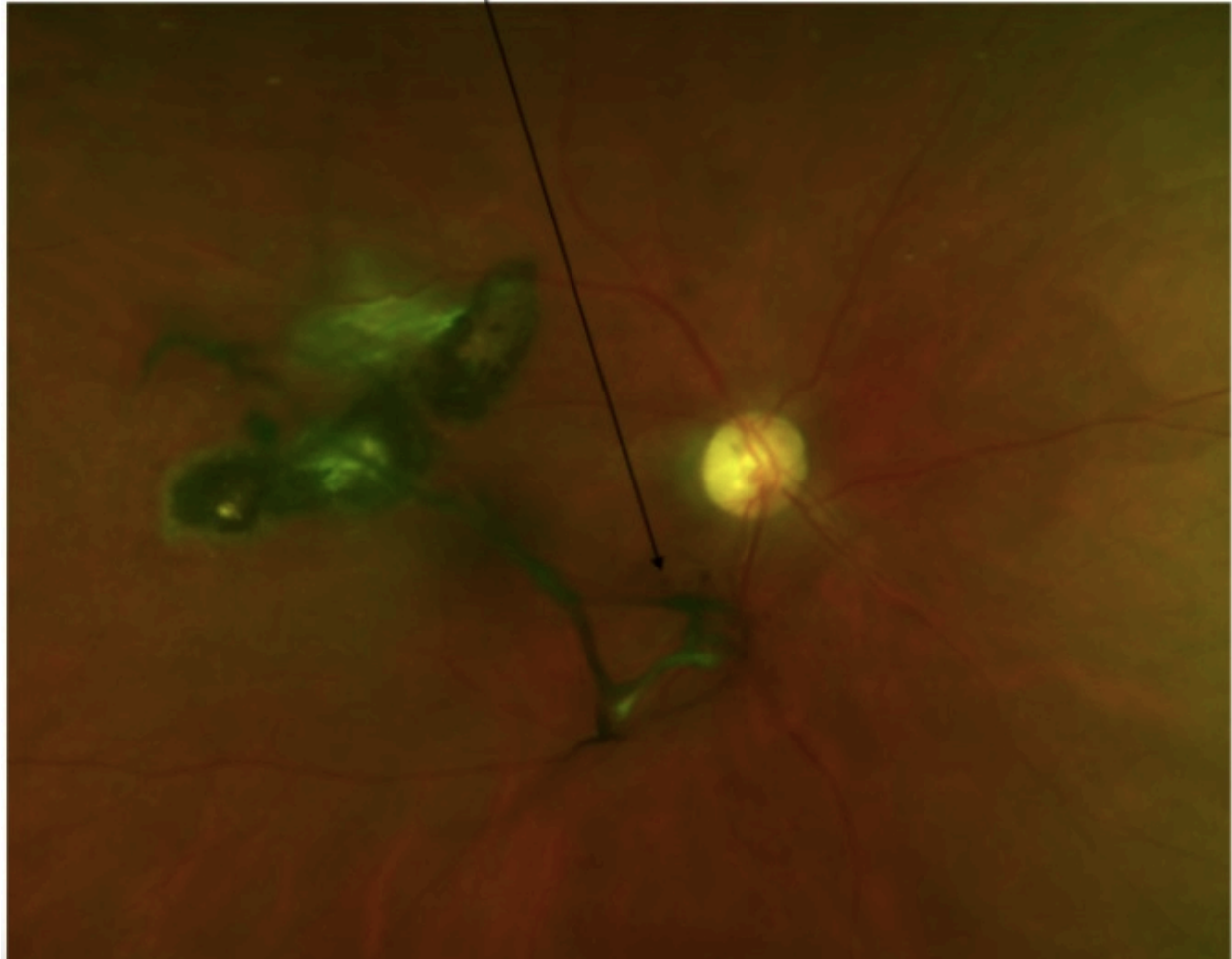
9

10

> 10

Pigmented vitreous floaters

Dark vitreous floater, irregular in shape. Overlying retinal structures



16. Are there any pigmented vitreous floaters?

Mark only one oval.

- Yes, definitely
- Yes, questionably
- No *Skip to question 19.*

Quantifying Vitreous Floaters

17. How many separate pigmented vitreous floaters are there?

Mark only one oval.

- 1
- 2
- 3
- 4
- 5
- 6
- 7
- 8
- 9
- 10
- > 10

18. What is the length of the longest pigmented floater seen in disc diameters?

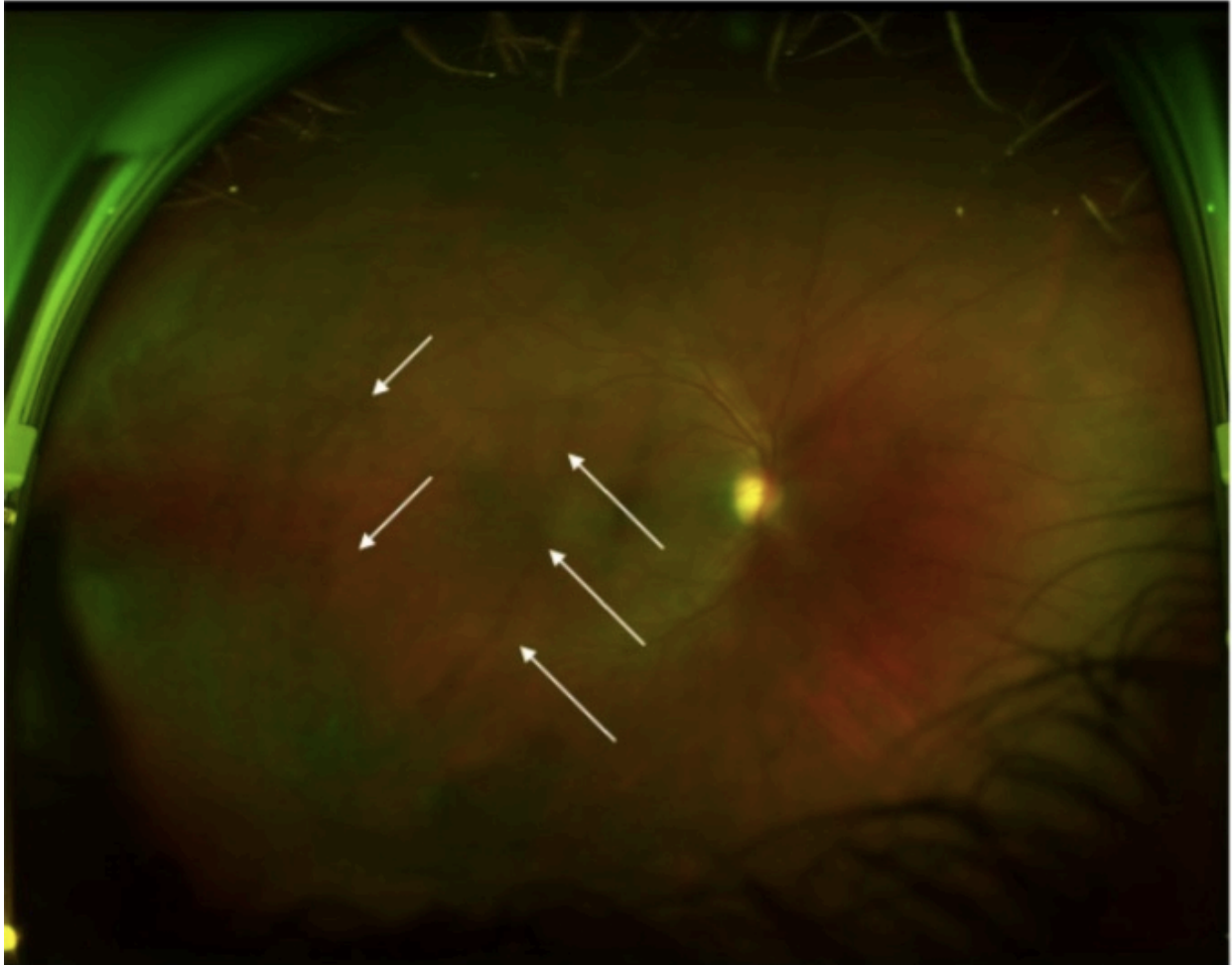
Mark only one oval.

- < 1 disc diameter
- 1 disc diameters
- 2 disc diameters
- 3 disc diameters
- 4 disc diameters
- 5 disc diameters
- 6 disc diameters
- 7 disc diameters
- 8 disc diameters
- 9 disc diameters
- 10 disc diameters

Vitreous Debris

Diffuse Vitreous debris

Often only obvious when 2 sequential fundal images are taken in quick succession and opacities are seen to shift position against the stationary retina.



19. Is there any vitreous debris?

Mark only one oval.

- Yes, definitely
- Yes, questionably
- No

Other Vitreous Observations

20. Are there any of the following signs?

Tick all that apply.

- Weiss ring
- Asteroid Hyalosis
- No

Classification of Fundal Lesions

21. Are there any pigmented lesions?

Mark only one oval.

- Yes, definitely
- Yes, questionably
- No

Pigmented Lesions Group 1

Look at the fundus and compare the lesions to those presented below.

Type 1 - Solid pigmented lesion

Type 2 - Pigmented lesion with a grey surrounding retinal halo

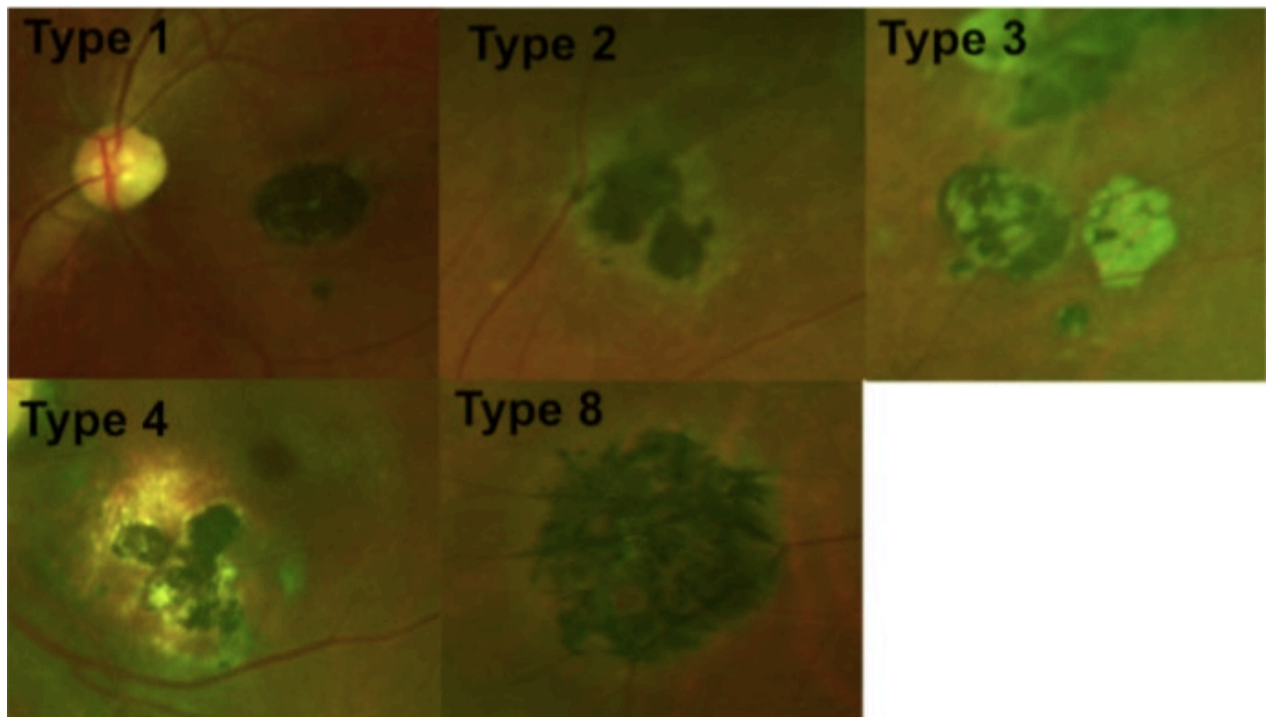
Type 3 - Pigmented lesion with pale lacunae

Type 4 - Pigmented lesion with surrounding deeper retinal involvement characterised by a yellow appearance

Type 8 - Pigmented lesion but non uniform, irregular border with linear projections following surrounding vessels

A single fundus may demonstrate more than one variety of lesion, for example you may see a type 3 lesion with surrounding grey retinal halo in which case select both type 2 and 3.

Also, select if there is any vessel tracking associated with the lesion or vitreous band adhesions



22. Tick all that apply

Mark only one oval per row.

	Yes, definitely	Yes, questionably
Type 1 - Solid pigmented lesion	<input type="radio"/>	<input type="radio"/>
Type 2 - Pigmented with grey halo	<input type="radio"/>	<input type="radio"/>
Type 3 - Pigmented with lacunae	<input type="radio"/>	<input type="radio"/>
Type 4 - Pigmented with yellow atrophy	<input type="radio"/>	<input type="radio"/>
Type 8 - "Spiky" pigmented lesion	<input type="radio"/>	<input type="radio"/>
Is there any pigment tracking along adjacent vessels?	<input type="radio"/>	<input type="radio"/>
Is there any associated vitreous bands?	<input type="radio"/>	<input type="radio"/>

Type 5 Pigmented Lesions

Atrophic lesions surrounding pigmented centre. The edge of the atrophic lesion often has a darker rim

Retinal image published in article J B Varkey et al, Persistence of Ebola Virus in ocular fluid during convalescence. N Engl J Med 2015; 372: 2423-2427

23. Are there any Type 5 lesions visible?

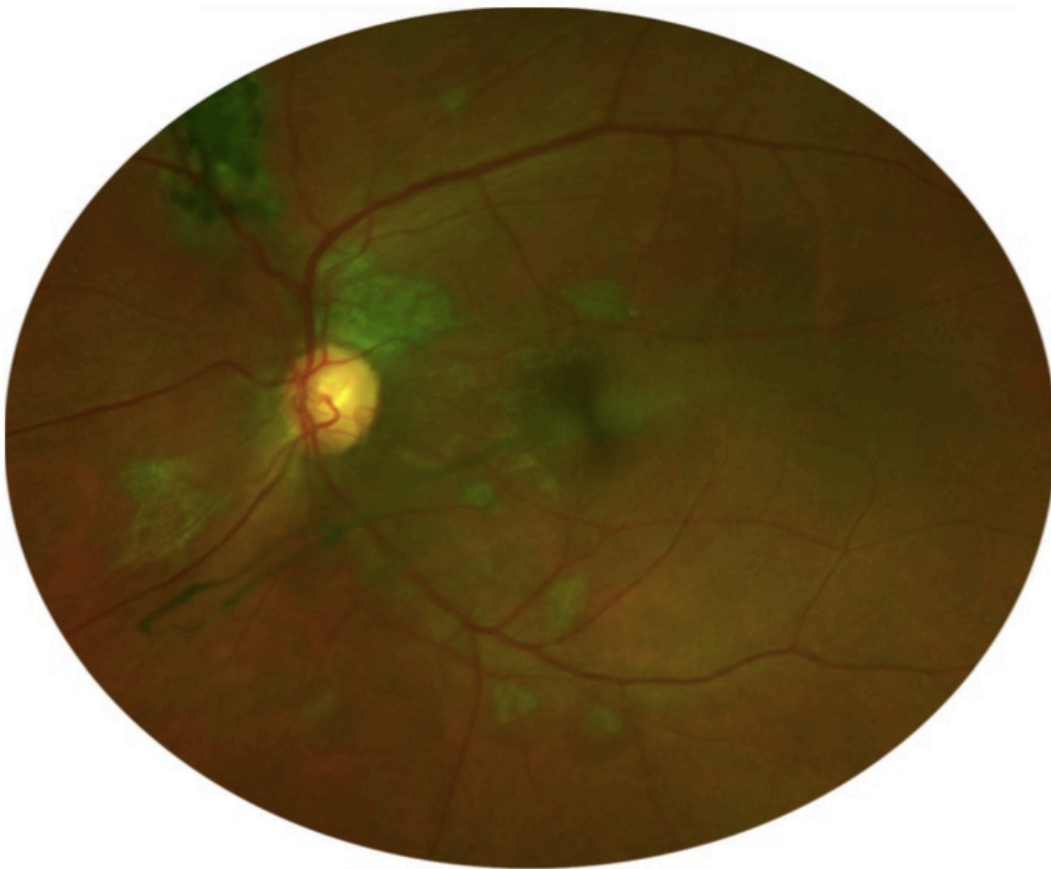
Mark only one oval.

- Yes, definitely
 Yes, questionably
 No **Skip to question 24.**

Type 6 Lesions

These lesions can appear in two forms. Here they can be seen emanating from the optic disc. Alternatively they can be seen in isolation in the peripheral fundus as demonstrated in the following question. They often are surrounded by a dark area of retina which most commonly is isolated to the lesion but in some occasions, extend more peripheral.

In all cases, they appear underneath the retinal vasculature without causing any vessel change. Occasionally large lesions can contain pigmented areas as seen in the top left.



24. Are there any Peripapillary Type 6 Lesions Visible?
Mark only one oval.

- Yes, definitely
- Yes, questionably
- No

Isolated Peripheral Type 6 Lesions

1. Small, often triangular lesions surrounded by darkened retina



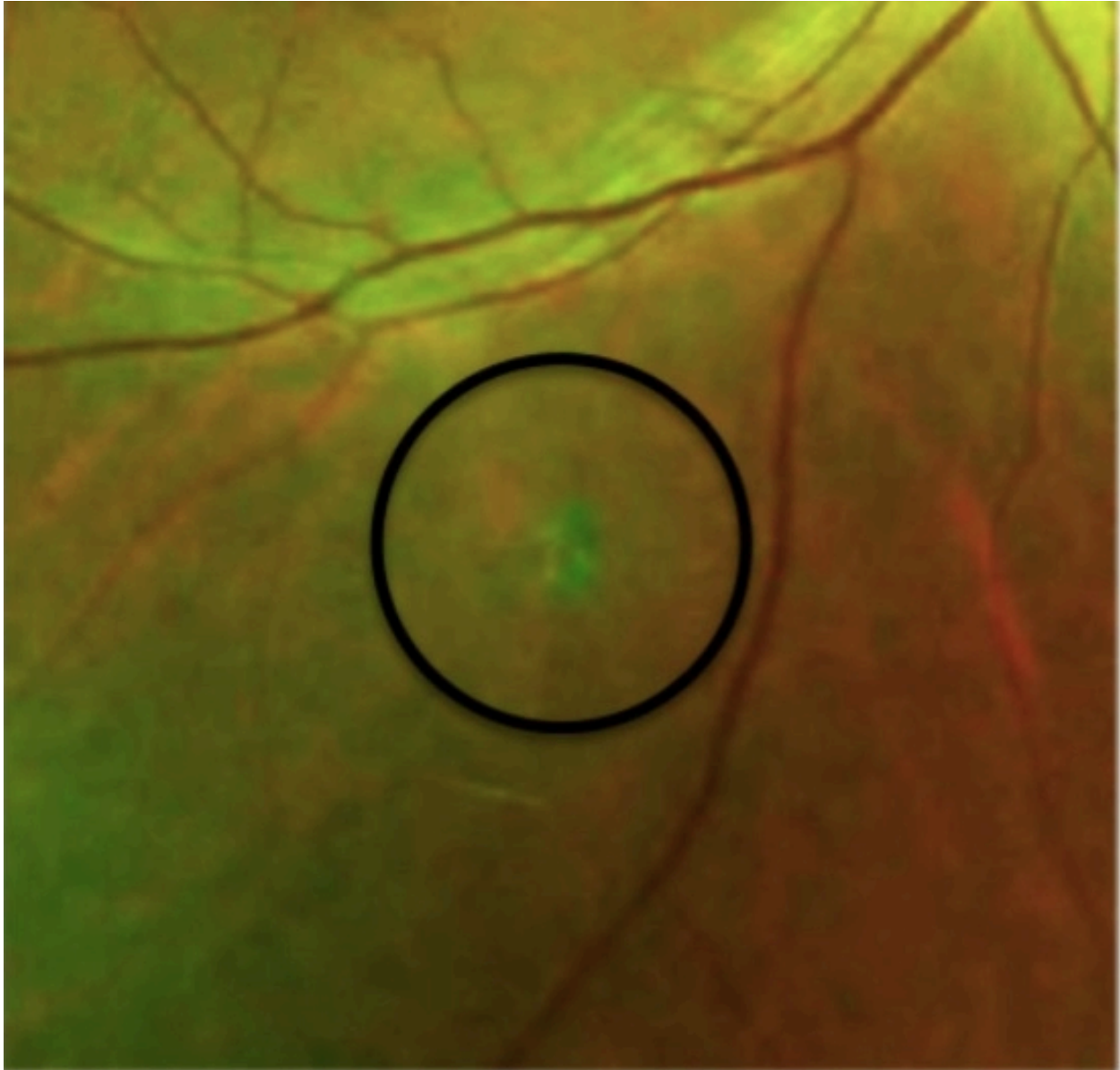
25. Are there any of Isolated Peripheral Type 6 lesions visible on the colour fundus image?

Mark only one oval.

- Yes, definitely
- Yes, questionably
- No

Type 7 Lesions

Type 7 lesions are small pigmentary disturbances. Their location is variable. They exhibit no surrounding retinal change.

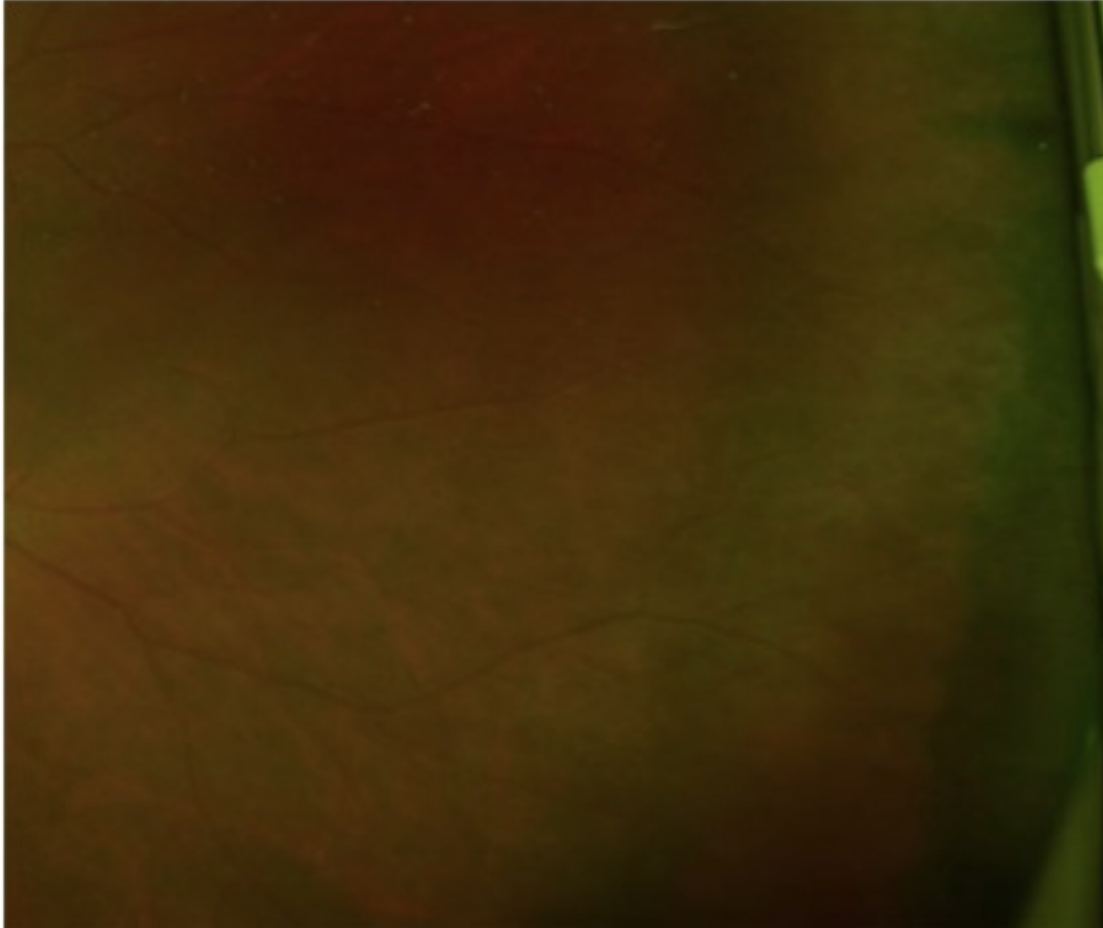


26. Are there any Type 7 lesions visible?
Mark only one oval.

- Yes, definitely
- Yes, questionably
- No

Type 9 Lesions

Type 9 lesions appear as pigmented bands along the retinal periphery. They are often only detected on the temporal aspect of the fundus image due to the greater temporal field of view obtained with an Optos Daytona image.



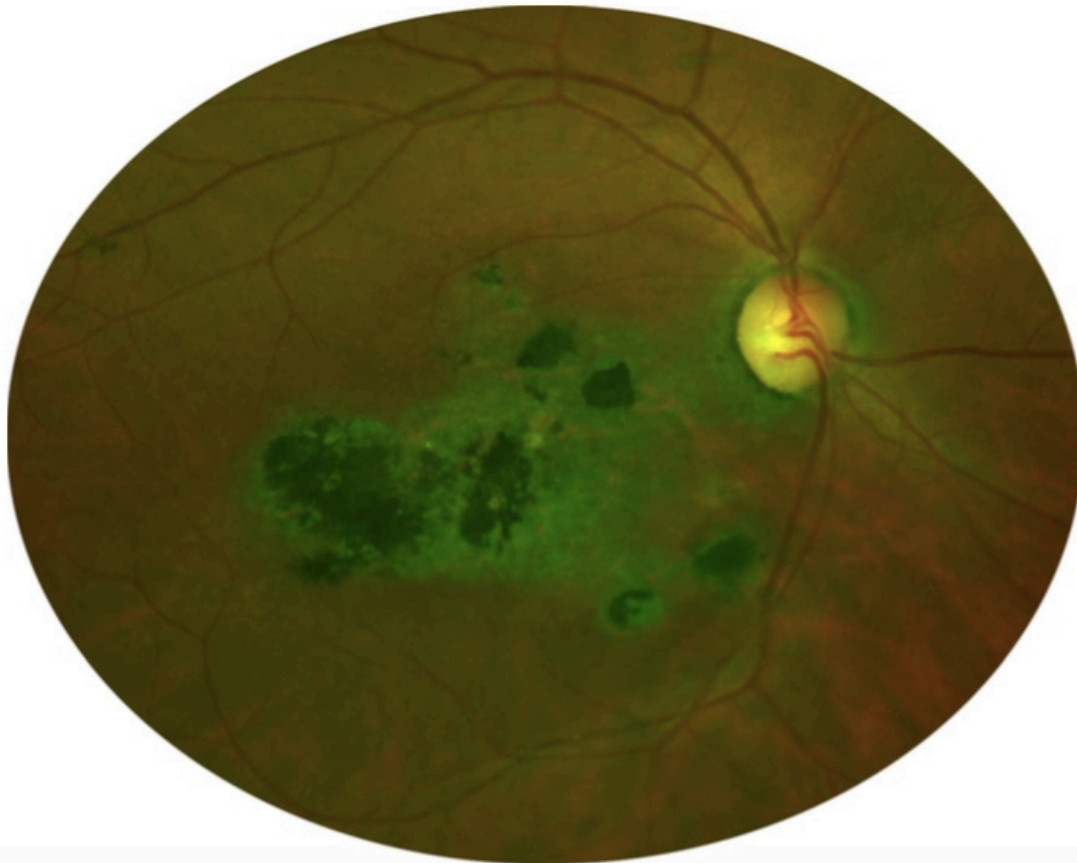
27. Are there any type 9 lesions?

Mark only one oval.

- Yes, definitely
- Yes, questionably
- No

Type 10 Lesions

These lesions seem to appear from the optic nerve. They always involve the macula. They appear diffuse grey in colour often with a halo around the optic nerve. They frequently have pigment sections but the area of hypo fluorescence extends further than the pigmented area. They appear to have a hyper fluorescent rim at the edge of the lesion on fundus autofluorescent imaging.

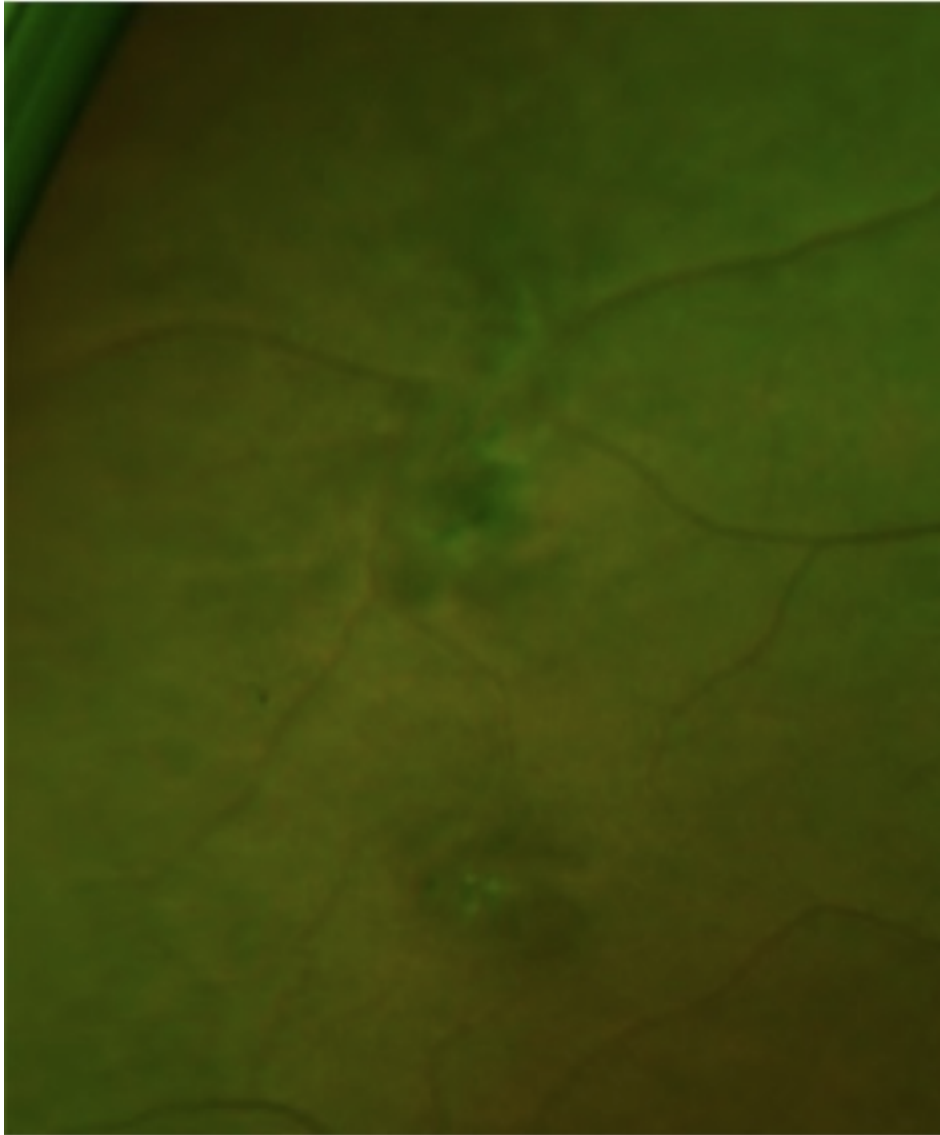


28. Are there any Type 10 lesions visible?

Mark only one oval.

- Yes, definitely
- Yes, questionably
- No

Peripheral Pigmentary Clumping



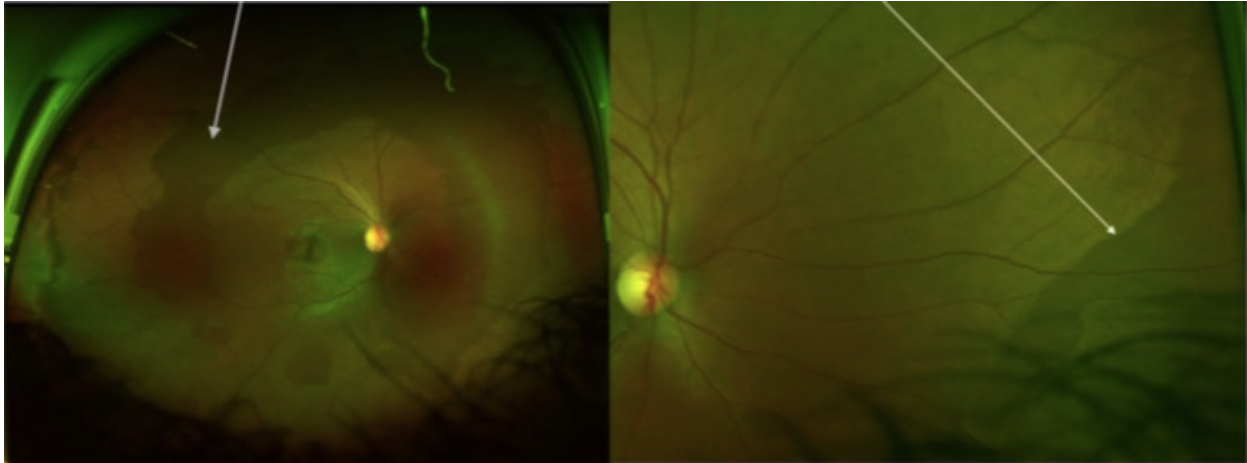
29. Are there any irregular pigmentary clumps within the peripheral that don't fit any of the lesion types? (see image above)

Mark only one oval.

- Yes, definite
- Yes, questionably
- No

Dark Without Pressure/Peripheral Dark Without Pressure

A well demarcated, geographic, dark retinal area. Often in the mid peripheral fundus but sometimes seen in the periphery. Retinal appearance on either side looks the same. There is no transitional zone.



30. Are there any areas of either of these signs?

Mark only one oval.

- Yes, definitely
- Yes, questionably
- No *Skip to question 32.*

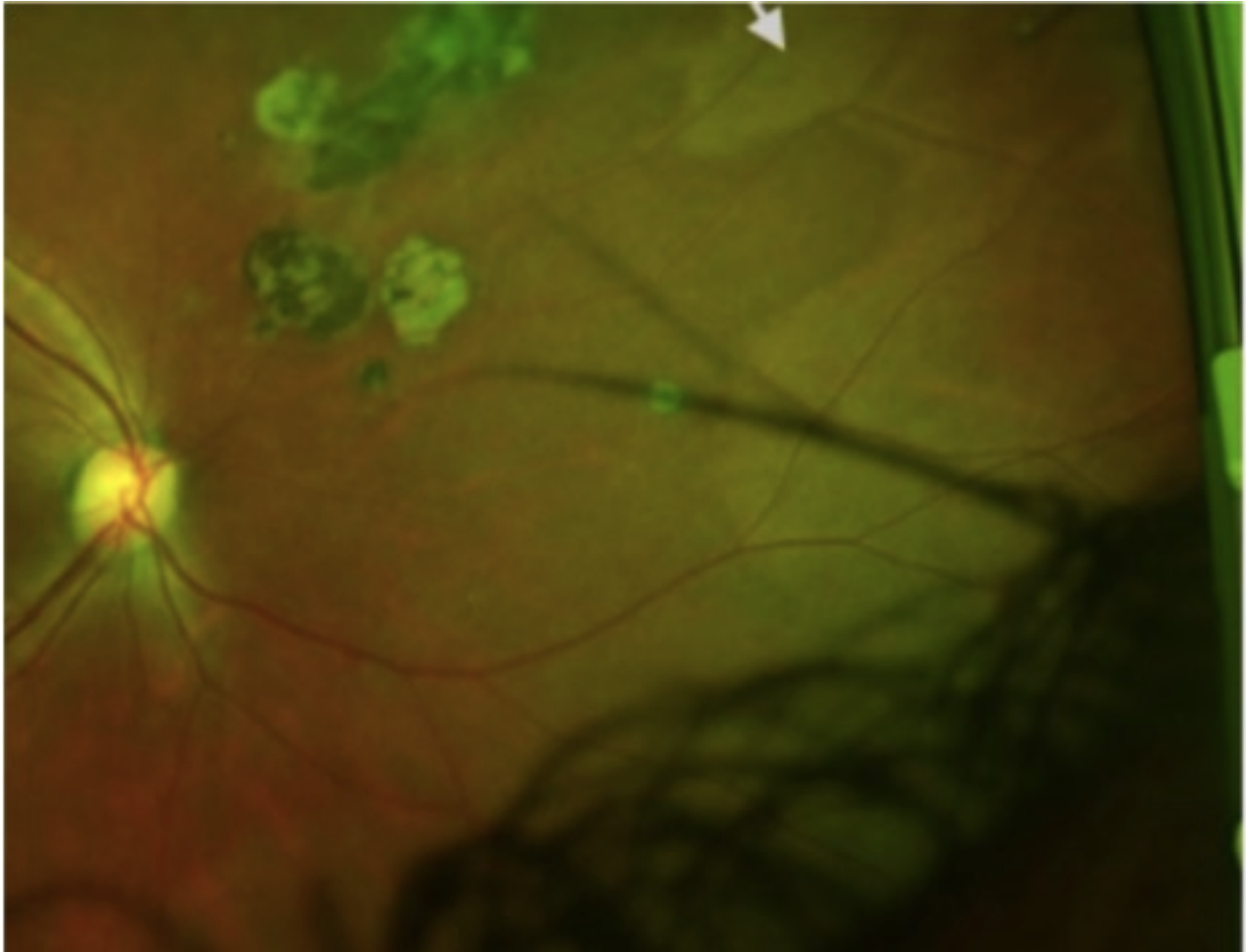
31. Does the Dark without pressure area contain any lesions?

Mark only one oval.

- Yes, definitely
- Yes, questionably
- No

Peripheral Whitening

A well demarcated peripheral LIGHTER zone of retina adjacent to what appears to be normal retina, which has a consistent colour with the rest of the posterior pole. There is no transitional zone.

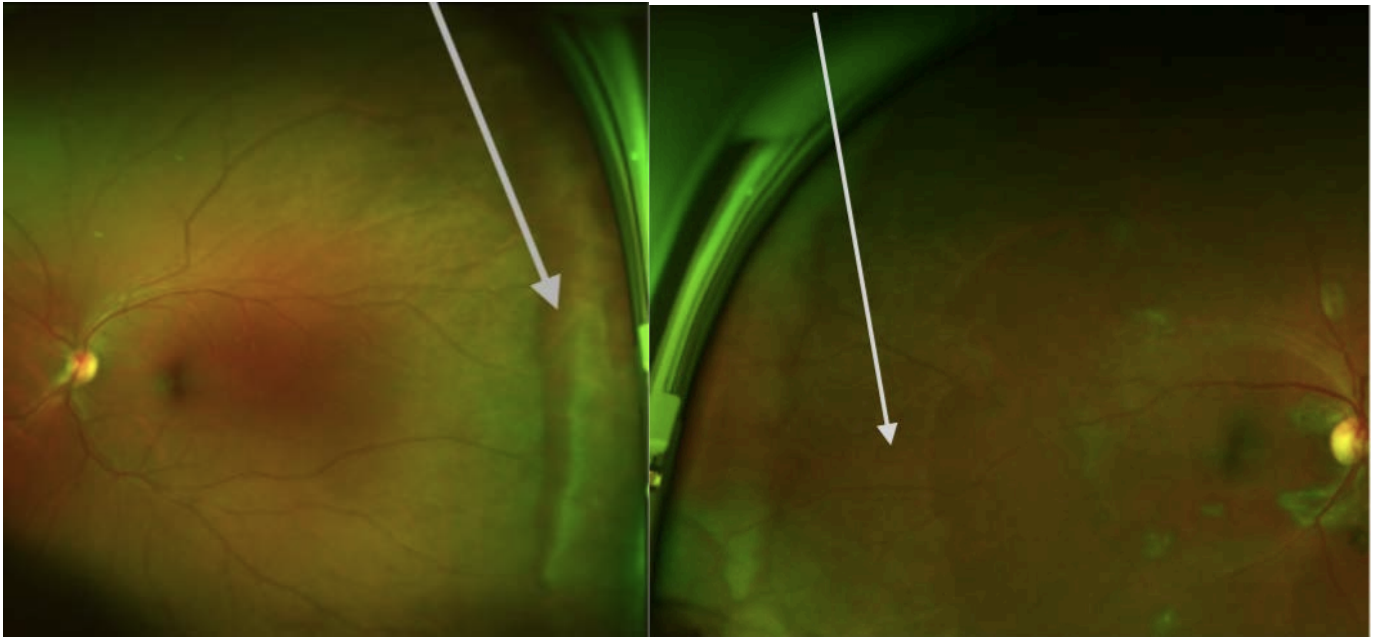


32. Are there any areas of peripheral whitening visible?
Mark only one oval.

- Yes, definitely
- Yes, questionably
- No *Skip to question 33.*

White Without Pressure

& associated Peripheral Retinal Glistening



Left Image - White without pressure (WWP)

Arrow points to the edge of the white without pressure lesion. The white area often increases in intensity before a darker border before a clear demarcation to normal retina.

Right Image - WWP associated retinal glistening.

In some eyes the area of retina adjacent to the WWP does not appear consistent with the posterior pole and instead has a patchy glistening appearance different to the solid appearance of peripheral whitening seen in the last question.

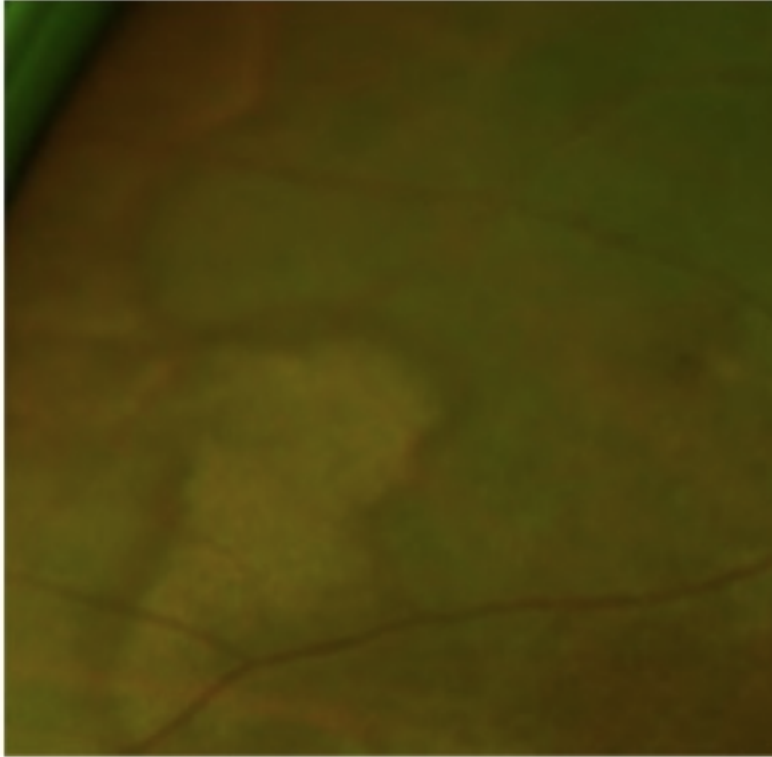
33. Are there any of the above lesions?

Mark only one oval per row.

	Yes, definitely	Yes, questionably	No
White without pressure	<input type="radio"/>	<input type="radio"/>	<input type="radio"/>
WWP associated retinal glistening	<input type="radio"/>	<input type="radio"/>	<input type="radio"/>

White with Dark Border

This differs to the previous types as there is an intermediate dark band between the peripheral white area and normal retina



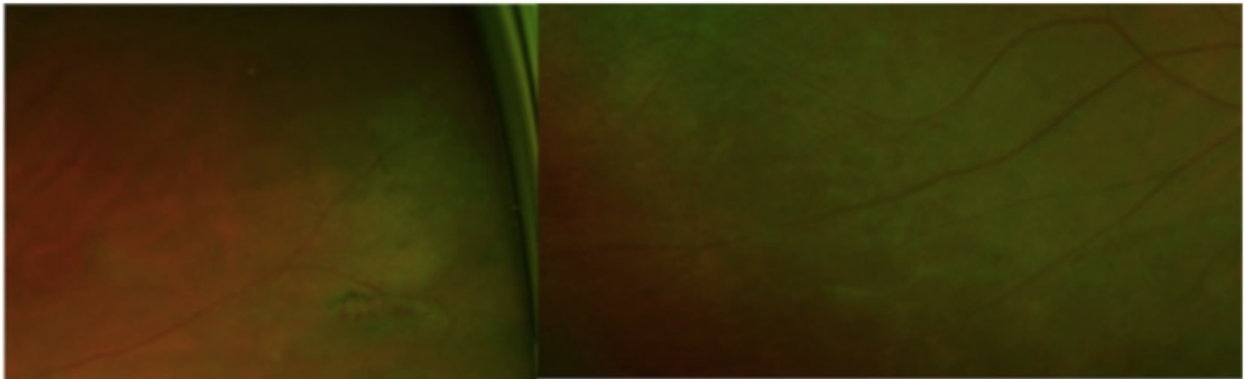
34. Are there any areas of White with Dark Border

Mark only one oval.

- Yes, definitely
- Yes, questionably
- No

Poorly demarcated peripheral whitening

These are ill defined peripheral changes with no clear border. They vary in intensity and are often patchy in nature

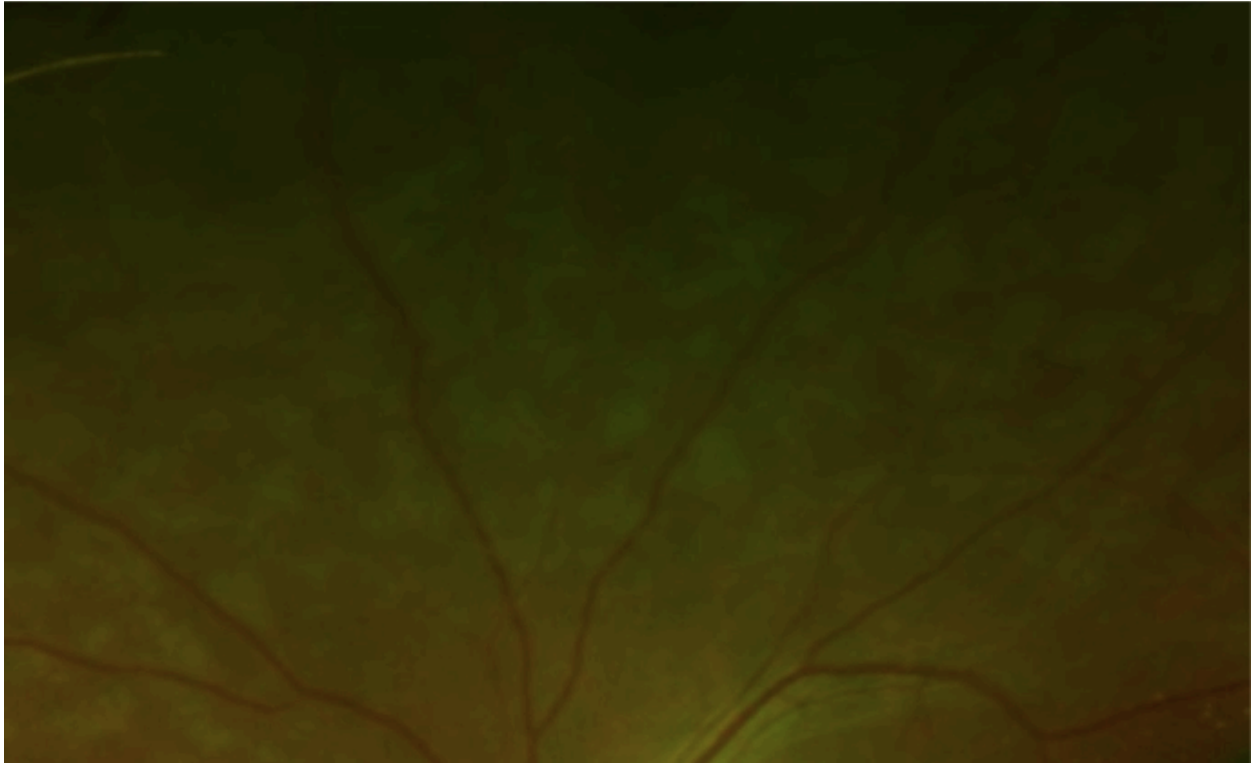


35. Are there any areas of poorly demarcated peripheral whitening?
Mark only one oval.

- Yes, definitely
- Yes, questionably
- No *Skip to question 36.*

Benign Flecked Retina

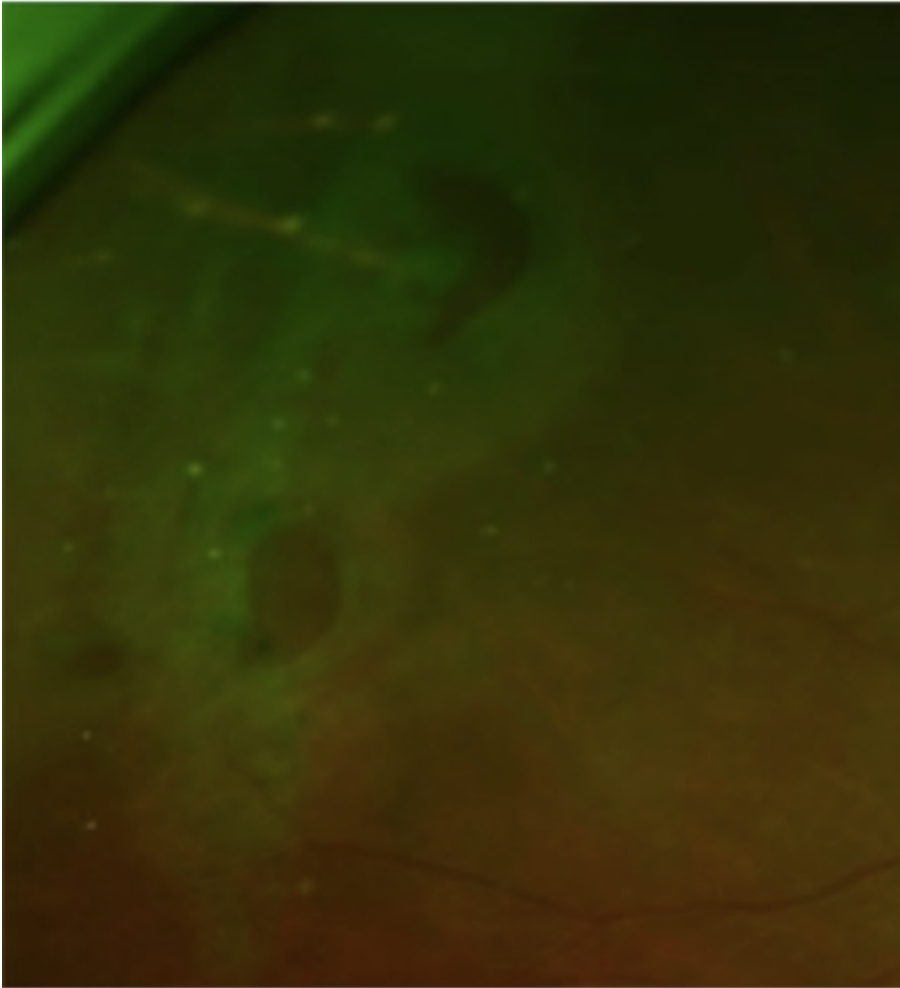
Previously described "benign flecked retina" appears as well demarcated islands of light zones in the mid peripheral retina



36. Are there any areas of the benign flecked retina in any areas of the retina?
Mark only one oval.

- Yes, definitely
- Yes, questionably
- No

Retinal Tears



37. Are there any retinal tears visible?

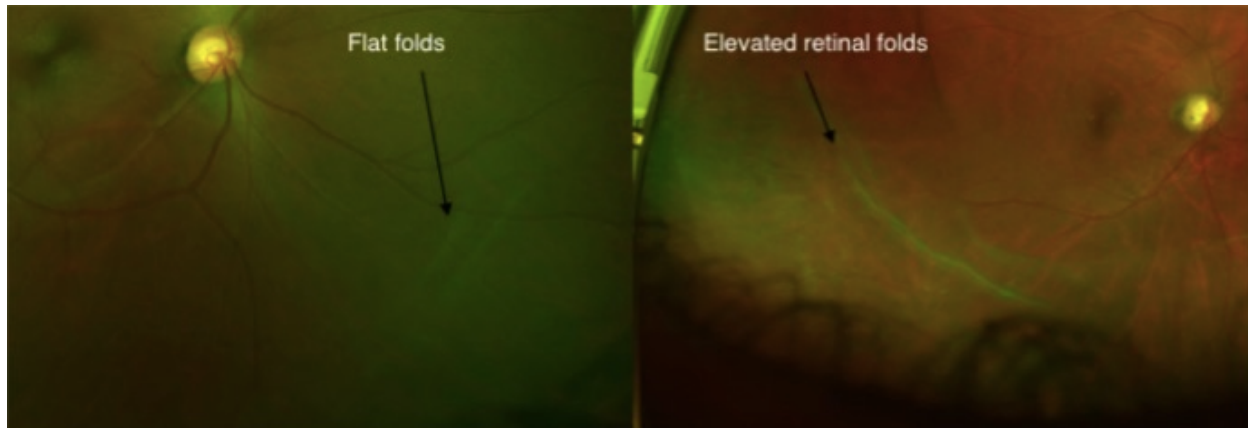
Mark only one oval.

- Yes, definitely
- Yes, questionably
- No *Skip to question 38.*

Retinal Detachments

Without slit lamp confirmation, the exact depth of the images below is difficult to assess. The image on the left may be in the vitreous whereas more prominent folds seen on the right may represent retinoschisis or detachment.

Retinal folds



38. Is there any evidence of the following?

Tick all that apply.

- Flat retinal fold appearance
- Appearance of Elevated Retinal Folds
- Definite retinal detachment
- Cheeky spots suggestive of retinoschisis
- No

Hemorrhages

39. Are there any retinal haemorrhages present?

Mark only one oval.

- Yes, definitely
- Yes, questionably
- No *Skip to question 40.*

Neovascularisation/Vasculitis

40. Is there any evidence of neovascularisation?

Tick all that apply.

- Definite New vessels at the disc**
- Definite New vessels elsewhere**
- Questionable New vessels elsewhere**
- None**

41. Is there any evidence of retinal vasculitis?

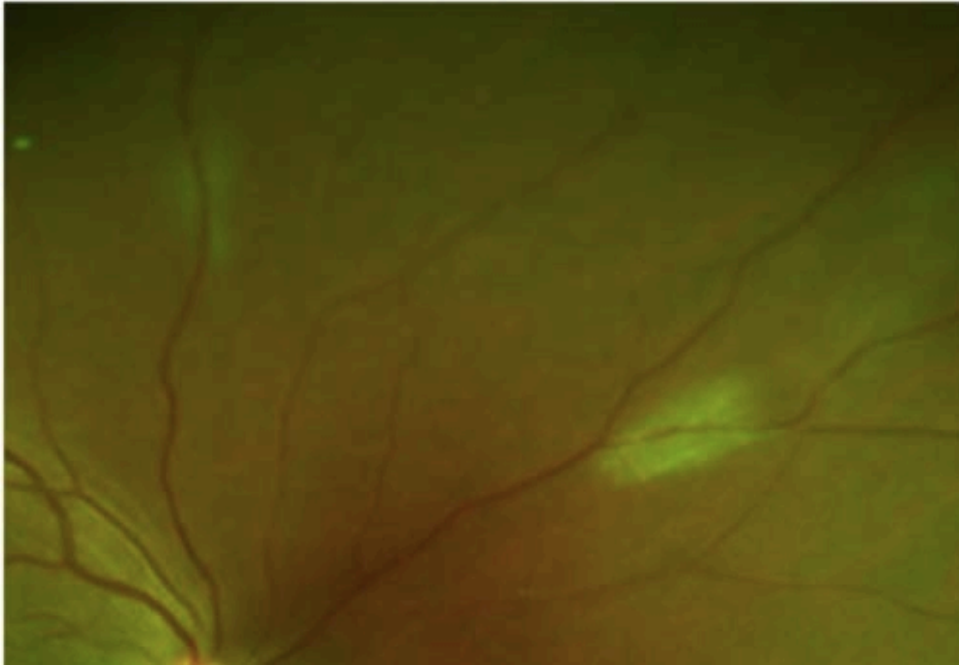
Mark only one oval.

- Yes, definitely**
- Yes, questionably**
- No**

Incidental retinal findings

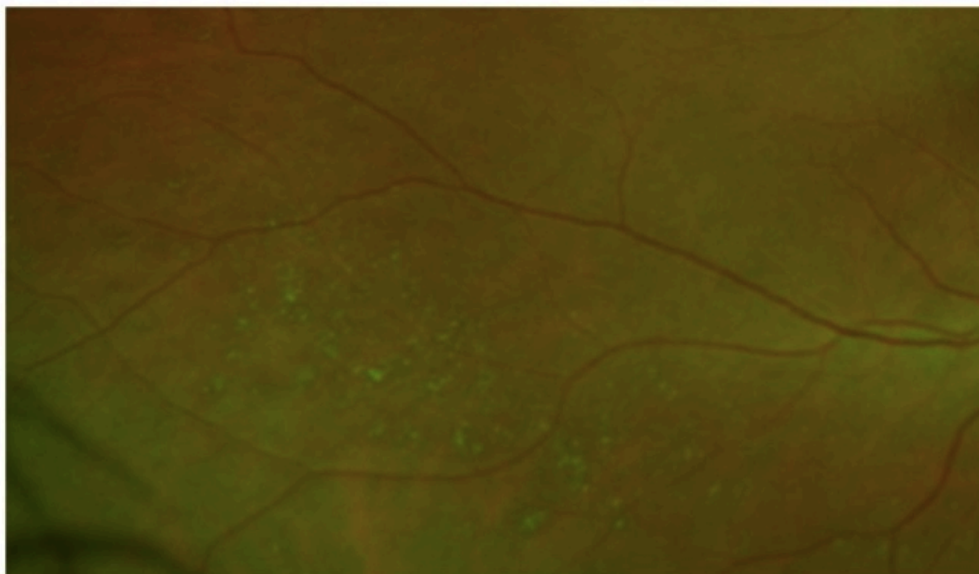
Retinal myelinated nerve fiber layer

Usually bright white streaks radiating to peripheral retina



Peripheral drusen

Small yellow dots usually outside the arcade. Some coalesce to form slightly larger areas



Snail Trail Degeneration

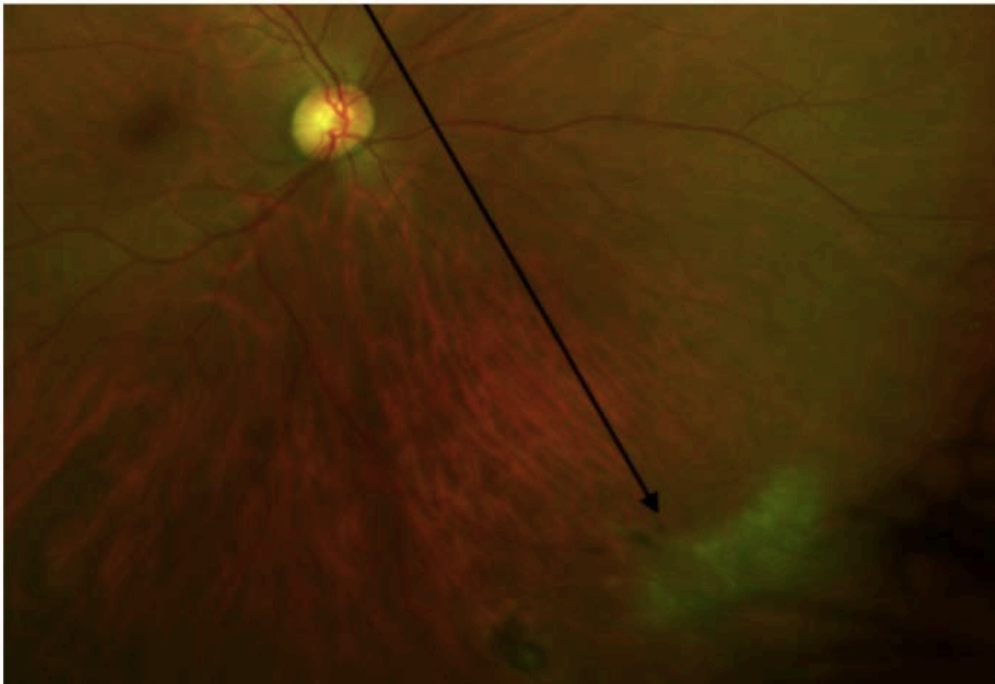
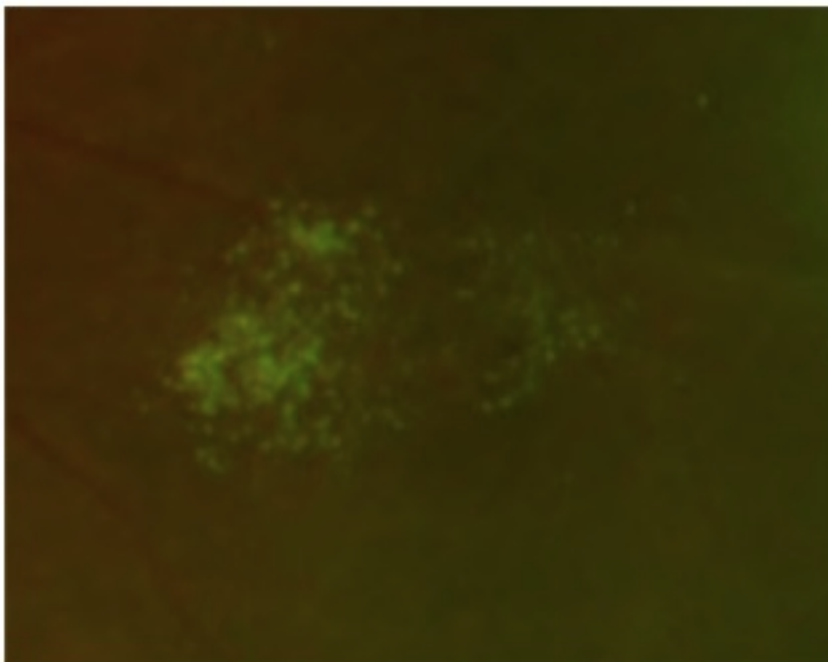


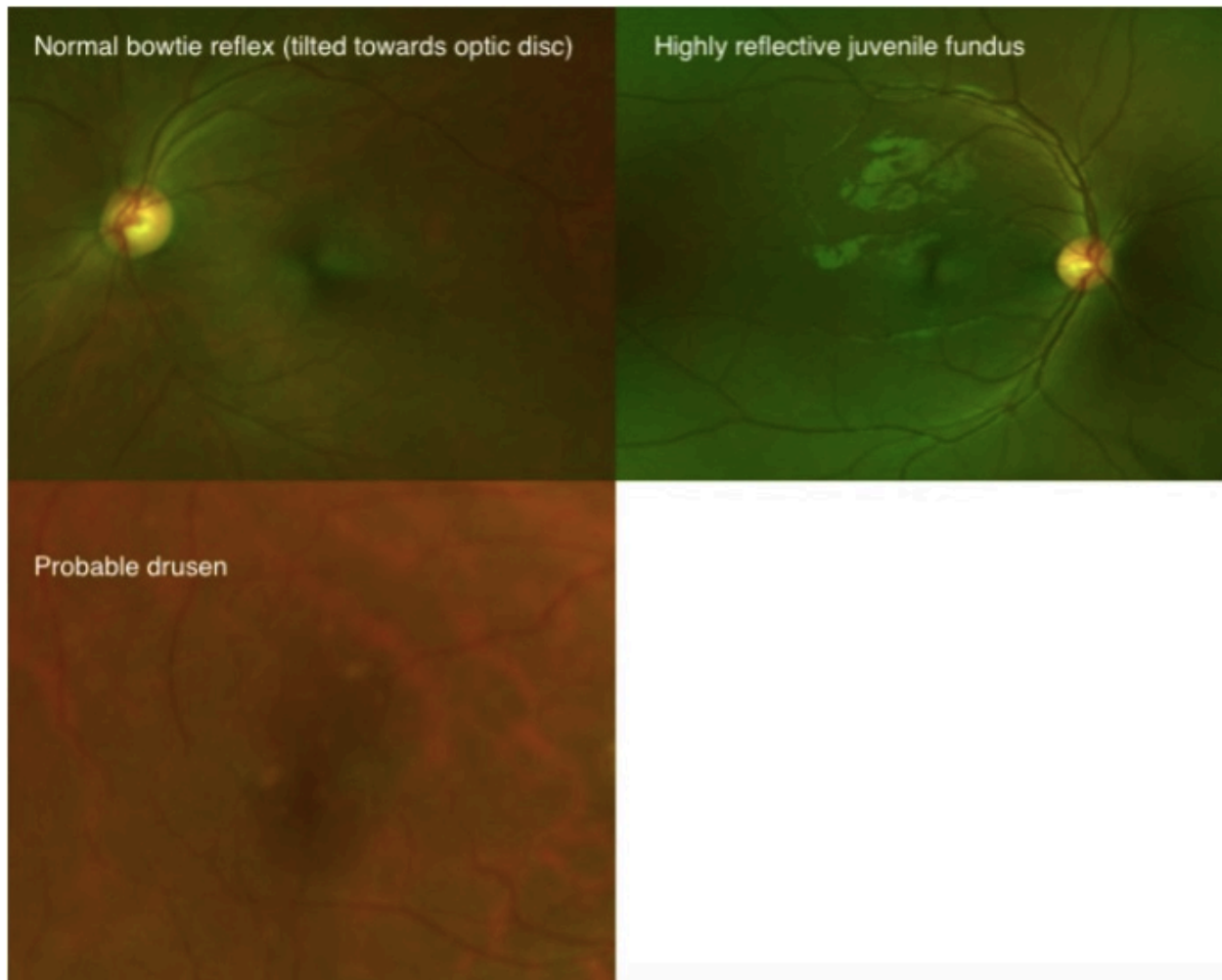
Image No 4



42. Are any of these findings visible on the retina?
Tick all that apply.

- Peripheral drusen**
- Myelinated nerve fibres**
- Snail Trail**
- Similar appearance to image 4**
- No**
- Other: _____**

Macula



43. Which of the following can be seen?
Tick all that apply.

- Normal bowtie reflex
- Reflective Juvenile fundus
- Macula hole
- Macula not visible
- Probable drusen

Optic Disc

44. Grade the cup to disc ratio

Mark only one oval.

- 0.1
- 0.2
- 0.3
- 0.4
- 0.5
- 0.6
- 0.7
- 0.8
- 0.9
- 1.0
- Unable to grade

45. Are there any optic disc abnormalities?

Mark only one oval.

- Yes, the disc is not normal
- No *Stop filling out this form.*

46. What are the optic disc abnormality?

Tick all that apply.

- Tilted disc
- Probable glaucoma
- Neovascularisation
- Vitreous adhesion to retinal scar
- Weiss ring
- Other: _____

Novel Retinal Lesion in Ebola Survivors, Sierra Leone, 2016

Technical Appendix 2

Anterior Chamber Sampling Protocol

All suitable candidates are assessed at the survivors' eye clinic 34 Military Hospital.

Signed or thumb print consent is sought and must be obtained with assistance of a local Krio interpreter. Where thumb print consent is obtained, a witness is also required.

The procedure will be undertaken in the survivors facility with the same inter sample cleaning protocol using 10% bleach as previous done in similar bodily fluid sample collection studies.

All aqueous taps will be taken by an experienced UK Ophthalmic specialist registrar with a Sierra Leone medical licence with the permission of the senior physician in charge of MH34, Col Prof Foday Sahr.

Method

1. Don full apron, facial protection and double gloves for personal protection.
2. Guttae (g) oxybutacaine hydrochloride 0.4% is instilled to anaesthetise the ocular surface.
3. A disposable speculum is inserted to maintain the palpebral aperture.
4. Topical g.povidone iodine 5% is instilled to decontaminate the ocular surface.
5. A 30 gauge needle on a 1ml syringe enters the cornea at the limbus in parallel to the iris.
6. A 0.1ml sample from the anterior chamber is withdrawn and the needle carefully retracted while external pressure is applied with a cotton tip.

7. The room and slit lamp will be deep cleaned and allowed to dry between patients.
8. A tear swab will be taken post procedure. Topical g.chloramphenicol 0.5% QDS is given for the next 4 days to reduce risk of any infection
9. The sample will be transported by means of a military ambulance to the Makeni lab facility for EBOV testing
10. The patient will be reviewed 30 minutes post procedure and one day following procedure for any adverse events.
11. Transportation costs will be refunded.

Potential Complications and Action Required

- Transient hypotony - recovery usually within 30 minutes post procedure.
- Sub-Conjunctival haemorrhage - Diffuse redness over conjunctiva due to a small degree of blood in between the conjunctiva and sclera. Reassure, self-resolving within 1-2 weeks. If bullous monitor for risk of dalen formation.
- Corneal abrasion – Identified with topical fluorescein. Very painful once topical anaesthetic worn off. Quick to heal within 1-2 days. Apply Chloramphenicol 1% ointment and pad overnight.
- Anterior chamber haemorrhage caused by inadvertent iris trauma - risk reduced by careful patient selection to ensure compliance with instruction and deep anterior chamber. Commence topical g.cyclopentolate 1% twice daily to reduce iris motility and g.prednisolone acetate 1% 4 times daily for 7 days. Monitor daily initially to ensure no associated elevation of intraocular pressure.
- Intraocular infection introduced by needle - risk reduced by pre-procedure povidone iodine 5% and post op chloramphenicol. Should endophthalmitis be suspected, this is an ophthalmic emergency. Follow the endophthalmitis treatment protocol.