

Review

Human Cancer Classification: A Systems Biology- Based Model Integrating Morphology, Cancer Stem Cells, Proteomics, and Genomics

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Abstract

Human cancer classification is currently based on the idea of cell of origin, light and electron microscopic attributes of the cancer. What is not yet integrated into cancer classification are the functional attributes of these cancer cells. Recent innovative techniques in biology have provided a wealth of information on the genomic, transcriptomic and proteomic changes in cancer cells. The emergence of the concept of cancer stem cells needs to be included in a classification model to capture the known attributes of cancer stem cells and their potential contribution to treatment response, and metastases. The integrated model of cancer classification presented here incorporates all morphology, cancer stem cell contributions, genetic, and functional attributes of cancer. Integrated cancer classification models could eliminate the unclassifiable cancers as used in current classifications. Future cancer treatment may be advanced by using an integrated model of cancer classification.

Key words: Human Cancer, Cancer Stem cells, Multiple Omics, Integrated Classification

Introduction

Human cancers occur worldwide. In 2008, 12.7 million new cancers and 7.6 million cancers were recorded; incidence and mortality rates varied with regions and levels of income around the world (1). These results require refocusing on all attributes of cancer.

Differentiating features of malignant and benign lesion are well established; these include rapid growth, increased cell turn-over, invasive growth, metastases, vascular or lymphatic channel invasion for malignant lesions. There are many exceptions to these attributes of cancer. There are overlaps between benign and malignant lesions. Benign (non-malignant) tumors do show chromosome aberrations; uveal melanomas and blue nevi share mutations in G protein (2). A good example is the recent attempt among dermatopathologists to segregate some melanocytic lesions as atypical melanocytic proliferations with low malignant potential

(MELTUMP) (3). Sometimes a cancer at given site/organ is classified as containing two cell types; for example, pancreatic cancer with neuroendocrine and acinar/ductal components (4). What model can accommodate unclassifiable cancers in a specific location? Cancer classification schemes always reserve a group as unclassifiable. How can this group be eliminated?

The last two decades have witnessed the surge in molecular profiling (5, 6) and has already expanded into predictive and diagnostic molecular classification of cancers (7, 8). As in the diagnosis of cancers, current molecular classification schemes are still dependent on morphologic variables. These classifications schemes use cell of origin as seen by light and electron microscopy. Inherently, all organs can generate multiple cancer types as multiple cell types exist in these organs- "the holy-grail of all subspecialties". Furthermore, cancer subtypes are generated under the

banner of a single, specific cell type of origin concept. Take the example of the common Basal cell carcinoma- it has variants and subtypes such as nodular, superficial, adenoid, morpheaform, infiltrative, keratotic, pigmented, basosquamous, clear cell, granular, eccrine, apocrine, fibroepitheliomatous, adamantoid, and basosebaceous (9). Do these entities have unique biological features or simply morphological variants of interest only to the diagnostic pathologists? Will the "cancer stem cell origin concept" cure this malady?

The current move to genomics {(gene and transcripts, kinomes, microRNAs, single nucleotide polymorphisms (SNPs), gene copy number variation (CNVs) and proteomics (antibody microarray and mass spectrometry)) brings change to the diagnostic information needed for treatment. Along with the genomic profiling, are efforts at targeted and gene therapy. Because of accumulated experience, in diagnosis, classifications and treatment of cancer that depends on morphology, the shift to genomic methods should be comprehensive and adequate for day-to-day clinical use. While future cancer classification schemes or models may not require morphological attributes, current dependencies on morphological phenotype requires its inclusion (10-12). Morphologic cancer phenotyping does not need to hide the compendium of genetic alterations, interactions with environment and alterations in transcriptional and protein interaction networks that are present in all cancers (11, 13).

Hallmarks of Cancer Cell

Hanahan and Weinberg (2000) (14, 15) listed the seven attributes of cancer; 1) Self sufficiency in growth signals, 2) Insensitivity to anti-growth signals, 3) Evading apoptosis, 4) Limitless replicative potential, telomerase and telomeres 5) Sustained angiogenesis, 6) Tissue invasion and metastasis, and 7) Genome instability. All seven attributes have received great attention in the past decade. Growth and anti-growth signaling are really complex (13). Protein-protein interaction and signaling networks, growth signaling pathways, the role of ubiquitination and protein degradation, and dysfunctional protein networks (16-18) and interactions are complex, described as hubs, modules and motifs (13). Information on cancer cell death and provocation by drugs and irradiation now requires all cell death types to be considered- apoptosis, necrosis, autophagy (19, 20). We now must include the pivotal role of microRNAs (21, 22), and methylation patterns (23). For example, microRNA-185 suppress cancer growth by interfering with Six1; when absent in cancers leads to increase growth and progression (24). Recent efforts have un-

covered the role of transposons in the induction of cancer in mouse models; the studies are generating previously unknown cancer related genes (25). Class II (DNA transposons) and class I retrotransposons contribute to DNA instability (26). Cancer cells use aerobic glycolysis to meet energy needs (Warburg effect) and presumed to be a response to hypoxia and tumor micro-environment; changes in metabolic needs of cancer cells such as need for glutamine and activation of hypoxia-inducible-factor (HIF) are interconnected to oncogene activation (27-29). These interacting functionalities of cancer cells impact prognostic and predictive models based on one or two functional attributes of cancer (30).

Origin of Cancer Cells -The Cancer Stem Cell Model

The traditional model of cancers envisaged a "normal cell" transformed to "atypical or dysplastic" cell with progression into invasive of malignant cell. This is the model that only assumes stochastic generation of cells capable of the behavior of metastasis and progression and cellular heterogeneity of cancers. The stochastic model is used to explain heterogeneity in cancers such as in prostate cancer (Fig 1). The stochastic model will have to assume that all genetic aberrations conferring advantages to the cancer cells "must be maintained in all subsequent cells as growth and proliferation continues and some maturation occurs". As cancers can also undergo senescence, apoptosis, autophagy and necrosis, the stochastic model must account for these changes (31, 32). Cancer senescence occurs via telomere shortening, oxidative stress, and oncogene activation, that can impair cancer progression (33-35). The stochastic model has to account for local recurrence and metastasis after long post-treatment intervals. Cell of origin models cannot exclude all interactions between cancer cells and any influence from the stroma (36, 37). Recent computational stochastic models, in part based on the hallmarks of cancer, suggest that onset of cancers depend on the first two (2) mutations and early-onset and late-onset cancer initiate these mutations at different times (38). Cancer initiation via DNA damage response and repair, induction of senescence and p53 mutation (39), the generation of driver mutations can be accommodated in stochastic and cancer stem cell models. Driver mutations in cancer initiation are suggested to accumulate over time. Recent detailed studies of human cancers and cancer cell lines show extensive and highly localized mutations following DNA damage and repair response; 2-3 % of human cancers develop these single chromosome mutations labeled as chrothripsis and are suggested as the gene-

sis of driver mutations generated (40). There are no unique driver mutations for the stochastic model.

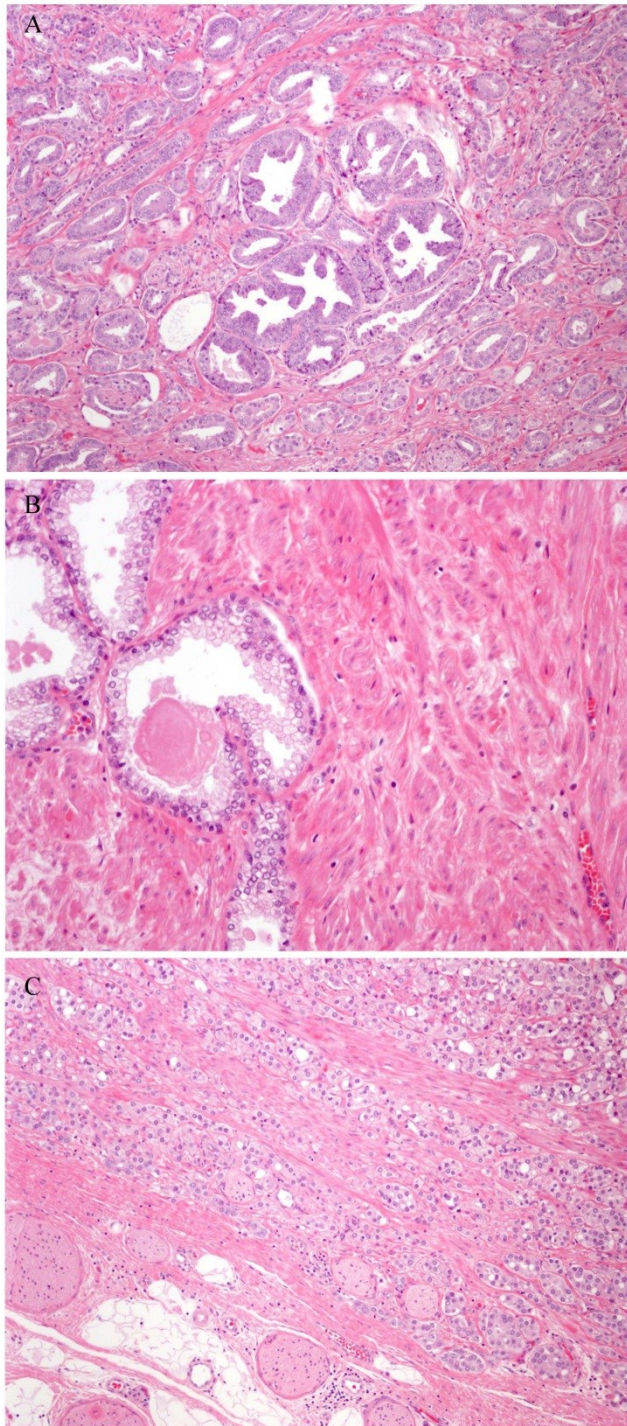


Figure 1. Patterns and heterogeneity in Cancerous Prostate Tissue (Hematoxylin and eosin stain). A. Normal prostate showing luminal and basal cells. B. Prostate cancer with definable glandular pattern, usually part of Gleason pattern 3. C. Prostate cancer with aggregation, clustering or individual infiltrating cells as described for Gleason 4.

Cancers of certain presumed cell types, synovial cells, occur in locations without defined synovium or other related cell type. Clear cell sarcoma is seen in deep soft tissues and is a copy of all features of melanoma except for genetic aberrations. How do we explain these cancers on the cell of origin concept? Prostate cancer cell of origin thought to be luminal cells, are now shown to be derived from basal cells with the attributes of androgen-independence (41).

The cancer stem cell model has been used to explain cancer cell origin, initiation, progression and metastasis (42-44) (Fig 2). Cancer stem cells as origin of cancers has attributes of hierarchical organization, may be under-estimated and assumed to be a minor population (45, 46). As in their resident or embryonic stem cell counterparts, there are known regulators, such as p53 and WNT signaling pathways. Cancer stem cells show c-Myc transcription profile, similar to embryonic stem cells (47). These cancer stem cells, initially described in breast cancers (48), are now described in liver, ovarian, prostate, head and neck, colon and brain cancers, melanomas (49-54). The cancer stem cells have their specific microenvironments to allow for their specific functions; epigenetic modifications may make cancer stem cells not reliant on its specific niche (42). Cancer stem cells are usually projected as a minor population of all cancer cells (46, 55). The number of identified cancer initiating stem cells may be affected by the background of animals used in xenotransplantation; in a mouse xenograft model of melanoma, 25% of cancer initiating cells could be found (50).

The functional attributes of cancer stem cells such as (i) evasion of cell death, i.e apoptosis, (ii) telomere activation, (iii) colony formation, tumor initiation and differentiation are suited to their role in human cancer (45). The contribution to recurrence, metastasis and treatment, especially radiotherapy-resistance, is now better appreciated (56, 57). Cancer stem cell markers for several human cancers are listed in Table 1. Cancer stem cells markers have functional attributes such as adhesion, cell invasion (CD44) and interactions with GLI1 and focal adhesion kinase (FAK) (CD24) (58).

Table 1. Biomarkers for Human Cancer Stem Cells

Cancer Type	Markers
Breast	CD44+CD24 ₋ /low
Ovary	CD133+/CD44+, CD117, Oct4, STELLAR, Nanog and ABCG2/BCRP1
Lung	Cd133+(ABCG2, Oct4, ESA)
Brain	CD133+
Colon	CD133+ CD44+ EpCam+, CD166+ (CD29+, CD24+)
Pancreas	CD44+EpCam+ CD24+

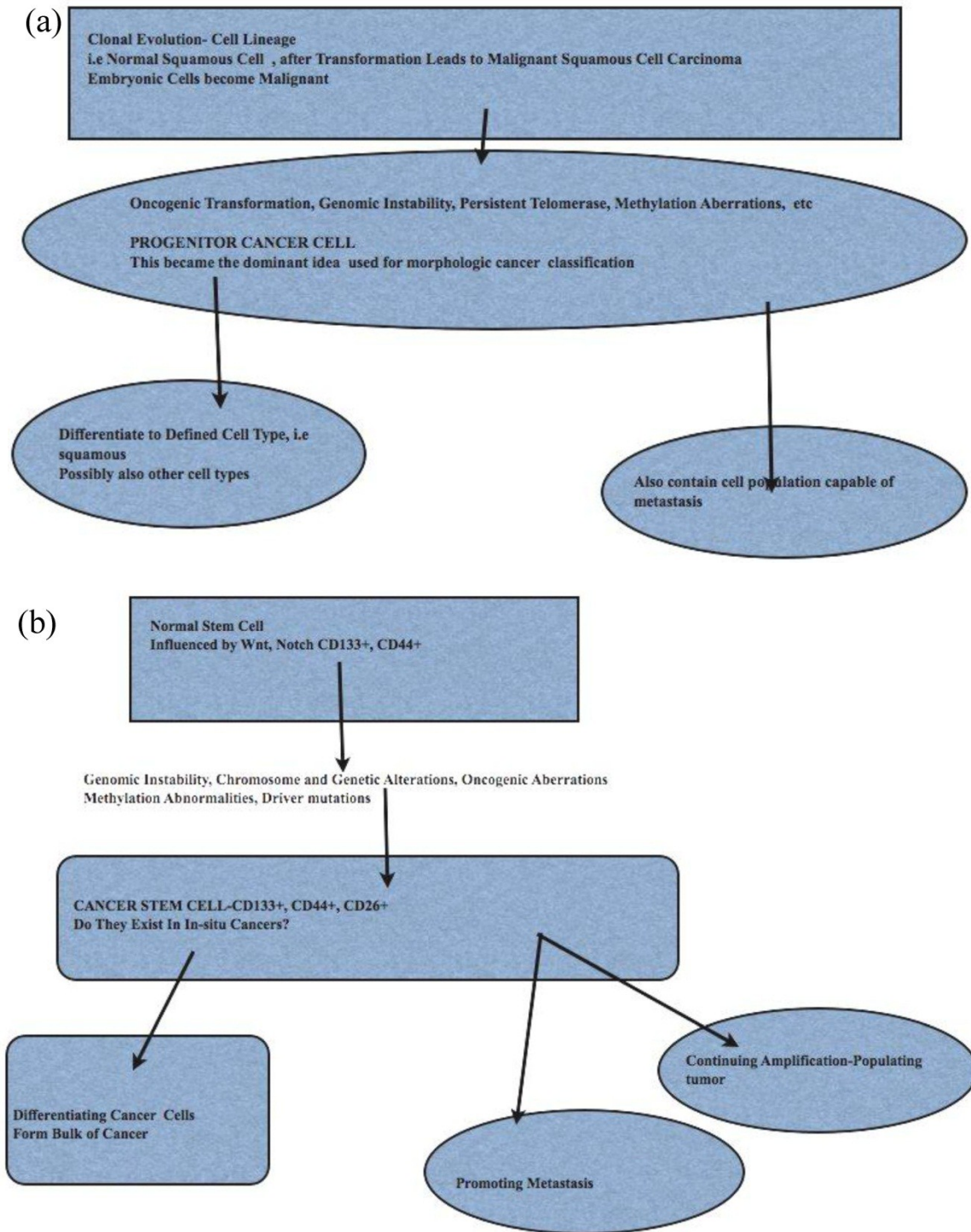


Figure 2 Comparison of Stem Cell and Lineage/Clonal Evolution Models of Cancer Cell Origin. (a) The lineage/clonal evolution model is used in morphological classification of cancer. One cell type gives rise to one cancer type. Squamous epithelium gives rise to squamous cell carcinoma. Pathologists do encounter squamous cell carcinoma in the urothelium of the urinary bladder. How we explain this is by “metaplasia of urothelium to squamous epithelium” and perhaps then to squamous cell carcinoma. An easier explanation will be cancer stem cell model as these have the capacity to become any cell type. (b) Cancer stem cells, with their inherent functional capacities including reduced cell death, are of interest when cancers are treated by irradiation or chemotherapy. Human cancers, examined in detail and extensively, do contain heterogeneous cell types; lung cancers are a good example. This leads to difficulties in some classification schemes depending on lineage.

Cancer Metastases and Cancer Stem Cell Model

Cancer metastasis occurs either through progressive acquisition of metastatic potential or the traits are acquired early during cancer initiation; this implies that metastases occurs early or late in cancer (59-61). The acquired traits include survival and evasion of cell death, dormancy, migration, immune escape, and ability to seed, home on targets (62). Support for late evolution of metastasis was described in pancreatic cancer using genome sequencing ; this study indicated clones with metastatic capacity evolve late (5years), and are present in the primary tumor (63, 64). Breast cancer cells reaching the bone marrow share stem cell features and markers CD24 and CD44 as determined by double immunohistochemistry in bone marrow; these stem-like cells ranged from 33-100% in metastases (65). CD26+CD24+ positive cancer cells found in colon cancers, not initiating CD133+CD26+CD44 positive cells, define occurrence of metastases (66). Some have separated cancer stem cells from pancreatic cancers capable of initiating metastases (67).

Personalized Genomic Medicine and Cancer Classification

How will the push for personalized medicine affect the present morphology based and the future trend of molecular cancer classification? Personalized medicine, as envisioned will require individual cancer genomics and proteomics for maximum benefit of targeted treatment for the individual; the implications of genomic cancer medicine should encourage use of integrated cancer classification models (68-73).

Integrated Model of Cancer Classification

The model envisaged (Fig 3) takes into account all elements of a cancer. We now can provide morphologic classes and subtypes, extract proteomic and gene profiles and gene copy number variations including cytogenetic and array comparative genomics. An added feature is that the functions of proteins and signaling pathways can be derived from gene expression and proteomics. This means that altered or dysfunctional pathways can be supported by adding cytogenetic or CNV data. There are emerging integrated models that use both genomics, exon resequencing, and proteomics in cancer analysis (74-76). Recent prostate cancer survival and post-surgical recurrence used modeling based on some aspects of the integrated classification model including mutation patterns, CNVs, targeted signaling pathway deregula-

tion, miRNA and cDNA profiling; new oncogenes, and CNV-based disease risk profiles over above morphology grades were found (77).

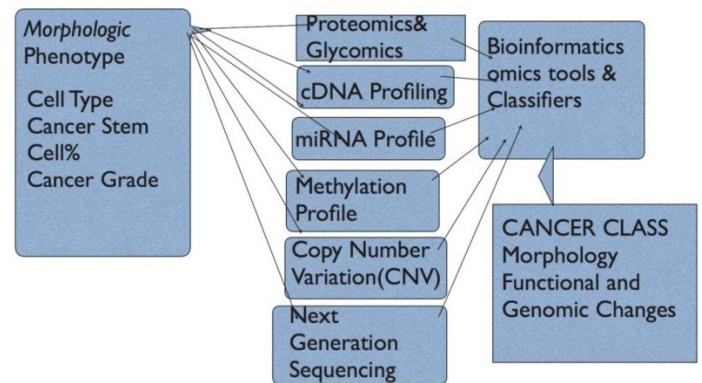


Figure 3 Figure Model of Cancer Classification. In this model, the Phenotype is represented by Morphological Characteristics/and subtypes; Proteomic profile can be derived from high-throughput tissue microarrays and immunohistochemistry plus automated computer-assisted quantitation with normalized intensities, protein microarrays and mass spectrometry; array comparative genomics for Copy Number Variation(CNV) and chromosomal aberrations, Genomic profiling using cDNA microarrays, and finally microRNA profiling. This provides protein profile and cDNA profiles for Gene Ontology and functional annotation. Signally pathways active or repressed can be derived. The CNV and microRNA data provide information to explain active oncogene induced pathways and miRNA targeted pathways that impact proliferation, cell survival, metastasis etc.

1) Proteomics: Normal and cancer tissues can be used for comparative proteomics. The methods available are mass spectrometry, protein and antibody microarray and tissue microarray. In mass spectrometry, tissue and cell protein content is quantitatively determined by analyzing peptide content and via bioinformatics the protein content, protein types and classes, protein interaction networks and via gene ontology functional protein mapping (78-80). Tissue microarray uses tissue cores for immunohistochemistry and indirect analysis of protein levels (81-83). In antibody and protein-antigen arrays, protein or antibody spotted on an array is probed with fluorophore-labelled protein or antibody and analyzed like cDNA microarrays (84-87). Flow cytometry can be used to catalogue both surface membrane, cytoplasmic and nuclear proteins in cells. Mass spectrometry, tissue microarray can be used to validate other profiling methods. Mass spectrometry is quantitative and can estimate both modified and unmodi-

fied proteins; 2-dimensional gel electrophoresis and protein isolation can be followed by mass spectrometry (78, 80, 88, 89). Glycomics and glycan profiling can add to the protein profile using both mass spectrometry and lectin microarrays to generate aberrant cell migration (90, 91).

2) cDNA Profiling and Transcriptomics; cDNA microarray profiling is the most common highthroughput method for determining expression levels of mRNA in cells and tissues (5). Isolated mRNA is transformed to cDNA, which is used to probe optimally designed oligonucleotide array that helped to generate new classes of breast cancer, leukemias and lymphomas. Using transcriptomics for clinical cancer treatment is an area of intense research (translational research) (12, 68, 72). Direct RNA sequencing overcomes drawbacks of cDNA based methods, detect low quantities of mRNA, detect chimeric transcripts, can use RNA from fixed tissues, and uses sequencing-by-synthesis (92).

3). Copy Number variation (CNV): Normal and cancer cells show variations in genes. The most common variation is single nucleotide polymorphisms (SNPs). Variations greater than 1kilobase of DNA is called CNV. Other variations include microsatellite instability (MSH), variable tandem repeats, transposable elements, deletions, inversions, and duplications. Furthermore, transcripts of gene fusions provide a wealth of information on the functional input of fused genes(64, 93-97)

4). Methylation status: Gene promoter methylation at the cytosine-guanine sites (CpG islands) leads to gene transcription suppression. Demethylation leads to transcript activation. Methylation status of cancer genes adds complexity to interpretation of gene profiling. Finding methylation status involves use of(i) methylation-sensitive or methylation-specific endonucleases, (ii) Sodium bisulphate treatment and DNA sequencing (iii) target amplification by capture and ligation (23, 98, 99).

5). miRNA Profiling: microRNAs (18-24 nucleotides) are present in plants and animals. miRNAs play significant roles in normal development, cellular responses and in human cancer. miRNAs represses mRNA trascription via partial complimentary alignment with targer mRNAs. miRNA profiling adds to complexity of cancer cell and tissue profiling. Alignment with their targets tells us why certain transcripts may be down-regulated (22, 100-105).

6). Gene Sequencing: Human genome sequencing ushered in the grand promise of genomics medicine in 2000. Now individual genes and chromosomes, and gene fusions are sequenced; these efforts will have impact on the information sets necessary for

diagnosis, treatment and predicting outcome in cancer (38). This enables direct analysis of mutations within the gene components of interest. Gene annotation enables linking of sequences to active transcripts (96). Next generation sequencing can overcome some drawbacks of qPCR and cDNA microarray and can enable assessment of cancer gene mutations, copy number variations (CNV), SNPs, miRNA and transcription profiling (106). Emerging sequencing methods, labeled third generation sequencing, include single-molecule-real-time sequencing (SMRT), direct sequencing using direct imaging and sequencing using nanopores (107).

7). Cancer Grading: Cancer grading to reflect the extent of differentiation is done for many cancers. Grading systems vary and depend on cancer type. Morphologic attributes are used to grade (score) prostate cancer (108, 109). Cancer grades will form an integral part of the model. Cancer stem cell compartment within the tumor can be estimated by proteomics- immunohistochemical methods using antibodies to CD24, CD44, CD133 and CD26.

The integrated model can be generated using comprehensive bioinformatics and data mining tools. OmicsAnalyzer is one such tool, which is a plug-in for Cytoscape (cytoscape.org), a web based platform for protein/gene network modeling, functional annotation and analysis. (110). Bioinformatics tools for classification, data mining and modeling are available (R-project.org). The integrated model of cancer classification can help stratify individuals for target treatment and disease outcome based on the treatment (Table 2).

Table 2. Anticipated data sets from a biopsy for decision making in targeted cancer treatment. Gene profiling signatures (growth, oncogene and stemness), and proteomics can be correlated with CNVs, mutations and gene fusions. Protein interaction maps can suggest treatment targets, possible pathways of resistance.

Cancer morphology	Adenocarcinoma, Grade 2 10% Mucinous component 10% Cancer stem cells
Proteomics/Transcriptome/Methylome	Up-regulated cell death pathway Up-regulated growth factor signaling Hyper-methylation of kinases Decreased expression of autophagy pathway miRNA clusters in some autophagy genes De-regulated metabolic genes
Mutations (Sequencing/CNV)	Mutations in 24 genes Gene fusions Copy number changes in 10 genes Clustered mutations in 8 genes BRAF, KRAS, EGFR, and p53 mutations

Conclusion

Cancer cells are endowed with capacities for uninhibited proliferation, invasion and metastasis. Cancer initiation, driver mutations, progression and metastases are common attributes for both stochastic and cancer stem cell models; cancer stem cells possess attributes that enhance survival in all environments hence more suitable as model of cancer. Integrated models that capture every essence of a cancer could enhance the ability to target different components of cancer for maximum therapeutic effect.

Conflict of Interest

The author has declared that no conflict of interest exists.

References

- Ferlay J, Shin H-R, Bray F, Forman D, Mathers C, Parkin D. Estimates of worldwide burden of cancer in 2008: GLOBOCAN2008. *Int J Cancer* 2010;127:2893-2917.
- Van Raamsdonk C, Bezrookove V, Green G, et al. Frequent somatic mutations of GNAQ in uveal melanoma and blue nevi. *Nature* 2009;457:599-602.
- Cerroni L, Barnhill R, Elder D, et al. Melanocytic Tumors of Uncertain Malignant Potential. *American J Surgical Pathology* 2008;34(3):314-326.
- Stelow E, Shaco-Levy R, Bao F, Garcia J, Klimstra D. Pancreatic Acinar Cell Carcinomas With Prominent Ductal Differentiation: Mixed Acinar Ductal Carcinoma and Mixed Acinar Endocrine Carcinoma. *American J Surgical Pathology* 2010;34(4):510-518.
- Golub T, Slonim D, Tamayo P, et al. Molecular Classification of Cancer: Class Discovery and Class Prediction by Gene Expression Monitoring. *Science* 1999;286(5439):531-537.
- Segal Ep, Friedman N, Kaminski N, Regev A, Koller D. From signature to models: understanding cancer using microarrays. *Nature Genetics* 2005;37:538-45.
- Bair E, Tibshirani R. Semi-Supervised Methods to Predict Patient Survival from Gene Expression Data. *PLoS Biology* 2004;2:0511.
- van de Vijver M, He Y, van't Veer LJ, et al. A Gene Expression Signature As A Predictor Of Survival In Breast Cancer. *NEJM* 2002;347(25):1999-2009.
- Crowson A. Basal cell carcinoma: biology, morphology and clinical implications. *Modern Pathology* 2006;19:S127-S147.
- Berman J. Modern classification of neoplasms: reconciling differences between morphologic and molecular approaches. *BMC Cancer* 2005;5:100.
- Loscalzo J, Kohane I, Barabasi A-L. Human disease classification in the postgenomic era: A complex systems approach to human pathobiology. *Molecular Systems Biology* 2007;3:124.
- Webb C, Pass H. Translation research: from accurate diagnosis to appropriate treatment. *Journal of Translational Medicine* 2004;2:35.
- Barabasi A-L, Oltvali Z. Network Biology: Understanding The Cells Functional Organization. *Nature Reviews Genetics* 2004;5:101-113.
- Hanahan D, Weinberg R. The Hallmarks of Cancer. *Cell* 2000;100:57-70.
- Hahn W, Weinberg R. Modeling the Molecular Circuitry of Cancer. *Nature Reviews Cancer* 2002;2:331-341.
- Chen Z, Sun L. Nonproteolytic Functions of Ubiquitin in Cell Signaling. *Molecular Cell* 2009;33:275-286.
- Goodarzi H, Elemento O, Tavazoie S. Revealing Global Regulatory Perturbations across Human Cancers. *Molecular Cell* 2009;36:900-911.
- Lemmon M, Schlessinger J. Cell Signaling by Receptor Tyrosine Kinases. *Cell* 2010;141:1117-1134.
- Amaravadi R, Thompson C. The Roles of Therapy-Induced Autophagy and Necrosis in Cancer Treatment. *Clin Cancer Res* 2007;13(24):7271-7279.
- Baehrecke E. Autophagy: dual roles in life and death? *Nature Reviews Molecular Cell Biology* 2005;6:505-510.
- Carthew R, Sontheimer E. Origins and Mechanisms of miRNAs and siRNAs. *Cell* 2009;136:642-655.
- Esquela-Kerscher A, Slack F. Oncomirs- microRNAs with a role in cancer. *Nature Reviews Cancer* 2006;6:259-269.
- Muntean A, Hess J. Epigenetic Dysregulation in Cancer. *American Journal of Pathology* 2009;175(4):1353-1361.
- Imam J, Buddavarapu K, Lee-Chang J, et al. MicroRNA-185 suppresses tumor growth and progression by targeting the Six1 oncogene in human cancers. *Oncogene* 2010; 29: 4971-4979.
- Copeland N, Jenkins N. Harnessing the transposons for cancer gene discovery. *Nature Reviews Cancer* 2010;10:696-706.
- O'Donnell K, Burns K. Mobilizing diversity: transposable element insertions in genetic variation and disease. *Mobile DNA* 2010;1:21.
- Gatenby R, Gillies R. Why do cancers have high aerobic glycolysis. *Nature Reviews Cancer* 2004;4(11):891-899.
- Kroemer G, Pouyssegur J. Tumor Cell Metabolism: Cancer's Achilles' Heel. *Cancer Cell* 2008;13:472-482.
- Levine A, Puzio-Kuter A. The Control of the Metabolic Switch in Cancers by Oncogenes and Tumor Suppressor Genes. *Science* 2010;330:1340-1344.
- Bild A, Potti A, Nevins J. Linking oncogenic pathways with therapeutic opportunities. *Nature Reviews Cancer* 2006;6:735-740.
- Collado M, Blasco M, Serrano M. Cellular Senescence in Cancer and Aging. *Cell* 2007;130:223-233.
- Levine B, Kroemer G. Autophagy in the Pathogenesis of Disease. *Cell* 2008;132:27-42.
- Sharpless N, DePinho R. Telomeres, stem cells, senescence, and cancer. *J Clinical Investigation* 2004;113(2):160-168.
- Feldser D, Greider C. Short Telomeres Limit Tumor Progression in Vivo by Inducing Senescence. *Cancer Cell* 2007;11(5):461-470.
- Di Micco R, Fumagalli M, Cicalese A, et al. Oncogene-induced senescence is a DNA damage response triggered by DNA hyper-replication. *Nature* 2006;444:638-642.
- Kassenbrock K, Plaks V, Werb Z. Matrix Metalloproteinases: Regulators of the Tumor Microenvironment. *Cell* 2010;141(1):52-57.
- Qian B-Z, Pollard J. Macrophage Diversity Enhances Tumor Progression and Metastases. *Cell* 2010;141(1):39-51.
- Bozic I, Antal T, Ohtsuki H, et al. Accumulation of driver and passenger mutations during tumor progression. *PNAS* 2010;107(43):18545-18550.
- Halazonetis T, Gorgoulis V, Bartek J. An Oncogene-Induced DNA Damage Model for Cancer Development. *Science* 2008;319:1352-1355.
- Stephens P, Greenman C, Fu B, et al. Massive Genomic Rearrangement Acquired in a Single Catastrophic Event during Cancer Development. *Cell* 2011;144(1):27-40.
- Goldstein A, Huang J, Guo C, Garraway I, Witte O. Identification of a Cell of Origin for Human Prostate Cancer. *Science* 2010;329:568-571.
- Clarke M, Fuller M. Stem Cells and Cancer: Two Faces of Eve. *Cell* 2006;124:1111-1115.
- Jordan C, Guzman M, Noble M. Cancer Stem Cells. *New Engl J Med* 2006;355:1253-1256.
- Fodde R. The Stem of Cancer. *Cancer Cell* 2009;15(2):87-90.

45. Shackelton M, Quintana E, Fearon E, Morrison S. Heterogeneity in Cancer: Cancer Stem Cells versus Clonal Evolution. *Cell* 2009;138:822-829.
46. Adams J, Strasser A. Is Tumor Growth Sustained by Rare Cancer Stem Cells or Dominant Clones? *Cancer Research* 2008;68(11):4018-4021.
47. Kim J, Woo A, Chu J, et al. A Myc Network Accounts for Similarities between Embryonic Stem and Cancer Cell Transcription Programs. *Cell* 2010;143(2):313-324.
48. Al-Hajji M, Wicha M, Benito-Hernandez A, Morrison S, Clarke M. Prospective identification of breast cancer stem cells. *PNAS* 2003;100:3983-3988.
49. Vescovi A, Galli R, Reynolds B. Brain tumor stem cells. *Nature Reviews Cancer* 2006;6:425-436.
50. Quintana E, Shackelton M, Sabel M, Fullen D, Johnson T, Morrison S. Efficient tumor formation by single human melanoma cells. *Nature* 2008;456:593-598.
51. Hu L, McArthur C, Jaffe R. Ovarian cancer stem-like side-population cells are tumorigenic and chemoresistant. *British J Cancer* 2010;102:1276-1283.
52. Lawson D, Witte O. Stem cells in prostate cancer initiation and progression. *The Journal of Clinical Investigation* 2007;117(8):2044-2050.
53. Sell S, Leffert H. Liver Cancer Stem Cells. *J Clin Oncol* 2008;26(17):2800-2805.
54. Ponnusamy M, Batra S. Ovarian cancer: emerging concept on cancer stem cells. *Journal of Ovarian Research* 2008;1:4.
55. Kern S, Shibata D. The Fuzzy Math of Solid Tumor Stem Cells: A Perspective. *Cancer Research* 2007;67:89850-8988.
56. Rich J. Cancer Stem Cells in Radiation Resistance. *Cancer Research* 2007;67:8980-8984.
57. Eyler C, Rich J. Survival of the Fittest: Cancer Stem Cells in Therapeutic Resistance and Angiogenesis. *J Clin Oncol* 2008;26(17):2839-2845.
58. Keysar S, Jimeno A. More than Markers: Biological Significance of Cancer Stem Cell-Defining Molecules. *Mol Cancer Ther* 2010;9(9):2450-7.
59. Talmadge J. Clonal Selection of Metastasis within the Life History of a tumor. *Cancer Research* 2007;67(24):11471-11475.
60. Klein C. The Metastasis Cascade. *Science* 2008;321:1785-1787.
61. Weinberg R. Leaving Home Early: Reexamination of the Canonical Models of Tumor Progression. *Cancer Cell* 2008;14:283-284.
62. Bogenrieder T, Herlyn M. Axis of evil: molecular mechanisms of cancer metastasis. *Oncogene* 2003;22:6524-6536.
63. Yachida S, Jones S, Bozic I, et al. Distant metastasis occurs late during genetic evolution of pancreatic cancer. *Nature* 2010;467:1114-1117.
64. Campbell P, Yachida S, Mudie L, et al. The patterns and dynamics of genomic instability in metastatic pancreatic cancer. *Nature* 2010;467:1109-1113.
65. Balic M, Lin H, Young L, et al. Most Early Disseminated Cancer Cells Detected in Bone Marrow of Breast Cancer Patients Have a Putative Breast Cancer Stem Cell Phenotype. *Clinical Cancer Research* 2006;12(19):5615-5621.
66. Pang R, Law W, Chu A, et al. A Subpopulation of CD26+ Cancer Stem Cells with Metastatic Capacity in Human Colorectal Cancer. *Cell Stem Cell* 2010;6:603-615.
67. Herman P, Huber S, Herrier T, et al. Distinct Populations of Cancer Stem Cells Determine Tumor Growth and Metastatic Activity in Human Pancreatic Cancer. *Cell Stem Cell* 2007;1:313-323.
68. de Bono J, Ashworth A. Translating cancer research into targeted therapeutics. *Nature* 2010;467:543-549.
69. Varmus H. Ten Years On- The Human Genome and Medicine. *New England Journal of Medicine* 2010;362:2028-2029.
70. Bentley D. Genomes for medicine. *Nature* 2004;429:440-445.
71. Evans W, Relling M. Moving towards individualized medicine with pharmacogenomics. *Nature* 2004;429:464-468.
72. Strausberg R, Simpson A, Old L, Riggins G. Oncogenomics and the development of new cancer therapies. *Nature* 2004;429:469-474.
73. Feero W, Guttmacher A, Collins F. Genomic Medicine- An Updated Primer. *N Engl J Med* 2010;362:2001-2011.
74. Kim S, Hahn W. Cancer genomics: integrating form and function. *Carcinogenesis* 2007;28(7):1387-1392.
75. Nevins J, Potti A. Mining gene expression profiles: expression signatures as cancer phenotypes. *Nature Reviews Genetics* 2007;8:601-609.
76. Rhodes D, Chinnaiyan A. Integrative analysis of cancer transcriptome. *Nature Genetics* 2005;17:S31-S37.
77. Taylor B, Schultz N, Hieronymus H, et al. Integrative Genomic Profiling of Human Prostate Cancer. *Cancer Cell* 2010;18:11-22.
78. Nomura D, Dix M, Cravatt B. Activity-based protein profiling for biochemical pathway discovery in cancer. *Nature Reviews Cancer* 2010;10:630-638.
79. Colinge J, Bennett K. Introduction to Computational Proteomics. *PLoS Computational Biology* 2007;3(7):e114.
80. Domon B, Aebersold R. Options and considerations when selecting a quantitative proteomics strategy. *Nature Biotechnology* 2010;28(7):710-721.
81. Kononen J, Bubendorf L, Kallioniemi A, et al. Tissue microarrays for high-throughput molecular profiling of tumor specimens. *Nature Medicine* 1998;4(7):844-847.
82. Celis J, Gromov P. Proteomics in translational cancer research: Toward an integrated approach. *Cancer Cell* 2003;3(1):9-16.
83. Celis J, Gromov P, Gromova I, Moreira J, Cabezon T, Ambartsumian Nea. Integrating Proteomic and Functional Genomic Technologies in Discovery-driven Translational Breast Cancer Research. *Mol and Cellular Proteomics* 2003;2:369-377.
84. Brennan D, O'Connor D, Rexhepaj E, Ponten F, Gallagher W. Antibody-based proteomics: fast-tracking molecular diagnostics in oncology. *Nature Reviews Cancer* 2010;10:605-617.
85. Iadevaia S, Lu Y, Morales F, Mills G, Ram P. Identification of Optimal Drug Combinations Targeting Cellular Networks: Integrating Phospho-Proteomics and Computational Network Analysis. *Cancer Research* 2010;70(17):6704-6714.
86. Xu Q, Lam K. Protein and Chemical Microarrays- Powerful Tools for Proteomics. *J Biomed and Biotechnology* 2003; 5:257-266.
87. MacBeath G. Protein microarrays and proteomics. *Nature genetics Supplement* 2002;32:526-532.
88. Domon B, Aebersold R. Mass Spectrometry and Protein Analysis. *Science* 2006;312:212-217.
89. Nesvizhskii A, Vitek O, Aebersold R. Analysis and validation of proteomic data generated by tandem mass spectrometry. *Nature Methods* 2007;4(10):787-797.
90. Hu S, Wong D. Lectin microarray. *Proteomics Clin Appl* 2009;3(2):148-154.
91. North S, Hitchen P, Haslam S, Dell A. Mass spectrometry in the analysis of N-and-O -linked glycans. *Curr Opin Struct Biol* 2009;19(5):498-506.
92. Ozsolak F, Platt A, Jones D, et al. Direct RNA sequencing. *Nature* 2009; 461: 814-818.
93. Yeang C-H, McCormick F, Levine A. Combinatorial patterns of somatic gene mutations in cancer. *The FASEB J* 2008;22:2605-2622.
94. Beroukhim R, CH M, Porter D, Wei G, Raychaudhuri S. The landscape of somatic copy-number alteration across human cancers. *Nature* 2010;463:899-905.
95. Freeman J, Perry G, Feuk L, et al. Copy number variation: New insights in genome diversity. *Genome Research* 2006;16:949-961.

96. Maher C, Kumar-Sinha C, Cao X, et al. Transcriptome sequencing to detect gene fusions in cancer. *Nature* 2009; 458: 97-101.
97. Network TCGAR. Comprehensive genomic characterization defines human glioblastoma genes and core pathways. *Nature* 2008;455(23 Oct):1061-1068.
98. Nautiyal S, Carlton V, Lu Y, et al. High-throughput method for analyzing methylation of CpGs in targeted genomic regions. *PNAS* 2010; 107(28): 12587-12592.
99. Lister R, Pelizzola M, Dowen R, et al. Human DNA methylomes at base resolution show widespread epigenomic differences. *Nature* 2009; 462: 315-322.
100. Zhang H, Li Y, Lai M. The microRNA network and tumor metastasis. *Oncogene* 2010;29:937-948.
101. Moazed D. Small RNAs in transcriptional gene silencing and genome defence. *Nature* 2009;457:413-420.
102. Lee Y, Dutta A. MicroRNAs in cancer. *Annu Rev Pathol* 2009;4:199-227.
103. Garofalo M, Condorelli G, Croce C, Condorelli G. MicroRNAs as regulators of death receptors signaling. *Cell Death and Differentiation* 2009; 17: 200-208.
104. Bandyopadhyay S, Mitra R, Maulik U, Zhang M. Development of the human cancer microRNA network. *Silence* 2010;1:6.
105. Sotiropoulou G, Pampalakis G, Lianidou E, Mourelatos Z. Emerging roles of microRNAs as molecular switches in the integrated circuit of the cancer cell. *RNA* 2009;15:1443-1461.
106. Dahl F, Stenberg J, Fredrikson S, et al. Multigene amplification and massively parallel sequencing for cancer mutation discovery. *PNAS* 2007;104(22):9387-9392.
107. Schadt E, Turner S, Kasarskis A. A window into third-generation sequencing. *Human Molecular Genetics* 2010;19(2):R227-R240.
108. Gleason D, Mellinger G. Prediction of prognosis for prostatic adenocarcinoma by combined histologic grading and clinical staging. *J Urology* 1974;111:58-64.
109. Gleason D. Histologic grading of prostate cancer: a perspective. *Human Pathology* 1992;23(3):273-279.
110. Xia T, Hemert J, Dickerson J. OmicsAnalyzer: a Cytoscape plug-in suite for modeling omics data. *Bioinformatics* 2010;26(23):2995-2996.