

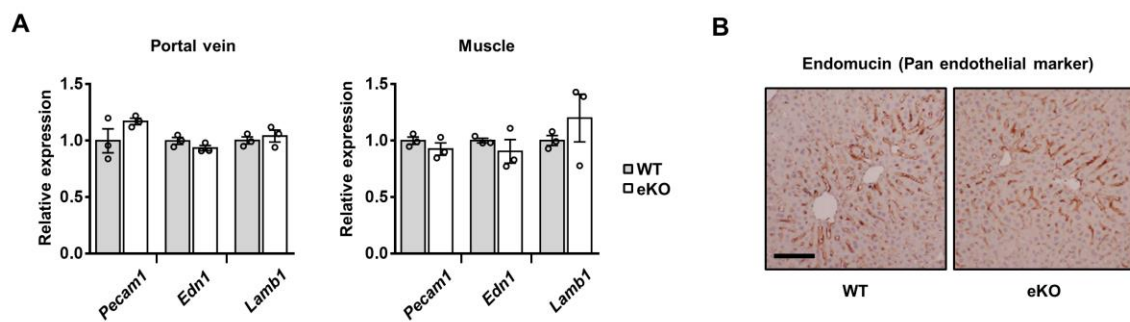
## Supplementary Figures and Table

### **Endothelial TAZ inhibits capillarization of liver sinusoidal endothelium and damage-induced liver fibrosis via nitric oxide production.**

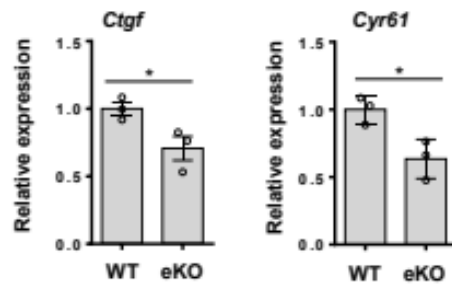
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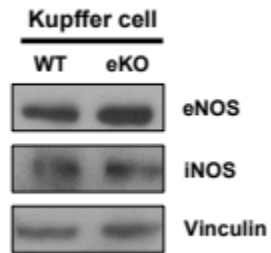
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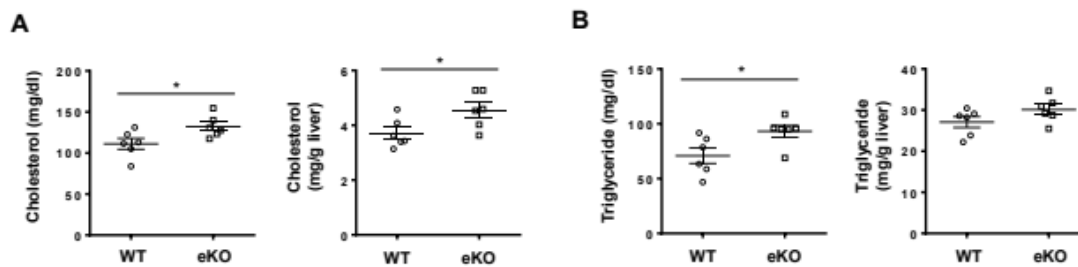
**Figure S1. A)** Descending aorta and tibialis anterior muscle were isolated from wild-type (WT) and endothelial TAZ-knockout (eKO) mice. After RNA extraction and cDNA synthesis, endothelial marker gene expression was assessed by qRT-PCR (n = 3). **B)** Liver endothelium of WT and eKO mice was visualized by immunostaining for the pan-endothelial marker, endomucin. Scale bar = 200  $\mu$ m. For panel **A**, data are presented as mean  $\pm$  SD.



**Figure S2.** Liver sinusoidal endothelial cells were isolated from wild-type (WT) and endothelial TAZ-knockout (eKO) mice. The relative expression of TAZ target genes was analyzed by quantitative real-time PCR (n = 3). Data are presented as mean ± SD. (\*P<0.05, as assessed using a one-tailed Student's t-test).



**Figure S3.** eNOS and iNOS levels in Kupffer cells. Kupffer cells were isolated from WT or eKO mice. The expression of eNOS, iNOS, and vinculin was analyzed by immunoblotting. Vinculin was used as a loading control.



**Figure S4.** **A)** Wild-type (WT) and endothelial TAZ-knockout (eKO) mice were fed with a 3,5-diethoxycarbonyl-1,4-dihydrocollidine (DDC) diet. Serum and liver tissue cholesterol levels were quantified using a cholesterol quantification kit (n = 6). **B)** Serum and liver tissue triglyceride levels of the mice in panel **A** were quantified using a triglyceride assay kit (n = 6). Data are presented as mean  $\pm$  SEM. (\*P<0.05, as assessed using a one-tailed Student's t-test).

**Table S1**

Primers used for gene expression analysis		
Gene	Direction	Sequence (5' → 3')
<i>Cd209b</i>	Forward	TGGGCTCCTGCTGATCATT
	Reverse	TTCCCTTGGGAGATGGGGAT
<i>Ehd3</i>	Forward	CGCCGTGCTTGAAAGTATCAG
	Reverse	ATAATTCGGTCCACCCGCTC
<i>Plvp1</i>	Forward	AGCACACTGCCTTCTCCTTG
	Reverse	AGCACACTGCCTTCTCCTTG
<i>Stab1</i>	Forward	TCACTGTCCCCACACTACTTT
	Reverse	TGTCGCAACGTTTAGACCGTA
<i>Stab2</i>	Forward	CACTATGTCGGGGATGGACG
	Reverse	GGGAGCGTAGGTGGAATACG
<i>Pecam1</i>	Forward	ACCGGGTGCTGTTCTATAAGG
	Reverse	TCACCTCGTACTCAATCGTGG
<i>Edn1</i>	Forward	CACCGTCCTCTTCGTTTTGC
	Reverse	GGCTCTGCACTCCATTCTCA
<i>Lamb1</i>	Forward	GAAAGGAAGACCCGAAGAAAA
	Reverse	CCATAGGGCTAGGACACCAAA
<i>Nos3</i>	Forward	CCTTCCGCTACCAGCCAG
	Reverse	CAGAGATCTTCACTGCATTGGCT
<i>Acta2</i>	Forward	GTTCAGTGGTGCCTCTGTCA
	Reverse	ACTGGGACGACATGGAAAAG
<i>Coll1a1</i>	Forward	TAGGCCATTGTGTATGCAGC
	Reverse	ACATGTTCACTTTGTGGACC
<i>Tgfb1</i>	Forward	GTGGAAATCAACGGGATCAG
	Reverse	ACTTCCAACCCAGGTCCTTC
<i>Timp1</i>	Forward	AGGTGGTCTCGTTGATTTCT
	Reverse	GTAAGGCCTGTAGCTGTGCC
<i>Fasn</i>	Forward	AGGGGTCGACCTGGTCCTCA
	Reverse	GCCATGCCCAGAGGGTGGTT
<i>Pparg</i>	Forward	ATGGGTGAAACTCTGGGAGA
	Reverse	CTTGTGAAGTGCTCAGC
<i>Scd1</i>	Forward	CGTCTGGAGGAACATCATTCT
	Reverse	CAGAGCGCTGGTCATGTAGT

<i>Srebp1c</i>	Forward	GTACCTGCGGGACAGCTTAG
	Reverse	TCAGGTCATGTTGGAAACCA
<i>Ctgf</i>	Forward	CGACTGGAAGACACATTTGG
	Reverse	CAGGTCTTAGAACAGGCG
<i>Cyr61</i>	Forward	GAGTGGGTTTGTGATGAAGAC
	Reverse	CTTCAGTGAGCTGCCTTTTCC
<i>Gapdh</i>	Forward	GCTTGTCATCAACGGGAAG
	Reverse	GATGTTAGTGGGGTCTCG
Primers used for chromatin immunoprecipitation		
Target	Direction	Sequence (5' → 3')
<i>Nos3</i> promoter	Forward	GGTCAGCGGGCATGAAG
	Reverse	AGCAGAGTCCTGGCCTT
Primers used for luciferase reporter construction		
Target	Direction	Sequence (5' → 3')
<i>Nos3</i> promoter	Forward	AAAAAACTCGAGGTGGGTTTCAGGAAATTGAGATGA
	Reverse	AAAAAAAAGCTTAGCAGAGTCCTGGCCTT