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#### **Discovery of Novel** *N***-Acetylpyrazolines as Microtubule Inhibitors:**

#### Design, Synthesis, Anticancer Evaluation, and Molecular Docking Study

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#### Abstract

In the current study, a new series of *N*-acetylpyrazolines (**6a-d**) were designed and synthesized from their corresponding chalcones and hydrazine hydrate in acidic medium. The *N*-acetylpyrazolines (**6a-d**) were tested for their antihepatocellular activity against liver cancer (HuH-7, and HepG-2), and normal BNL cell lines and compared with paclitaxel, colchicine, and combrestatin A-4 (CA-4), as standards, and their IC<sub>50</sub> values were determined. The 3',4',5'-trimethoxyphenyl *N*-acetylpyrazoline derivative **6d** was the found the most potent *N*-acetylpyrazoline derivative IC<sub>50</sub> = 0.30, and 77.30  $\mu$ M, respectively, and found non-cytotoxic to the normal BNL cell line. While compounds **6b**, and **6c** revealed lower anticancer activity against HuH-7 cell line IC<sub>50</sub> = 14.50, and 11.00  $\mu$ M, respectively. Moreover, the *N*-acetylpyrazolines **6a-d** were evaluated for their anticancer screening against different cancer cell lines at 10  $\mu$ M by the Developmental Therapeutic Program (DTP) - NCI - USA, and they showed mean GI% ranges 8.87-64.54%. The 3',4',5'-trimethoxyphenyl *N*-acetylpyrazolines **6c**, and **6d** revealed potent anticancer activities and lethal effects against lung cancer cell line (HOP-92) for **6c** and melanoma cell line (SK-MEL-5) for **6d** with GI% values of 104.80, and 109.52%, respectively. Furthermore, the *N*-acetylpyrazolines **6a-d** enhanced tublin polymerization, and showed tubulin-stabilizing effects as paclitaxel at 50  $\mu$ M. A molecular docking study was performed for the *N*-acetylpyrazolines **6a-d** to investigate the binding pattern at the Taxol-binding site of microtubules. A physicochemical prediction and ADME properties as well as drug-likeness and medicinal chemistry friendliness of compounds **6a-d** were also performed.

Keywords: N-acetylpyrazolines; Tubulin polymerization inhibitors; Anticancer activity; Molecular docking study

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#### 1. Introduction

Cancer is the most serious life-threatening disease, and one of the leading causes of global mortality [1, 2]. Liver cancer is considered as one of the global health challenges and its incidence is increasing worldwide [3], and it is estimated that more than one million individuals will be affected annually by liver cancer by 2025 [4]. Eighty percent of instances of liver cancer are attributed to hepatocellular carcinoma (HCC), which is the most common type of liver cancer [5]. Tubulin is one of the molecular targets for antitumor agents via targeting the dynamic process of microtubules assembly, and disassembly which can be hindered through various targeting agents that can bind to different sites in the  $\beta$ -tubulin subunit [6]. The microtubule targeting agents, Interference with the microtubule function, can do cellular arrest at G2/M phase, hence, leading to cell death through both induction of apoptosis and necrosis [7, 8]. The structural elements of the mitotic spindle are microtubules, which are hollow cylindrical filaments that are assembled by the aggregation of  $\alpha$ - and  $\beta$ tubulin heterodimers [9]. Numerous biological activities, such as intracellular trafficking, the formation of the cell skeleton, cell migration, and cell division, depend critically on microtubules [10, 11]. Paclitaxel (I), and colchicine (II) are naturally occurring microtubules targeting agents at distinctive binding sites (Figure 1), and act as a microtubule stabilizing and destabilizing agent, respectively [12]. The microtubule stabilizing agents bind to taxane site that can accelerate microtubule assembly; paclitaxel (I), have been successfully applied in tumor therapy, and some microtubule stabilizing agents have entered also in clinical studies [13, 14]. However, these tubulin targeting agents have various drawbacks; poor pharmacokinetic properties, high lipophilicity, low water solubility, and multi-drug resistance [15, 16]. Pyrazoline containing compounds, privileged scaffold found in many biologically active agents, that are characterized with diversity in their biological activities including anticancer activity [17-23]. Pyrazolines act as anticancer agents via different mechanisms including; tyrosine kinase inhibitors [24-28], and tubulin suppressors [29-32]. The Nmethylindole-pyrazoline hybrids (IIIa-c) (Figure 2) showed tubulin polymerization inhibition and anticancer activities [33], whereas, the

3',4',5'trimethoxyphenyl N-methylindole-pyrazoline hybrid (**IIIa**) revealed tubulin inhibition  $IC_{50} = 2.12$ µM, and potent anticancer activity against diverse cancer cell lines; HepG2, HeLa, MCF-7, and A549 with IC<sub>50</sub> values 0.31, 0.21, 0.29, and 0.26 µM, respectively [34]. The 3-methoxyphenyl Nmethylindole-pyrazoline hybrid (IIIb) (YMR-65) showed relatively similar tubulin polymerization inhibition as compound (IIIa),  $IC_{50} = 2.44 \mu M$ , and high anticancer activity  $IC_{50} = 0.31, 0.25, 0.32$ , and 0.28 against HepG2, HeLa, MCF-7, A549 cancer cell lines, respectively [33, 35]. Furthermore, it demonstrated potent in vivo anticancer efficacy, and toxicity low in pharmacokinetic and pharmacodynamics studies [36]. Later, the Nnicotinoylpyrazoline derivative (IIIc) was developed for better stability, and less susceptibility to metabolism [37]. Moreover, it displayed more potent tubulin inhibition  $IC_{50} = 1.60 \ \mu M$ , and potent anticancer activity GI<sub>50</sub> ranges 0.09-0.59 µM against HepG2, HeLa, A-549, and MCF-7 cancer cell lines. An in vivo study was also performed and demonstrated a comparable anticancer activity to combrestatin A-4 (CA-4) in HeLa-xenograft mice model without tissue damage, and weight loss [37]. Besides, the *N*-phenylpyrazole derivative (IVa) showed anticancer activity against the colon cancer cell line HCT-116 (IC<sub>50</sub> =  $2.65 \mu$ M), and tubulin polymerization inhibition  $IC_{50} = 10.9 \mu M$ , as well as induction of cell cycle arrest in HCT-116 cancer cell line at G2/M phase. in vivo study for this pyrazole derivative (IVa) showed effect in decreasing tumor growth in a HCT-116 mouse model and showed no toxicity [38]. In contrast, the N-phenylpyrazole derivative (IVb) caused cell cycle arrest in G2/M phase and interfered with the production of mitotic spindles in MCF-7 cells, demonstrating strong anticancer activity against the cancer cell lines B16F10, Hela, and MCF-7 (IC<sub>50</sub> = 1.3, 6.0, and 5.5µM, respectively) (Figure 2) [30]. The indole-pyrazole hybrid (V) showed potent anticancer activity against the hepatocellular cancer cell (HCC) lines; HuH-7, HepG-2, Mahlavu, and SNU475 with IC<sub>50</sub> range 0.6-2.9 µM, and showed moderate inhibitory activity against tubulin polymerization (IC<sub>50</sub> = 19  $\mu$ M). It also induced cell cycle arrest at the G2/M phase in HuH-7 and Mahlavu cell lines, as well as induction of apoptosis in HCC cells (Figure 2) [32].

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Figure 1: Chemical structures of some naturally occurring tubulin polymerization inhibitors: paclitaxel (I), and colchicine (II).

Based on the above mentioned findings, a series of novel *N*-acetylpyrazolines **6a-d** were designed, synthesized from the corresponding chalcones, and evaluated *in vitro* for their anticancer activity against liver cancer and normal (BNL) cell lines, as well as anticancer screening against a panel of cancer cell lines at 10  $\mu$ M. This was coupled with tubulin polymerization assay for the synthesized *N*acetylpyrazolines. To investigate the binding pattern of the synthesized compounds, a molecular docking study was also performed for the *N*-acetylpyrazolines to at the paclitaxel-binding site of microtubules *via* key amino acids.

### 2. Experimental 2.1. Chemistry:

The measured melting points were determined using a Daihan Melting Point Analyzer, and were uncorrected. A 400 (100) MHz Bruker Spectrometer at the Microanalytical Unit, Faculty of Pharmacy, Cairo University, Egypt, was used to report the NMR spectra. An Advion compact mass spectrometer (CMS) at Nawah Scientific in Cairo, Egypt, was used to report mass spectra (MS-ESI).



Figure 2: Chemical structures of some reported pyrazolines (IIIa-c), and pyrazoles (IV, V) as tubulin polymerization inhibitors

(V)

### **2.1.1.** General procedure for the synthesis of chalcones 5a-d

To a stirred solution of the appropriate 4-(benzyloxy)-3-methoxybenzaldehyde **3a-c** (5 mmol) and 3',4'dimethoxyacetophenone **4a** or 3',4',5'trimethoxyacetophenone **4b** (5 mmol) in MeOH (40 ml) was added 50% aqueous KOH (15 ml) and heated under reflux with stirring for 1-4 h. The solvent was evaporated and the precipitate was formed, filtered, evaporated and purified either by recrystallization from absolute ethanol or by column chromatography ethyl acetate: petroleum ether (60-80 °C) (1:1).

## 2.1.1.1. (*E*)-3-(4-(Benzyloxy)-3-methoxyphenyl)-1-(3,4-dimethoxyphenyl)prop-2-en-1-one (5a)

Yellow solid; Yield (80%); mp 173-174 °C; IR (KBr) vmax: 3062, 1661 cm-1; 1H NMR (400 MHz, DMSOd<sub>6</sub>) δ: 3.85 (s, 3H, OCH<sub>3</sub>), 3.87 (s, 6H, 2 OCH<sub>3</sub>), 5.16 (s, 2H, OCH<sub>2</sub>), 7.10 (dd, *J* = 2.4 Hz, *J* = 8.4 Hz, 2H, H<sub>ar</sub>), 7.30-7.42 (m, 4H, H<sub>ar</sub>), 7.46 (d, J = 8.0 Hz, 2H,  $H_{ar}$ ), 7.54 (s, 1H,  $H_{ar}$ ), 7.60 (s, 1H,  $H_{ar}$ ), 7.67 (d, J =12.0 Hz, 1H, CH=CH-CO), 7.83 (d, J = 16.0 Hz, 1H, CH=CH-CO), 7.92 (d, J = 8.0 Hz, 1H, H<sub>ar</sub>) ppm; <sup>13</sup>C NMR (100 MHz, DMSO-d<sub>6</sub>) δ: 55.63 (OCH<sub>3</sub>), 55.82 (OCH<sub>3</sub>), 55.97 (OCH<sub>3</sub>), 69.89 (OCH<sub>2</sub>), 110.73 (CH<sub>ar</sub>), 110.83 (CHar), 111.39 (CHar), 113.18 (CHar), 119.77 (CHar), 123.41 (CH=CH-CO), 127.96, (CHar), 128.03 (Car), 128.05 (CHar), 128.53 (CHar), 130.82 (CHar), 136.81 (Car), 143.59 (CH=CH-CO), 148.87 (Car), 149.34 (Car), 150.11 (Car), 154.16 (Car), 187.40 (C=O) ppm; MS (ESI<sup>+</sup>, m/z): 427.1 (89.6%) [M + Na]<sup>+</sup>; Anal. Calcd for C<sub>25</sub>H<sub>24</sub>O<sub>5</sub>: C, 74.24; H, 5.98; Found: C, 74.33; H, 6.11.

#### 2.1.1.2. (*E*)-3-(4-(4-Fluorobenzyloxy)-3-methoxy phenyl)-1-(3,4-dimethoxyphenyl)prop-2-en-1-one (5b)

Yellow solid; Yield (86%); mp 135-136 °C; IR (KBr) umax: 3068, 1665 cm<sup>-1</sup>; <sup>1</sup>H NMR (400 MHz, DMSOd<sub>6</sub>) δ: 3.86 (s, 3H, OCH<sub>3</sub>), 3.87 (s, 6H, 2 OCH<sub>3</sub>), 5.14 (s, 2H, OCH<sub>2</sub>), 7.10 (dd, J = 3.2 Hz, J = 8.4 Hz, 2H,  $H_{ar}$ ), 7.23 (t, J = 8.4 Hz, 2H,  $H_{ar}$ ), 7.40 (d, J = 8.0 Hz, 1H, Har), 7.49-7.55 (m, 3H, Har), 7.60 (s, 1H, Har), 7.68 (d, J = 15.6 Hz, 1H, CH=C<u>H</u>-CO), 7.84 (d, J = 15.6Hz, 1H, CH=CH-CO), 7.93 (d, J = 8.4 Hz, 1H, H<sub>ar</sub>) ppm; <sup>13</sup>C NMR (100 MHz, DMSO-*d*<sub>6</sub>) δ: 55.58 (OCH<sub>3</sub>), 55.77 (OCH<sub>3</sub>), 55.82 (OCH<sub>3</sub>), 69.12 (OCH<sub>2</sub>), 110.70 (CHar), 110.78 (CHar), 111.40 (CHar), 113.21 (CH<sub>ar</sub>), 115.18 (d,  ${}^{2}J_{CF} = 22.0$  Hz, <u>C</u>H<sub>ar</sub>-C<sub>ar</sub>F), 119.78 (CH=CH-CO), 123.32 (CH<sub>ar</sub>), 128.07 (C<sub>ar</sub>), 130.11 (d,  ${}^{3}J_{CF} = 8.0 \text{ Hz}, \underline{C}H_{ar}-CH_{ar}-C_{ar}F$ ), 130.77 (CH<sub>ar</sub>), 133.00 (CHar), 133.03 (Car), 143.47 (CH=CH-CO), 148.81  $(C_{ar}), 149.29 (C_{ar}), 149.92 (C_{ar}), 153.10 (C_{ar}), 160.65$ (d,  ${}^{1}J_{CF} = 242.0$  Hz, C<sub>ar</sub>-F), 187.30 (C=O) ppm; MS (ESI+, m/z): 445.1 (79.8%) [M + Na]+; Anal. Calcd for C<sub>25</sub>H<sub>23</sub>FO<sub>5</sub>: C, 71.08; H, 5.49; Found: C, 71.23; H, 5.56.

### **2.1.1.3.** (*E*)-**3**-(**4**-(**Benzyloxy**)-**3**-methoxyphenyl)-**1**-(**3**,**4**,**5**-trimethoxyphenyl)prop-**2**-en-**1**-one (**5**c)

Yellow solid; Yield (88%); mp 128 °C; IR (KBr)  $\upsilon_{max}$ : 3063, 1651 cm<sup>-1</sup>; <sup>1</sup>H NMR (400 MHz, DMSO- $d_0$ )  $\delta$ : 3.77 (s, 3H, OCH<sub>3</sub>), 3.87 (s, 3H, OCH<sub>3</sub>), 3.90 (s, 6H,

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2 OCH<sub>3</sub>), 5.17 (s, 2H, OCH<sub>2</sub>), 7.11 (d, J = 8.4 Hz, 1H, H<sub>ar</sub>), 7.32-7.36 (m, 1H, H<sub>ar</sub>), 7.38-7.42 (m, 4H, H<sub>ar</sub>), 7.43-7.47 (m, 3H, H<sub>ar</sub>), 7.53 (s, 1H, H<sub>ar</sub>), 7.71 (d, J =15.2 Hz, 1H, CH=C<u>H</u>-CO), 7.80 (d, J = 15.6 Hz, 1H, C<u>H</u>=CH-CO) ppm; <sup>13</sup>C NMR (100 MHz, DMSO- $d_6$ )  $\delta$ : 55.88 (OCH<sub>3</sub>), 56.25 (OCH<sub>3</sub>), 60.20 (OCH<sub>3</sub>), 69.85 (OCH<sub>2</sub>), 106.21 (CH<sub>ar</sub>), 112.07 (CH<sub>ar</sub>), 113.25 (CH<sub>ar</sub>), 119.85 (CH<sub>ar</sub>), 123.20 (CH=<u>C</u>H-CO), 127.86 (CH<sub>ar</sub>), 127.96 (C<sub>ar</sub>), 128.46 (CH<sub>ar</sub>), 133.31 (C<sub>ar</sub>), 136.76 (C<sub>ar</sub>), 141.87 (C<sub>ar</sub>), 144.27 (<u>C</u>H=CH-CO), 149.26 (C<sub>ar</sub>), 150.19 (C<sub>ar</sub>), 152.90 (C<sub>ar</sub>), 187.93 (C=O) ppm; Anal. Calcd for C<sub>26</sub>H<sub>26</sub>O<sub>6</sub>: C, 71.87; H, 6.03; Found: C, 71.93; H, 6.14.

#### 2.1.1.4. (*E*)-3-(4-(4-Methylbenzyloxy)-3-methoxy phenyl)-1-(3,4,5-trimethoxyphenyl)prop-2-en-1one (5d)

Yellow solid; Yield (84%); mp 103 °C; IR (KBr) v<sub>max</sub>: 3078, 1651 cm<sup>-1</sup>; <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>) δ: 2.30 (s, 3H, CH<sub>3</sub>), 3.76 (s, 3H, OCH<sub>3</sub>), 3.85 (s, 3H, OCH<sub>3</sub>), 3.89 (s, 6H, 2 OCH<sub>3</sub>), 5.10 (s, 2H, OCH<sub>2</sub>), 7.09 (d, J = 8.4 Hz, 1H, H<sub>ar</sub>), 7.19 (d, J = 8.0 Hz, 2H, H<sub>ar</sub>), 7.33 (d, J = 7.6 Hz, 2H, H<sub>ar</sub>), 7.40-7.43 (m, 3H, H<sub>ar</sub>), 7.51 (s, 1H, H<sub>ar</sub>), 7.69 (d, J = 15.6 Hz, 1H, CH=CH-CO), 7.79 (d, J = 15.6 Hz, 1H, C<u>H</u>=CH-CO) ppm; <sup>13</sup>C NMR (100 MHz, DMSO-d<sub>6</sub>) δ: 20.90 (CH<sub>3</sub>), 55.99 (OCH<sub>3</sub>), 56.37 (OCH<sub>3</sub>), 60.36 (OCH<sub>3</sub>), 69.89 (OCH<sub>2</sub>), 106.30 (CHar), 112.03 (CHar), 113.37 (CHar), 119.88 (CH<sub>ar</sub>), 123.45 (CH=<u>C</u>H-CO), 127.89 (C<sub>ar</sub>), 128.12 (CHar), 129.16 (CHar), 133.45 (Car), 133.80 (Car), 137.43 (Car), 141.99 (Car), 144.51 (CH=CH-CO), 149.40 (Car), 150.37 (Car), 153.03 (Car), 188.16 (C=O) ppm; Anal. Calcd for C<sub>27</sub>H<sub>28</sub>O<sub>6</sub>: C, 72.30; H, 6.29; Found: C, 72.41; H, 6.38.

### 2.1.2. General procedure for the synthesis of 1-acetyl-4,5-dihydropyrazoles 6a-d

To a solution of 15 mL of glacial acetic acid, (0.016 mol) of 80% hydrazine hydrate, (0.004 mol) of previously synthesized chalcones **5a-d** in absolute ethanol was added and the mixture was stirred and heated under reflux for 2-5 h, the solid precipitated and was filtered and dried. The crude product was purified using column chromatography. The eluent was ethyl acetate: petroleum ether (60-80 °C) (2: 3).

#### 2.1.2.1. 1-(5-(4-(Benzyloxy)-3-methoxyphenyl)-4,5dihydro-3-(3,4-dimethoxyphenyl)pyrazol-1-yl) ethanone (6a)

Off-white crystals; Yield (69%); mp 132-133 °C; IR (KBr) v<sub>max</sub>: 3074, 2928, 1651 cm<sup>-1</sup>; <sup>1</sup>H NMR (400 MHz, DMSO-d<sub>6</sub>) δ: 2.30 (s, 3H, CO-CH<sub>3</sub>), 3.14 (dd, J = 4.0 Hz, J = 12.0 Hz, 1H, H<sub>pyrazoline</sub>), 3.75 (s, 3H, OCH<sub>3</sub>), 3.76-3.78 (m, 1H, H<sub>pyrazoline</sub>), 3.80 (s, 3H, OCH<sub>3</sub>), 3.81 (s, 3H, OCH<sub>3</sub>), 5.04 (s, 2H, OCH<sub>2</sub>), 5.47 (dd, J = 4.0 Hz, J = 8.0 Hz, 1H, H<sub>pyrazoline</sub>), 6.62 (1H, d, J = 8.0 Hz, H<sub>ar</sub>), 6.83 (s, 1H, H<sub>ar</sub>), 6.95 (d, J = 8.0Hz, 1H, H<sub>ar</sub>), 7.01 (d, J = 8.4 Hz, 1H, H<sub>ar</sub>), 7.27-7.33 (m, 2H, H<sub>ar</sub>), 7.36-7.43 (m, 5H, H<sub>ar</sub>) ppm; <sup>13</sup>C NMR (100 MHz, DMSO-d<sub>6</sub>) δ: 21.80 (CO-<u>C</u>H<sub>3</sub>), 42.30 (CH<sub>Pyrazoline</sub>), 55.56 (OCH<sub>3</sub>), 55.62 (OCH<sub>3</sub>), 55.64 (OCH<sub>3</sub>), 59.11 (CH<sub>Pyrazoline</sub>), 70.00 (OCH<sub>2</sub>), 109.11 (CHar), 109.97 (CHar), 111.48 (CHar), 113.72 (CHar), 116.99 (CHar), 120.48 (CHar), 123.81 (CHar), 127.76 (CHar), 127.88 (CHar), 128.47 (CHar), 135.49 (Car), 137.25, 146.96, (Car), 148.82 (Car), 149.21 (Car), 150.85 (C<sub>Pvrazoline</sub>), 154.29 (C<sub>ar</sub>), 167.29 (C=O) ppm; MS (ESI+, m/z): 483.2 (100.0%) [M + Na]+; Anal. Calcd for C<sub>27</sub>H<sub>28</sub>N<sub>2</sub>O<sub>5</sub>: C, 70.42; H, 6.13; N, 6.08; Found: C, 70.58; H, 6.25; N, 6.19.

#### 2.1.2.2. 1-(5-(4-(4-Fluorobenzyloxy)-3-methoxy phenyl)-4,5-dihydro-3-(3,4-dimethoxyphenyl) pyrazol-1-yl)ethanone (6b)

Yellowish brown solid; Yield (65%); mp 82-83 °C; (KBr) v<sub>max</sub>: 3071, 2936, 1659 cm<sup>-1</sup>; <sup>1</sup>H NMR (400 MHz, DMSO-d<sub>6</sub>) δ: 2.30 (s, 3H, CO-CH<sub>3</sub>), 3.14 (dd, J = 4.0 Hz, J = 12.0 Hz, 1H, H<sub>pyrazoline</sub>), 3.75 (s, 3H, OCH<sub>3</sub>), 3.80 (s, 3H, OCH<sub>3</sub>), 3.81 (s, 3H, OCH<sub>3</sub>), 3.84-3.86 (m, 1H, H<sub>pyrazoline</sub>), 5.01 (s, 2H, OCH<sub>2</sub>), 5.47 (dd, J = 4.0 Hz, J = 8.0 Hz, 1H, H<sub>pyrazoline</sub>), 6.63 (d, J = 8.0Hz, 1H, H<sub>ar</sub>), 6.84 (s, 1H, H<sub>ar</sub>), 6.95 (d, J = 8.0 Hz, 1H,  $H_{ar}$ ), 7.00 (d, J = 8.0 Hz, 1H,  $H_{ar}$ ), 7.20 (t, J = 8.4 Hz, 2H,  $H_{ar}$ ), 7.28 (d, J = 8.0 Hz, 1H,  $H_{ar}$ ), 7.36 (s, 1H,  $H_{ar}$ ), 7.45-7.48 (m, 2H, H<sub>ar</sub>) ppm; <sup>13</sup>C NMR (100 MHz, DMSO-d<sub>6</sub>) δ: 21.73 (CO-<u>C</u>H<sub>3</sub>), 42.23 (CH<sub>Pyrazoline</sub>), 55.50 (OCH<sub>3</sub>), 55.57 (OCH<sub>3</sub>), 59.04 (CH<sub>Pyrazoline</sub>), 69.28 (OCH<sub>2</sub>), 109.11 (CH<sub>ar</sub>), 109.95 (CH<sub>ar</sub>), 111.44 (CH<sub>ar</sub>), 113.84 (CH<sub>ar</sub>), 115.09 (d,  ${}^{2}J_{CF} = 21.0$  Hz, <u>C</u>H<sub>ar</sub>- $C_{ar}F$ ), 116.96 (CH<sub>ar</sub>), 120.38 (CH<sub>ar</sub>), 129.82 (d, <sup>3</sup> $J_{CF}$  = 8.0 Hz, CHar-CHar-CarF), 133.42 (Car), 133.45 (Car), 135.59 (Car), 146.78 (Car), 148.77 (Car), 149.20 (Car), 150.79 (C<sub>Pyrazoline</sub>), 154.15 (C<sub>ar</sub>), 160.53 (d,  ${}^{1}J_{CF} =$ 242.0 Hz, C<sub>ar</sub>-F), 167.17 (C=O) ppm; <sup>13</sup>C DEPT-135 (100 MHz, DMSO-d<sub>6</sub>) δ: 21.51 (CO-<u>C</u>H<sub>3</sub>), 42.01 (CH<sub>Pyrazoline</sub>), 55.28 (OCH<sub>3</sub>), 55.36 (OCH<sub>3</sub>), 58.82 (CH<sub>Pyrazoline</sub>), 69.06 (OCH<sub>2</sub>), 108.88 (CH<sub>ar</sub>), 109.73 (CH<sub>ar</sub>), 111.22 (CH<sub>ar</sub>), 113.61 (CH<sub>ar</sub>), 114.87 (d,  ${}^{2}J_{CF} =$ 22.0 Hz, CHar-CarF), 116.73 (CHar), 120.18 (CHar), 129.61 (d,  ${}^{3}J_{CF} = 9.0 \text{ Hz}$ , <u>C</u>H<sub>ar</sub>-CH<sub>ar</sub>-C<sub>ar</sub>F) ppm; Anal.

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Calcd for C<sub>27</sub>H<sub>27</sub>FN<sub>2</sub>O<sub>5</sub>: C, 67.77; H, 5.69; N, 5.85; Found: C, 67.86; H, 5.78; N, 5.96.

#### 2.1.2.3. 1-(5-(4-(Benzyloxy)-3-methoxyphenyl)-4,5dihydro-3-(3,4,5-trimethoxyphenyl)pyrazol-1yl)ethanone (6c)

Brown solid; Yield (64%); mp 68-69 °C; IR (KBr)  $\upsilon_{max}\!\!:$  3063, 2936, 1659 cm^-1;  $^1\!H$  NMR (400 MHz, DMSO-*d*<sub>6</sub>) δ: 2.32 (s, 3H, CO-CH<sub>3</sub>), 3.22 (dd, *J* = 4.0 Hz, J = 12.0 Hz, 1H, H<sub>pyrazoline</sub>), 3.70 (s, 3H, OCH<sub>3</sub>), 3.76 (s, 3H, OCH<sub>3</sub>), 3.77-3.79 (m, 1H, H<sub>pyrazoline</sub>), 3.83 (s, 6H, 2 OCH<sub>3</sub>), 3.87 (s, 3H, OCH<sub>3</sub>,), 5.04 (s, 2H  $OCH_2$ ), 5.49 (dd, J = 4.0 Hz, J = 8.0 Hz, 1H, H<sub>pyrazoline</sub>), 6.63 (d, J = 8.4 Hz, 1H, H<sub>ar</sub>), 6.84 (s, 1H, H<sub>ar</sub>), 6.95 (d, J = 8.4 Hz, 1H, H<sub>ar</sub>), 7.05 (s, 1H, H<sub>ar</sub>), 7.16 (s, 1H, H<sub>ar</sub>), 7.36-7.41 (m, 5H, Har) ppm; <sup>13</sup>C NMR (100 MHz, DMSO-*d*<sub>6</sub>) δ: 21.73 (CO-<u>C</u>H<sub>3</sub>), 42.27 (CH<sub>Pyrazoline</sub>), 55.61 (OCH<sub>3</sub>), 55.99 (OCH<sub>3</sub>), 59.24 (OCH<sub>3</sub>), 60.14 (CH<sub>Pyrazoline</sub>), 69.98 (OCH<sub>2</sub>), 104.11 (CH<sub>ar</sub>), 109.95 (CHar), 113.77 (CHar), 116.94 (CHar), 126.64 (CHar), 127.66 (CHar), 127.79 (Car), 127.86 (CHar), 128.40 (CH<sub>ar</sub>), 128.41 (CH<sub>ar</sub>), 135.43 (C<sub>ar</sub>), 137.22 (C<sub>ar</sub>), 139.43 (Car), 146.94 (Car), 149.22 (CPyrazoline), 153.02 (Car), 154.19 (Car), 167.35 (C=O) ppm; Anal. Calcd for C<sub>28</sub>H<sub>30</sub>N<sub>2</sub>O<sub>6</sub>: C, 68.56; H, 6.16; N, 5.71; Found: C, 68.67; H, 6.27; N, 5.80.

#### 2.1.2.4. 1-(5-(4-(4-Methylbenzyloxy)-3-methoxy phenyl)-4,5-dihydro-3-(3,4,5-trimethoxy phenyl)pyrazol-1-yl)ethanone (6d)

Brown solid; Yield (67%); mp 89 °C; IR (KBr) v<sub>max</sub>: 3001, 2932, 1659 cm<sup>-1</sup>; <sup>1</sup>H NMR (400 MHz, DMSO*d*<sub>6</sub>) δ: 2.29 (s, 3H, CH<sub>3</sub>), 2.30 (s, 3H, CO-CH<sub>3</sub>), 3.13 (dd, J = 4.0 Hz, J = 12.0 Hz, 1H, H<sub>pyrazoline</sub>), 3.74 (s, 3H, OCH<sub>3</sub>), 3.80 (s, 3H, OCH<sub>3</sub>), 3.81 (s, 6H, 2 OCH<sub>3</sub>), 3.85 (d, J = 4.0 Hz, 1H, H<sub>pyrazoline</sub>), 4.98 (s, 2H, OCH<sub>2</sub>), 5.46 (dd, J = 4.0 Hz, J = 8.0 Hz, 1H, H<sub>pyrazoline</sub>), 6.82 (s, 1H, H<sub>ar</sub>), 6.93 (d, J = 8.4 Hz, 1H, H<sub>ar</sub>), 7.01 (d, J =8.4 Hz, 1H, H<sub>ar</sub>), 7.16-7.18 (m, 2H, H<sub>ar</sub>), 7.29-7.35 (m, 4H, H<sub>ar</sub>) ppm; <sup>13</sup>C NMR (100 MHz, DMSO- $d_6$ )  $\delta$ : 20.78 (CH<sub>3</sub>), 21.76 (CO-<u>C</u>H<sub>3</sub>), 42.26 (CH<sub>Pyrazoline</sub>), 55.54 (OCH<sub>3</sub>), 55.61 (OCH<sub>3</sub>), 59.06 (CH<sub>Pyrazoline</sub>), 69.86 (OCH<sub>2</sub>), 109.14 (CH<sub>ar</sub>), 109.98 (CH<sub>ar</sub>), 111.48 (CHar), 113.74 (CHar), 116.97 (CHar), 120.42 (CHar), 123.79 (CHar), 127.80 (CHar), 127.98 (Car), 128.00 (Car), 128.96 (CHar), 134.17 (Car), 135.37 (Car), 137.06 (Car), 146.95 (Car), 148.80 (Car), 149.19 (Car), 150.82 (C<sub>Pyrazoline</sub>), 154.21 (C<sub>ar</sub>), 167.22 (C=O) ppm; Anal. Calcd for C<sub>29</sub>H<sub>32</sub>N<sub>2</sub>O<sub>6</sub>: C, 69.03; H, 6.39; N, 5.55; Found: C, 69.18; H, 6.47; N, 5.67.

#### 2.2. Biology:

The Medical Research Ethics Committee at National Research Centre - Egypt has approved this study with Ethical Approval Number: **19280**.

### 2.2.1. Anticancer activity: 2.2.1.1. Cell culture:

Two human hepatoma cell lines (HuH-7, and HepG-2) and a normal hepatocyte cell line (BNL) (Nawah Scientific Inc., Cairo, Egypt) were grown in Dulbecco's Modified Eagle Medium (DMEM) media supplemented with streptomycin (100 mg/mL), penicillin (100 units/mL), and heat-inactivated fetal bovine serum (10% FBS) in dampened atmosphere with 5% CO<sub>2</sub> at 37 °C [39].

#### 2.2.1.2. Cytotoxicity assay:

Cell suspension (100 µL of 4 x10^3 cells /well) was seeded in 96-well plates, incubated overnight, and treated with serial dilutions of test compounds (6a-d) for 72 hours. Then, cells were fixed by adding trichloroacetic acid (150 µL of 10% TCA) for one hour at 4°C. To dissolve the protein-bound sulforhodamine B (SRB) stain, we added 150 µL of tris base (10 mM) and measured absorbance at 540 nm with a microplate reader (FLUOstar Omega, Ortenberg, Germany). IC50 values are measured as mean  $\pm$  SD. The selectivity index the (SI), cytotoxic selectivity for the proposed treatments, was reported as  $SI = IC_{50}$  of normal cells /  $IC_{50}$  of cancer cells [40].

#### 2.2.2. NCI-60 anticancer screening:

All the newly synthesized *N*-acetylpyrazolines **6a-d** were selected and screened by the Developmental Therapeutic Program (DTP) at the National Cancer Institute (NCI) in Bethesda, Maryland, US [41-48]. The anticancer screening results were reported as growth inhibition percentage (GI) % at 10  $\mu$ M.

#### 2.2.3. Tubulin polymerization assay:

Tubulin polymerization assay kit (Cat. BK006P, Cytoskeleton, Inc., US) was used, and the experimental protocol was done according to manufacturer [49]. Paclitaxel, a microtubules stabilizing agent, and combrestatin A-4 (CA-4), a microtubules destabilizing agent, were purchased from Sigma Aldrich, and used as positive controls. The direct effect of compounds **6a-d** on tubulin polymerization was determined in an *in vitro* fluorescent-based assay (Cytoskeleton, Denver, CO). Briefly, in 96-well plates tubulin reaction mix (2 mg/ml porcine brain tubulin in 80 nM Pipes, pH 6.9, 2

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nM MgCl<sub>2</sub>, 0.5 mM EGTA, 1 mM GTP and 15% glycerol) were used to dilute each 10x compounds. Fluorescence changes were monitored kinetically in the Cytation 3 Multi-Mode Reader (Biotek, Canada) at 37 °C for 1 h, one read per minute, using excitation at 360 nm and emission at 420 nm.

#### 2.3. Molecular docking study:

Molecular docking simulations were performed using Molecular Operating Environment software (MOE, 2022.02), using high-resolution cryo-EM reconstruction of Taxol-stabilized microtubule (PDB ID: 5SYF) [50, 51]. The detailed docking protocol was reported in supporting information.

## 2.4. Prediction of physicochemical and ADME properties as well as drug-likeness and medicinal chemistry friendliness:

*SwissADME* online web tool was employed to predict the physicochemical and ADME properties as well as drug-likeness and medicinal chemistry friendliness of the target compounds **6a-d**. The compounds' SMILES were generated utilizing (MOE, 2022.02) software then they were submitted to *SwissADME* [52-54].

### 3. Results and Discussion 3.1. Chemistry:

The target compounds 6a-d were synthesized via the reaction of the previously synthesized *p*-benzyloxy-3methoxybenzaldehydes 3a-c according to reported method [55-57] and 3',4'-dimethoxyacetophenone 4a or 3',4',5'-trimethoxyacetophenone 4b through Claisen-Shmidt condensation reaction forming the novel chalcones 5a-d using 50% aqueous potassium hydroxide. The synthesized chalcones 5a-d was cyclized using hydrazine hydrate in the presence of glacial acetic acid forming the novel Nacetylpyrazolines 6a-d (Scheme 1). Chalcones 5a-d were found in (E) configuration based on the coupling constant (J = 12-16 Hz) for the  $\alpha$ . $\beta$ -unsaturated ketone  $^{13}C$ DEPT-135 protons. The (Distortionless Enhancement by Polarisation Transfer) spectrum of the *N*-acetylpyrazoline derivative **6b** was performed to confirm the presence of the methylene group of the 4,5-dihydropyrazole ring that resonated at 42.01 ppm, whereas, the OCH<sub>2</sub> group at 69.06 ppm in the negative/inverted mode, while, the methyl and methine carbons resonated in the positive/upright

mode (Figure 3). For <sup>1</sup>H and <sup>13</sup>C NMR, and mass spectral data of the synthesized chalcones **5a-d**, and *N*-acetylpyrazolines **6a-d**, see supporting information (Figures S.1-S.19).





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#### **3.2. Biology**

### **3.2.1.** Anticancer activity against HuH-7, HepG-2, and BNL cell lines:

The cytotoxic activity of compounds **6a-d** was first evaluated against liver cancer cells (HuH-7, and HepG-2) and normal cells (BNL) at two concentrations (10 and 100  $\mu$ M). Growth inhibition percentage (GI%) was calculated and compared to paclitaxel, colchicine, and CA-4 as standards (**Table 1**).

Compounds 6a-d inhibited growth in HuH-7 liver cancer cells with GI% values of 12.1-39.4% and 65.2-82.9% at 10 and 100 µM, respectively. However, they had lower GI% values against HepG-2 cells at both (6.0-22.3%) concentrations and 36.7-51.3%), indicating that compounds 6a-d had more potent anticancer activity against the HuH-7 cell line than the HepG-2 cell line. Furthermore, the four synthesized compounds **6a-d** were further tested for  $IC_{50}$ determination (Table 2). The results depicted that the 3',4',5'-trimethoxyphenyl N-acetylpyrazolines (6c and 6d) showed more potent anticancer activity than the 3',4'-dimethoxyphenyl pyrazolines (6a and 6b) with IC<sub>50</sub> values of 11.00 and 0.30 µM vs 50.20 and 14.50 µM, respectively against the HuH-7 cell line. Regarding HepG-2 the cell line, only compounds **6b** and **6d** showed weak to moderate anticancer activities with IC50 values of 90.00 and 77.30 µM, respectively. While compounds **6a** and **6c** didn't show noticeable cytotoxicity till the concentration of 100 µM. Fortunately, all the tested compounds **6a-d** showed IC<sub>50</sub> values of more than 100 µM against the normal mouse BNL liver cell line and their selectivity index values were calculated and presented in Table 2, confirming the active targeting power of the pyrazoline compounds against the cancerous cells while providing the safety of normal cells [58, 59]. The tested compounds displayed the highest selectivity and potency towards HuH-7 cells rather than HepG-2 cells. Among them, compound 6d was the safest compound (SI = > 333 and 1.29 for HuH-7 and HepG-2, respectively). The IC<sub>50</sub> graphs for compounds **6b-d** are inserted in supporting information (Figures S.20-S.22).

Table 1

Growth inhibition (GI) % of compounds 6a-d against liver cancer and normal cell lines at 10 and 100 µM.

	HuH-7		HepG-2		BNL	
Compound ID			GI%	, 0		
	10 µM	100 µM	10 µM	100 µM	10 µM	100 µM
6a	12.1	65.4	6.0	6.98	1.45	4.52
6b	39.4	82.9	22.3	54.7	2.5	6.5
6с	33.8	89.2	5.31	8.65	0.94	2.87
6d	88.8	99.4	17.3	64.0	5.21	7.59
Paclitaxel	81.0	99.91	88.85	97.2	97.4	100.9
Colchicine	89.4	99.63	56.33	93.8	90.3	100.1
CA-4	80.0	99.0	86.22	98.4	88.5	99.99

Table 2

IC50 values of compounds 6a-d against HepG-2, HuH-7, and BNL cell lines and the selectivity index (SI).

Compound ID	HuH-7	HepG-2	BNL	HuH-7	HepG-2
		IC <sub>50</sub> (µM)		SI	[
6a	50.20	>100	>100	> 1.99	> 1
6b	14.50	90.00	>100	> 6.89	> 1.1
6с	11.00	>100	>100	> 9.09	> 1
6d	0.30	77.30	>100	> 333	1.29
Paclitaxel	0.02	1.10	0.001	0.05	0.001
Colchicine	0.056	6.61	0.014	0.25	0.002
CA-4	0.026	1.47	0.499	19.19	0.339

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#### 3.2.2. NCI-60 anticancer screening:

All the newly synthesized *N*-acetylpyrazolines **6a-d** were screened for their possible anticancer activity by the National Cancer Institute (NCI - US) against a cancer cell lines panel *via* a sulforhodamine B (SRB) assay.

Table 3 presents the growth inhibition (GI) percentage of the target compounds 6a-d at 10 µM. The one-dose mean graphs were presented in supporting information (Figures S.23-S.26). The N-acetylpyrazolines 6a-d showed mean GI% values of 8.87, 46.54, 64.54, and 54.63%, respectively. The 3',4'-dimethoxyphenyl analogue 6a showed the lowest anticancer activity among the synthesized N-acetylpyrazolines with GI% of 8.87%, while, it revealed a moderate anticancer activity against lung (EKVX, and NCI-H522), colon (HCT-15), melanoma (UACC-62), renal (UO-31), and breast (MCF-7, and T-47D) cancer cell lines with GI% values 31.83, 40.68, 30.09, 36.80, 38.14, 32.39, and 39.94%, respectively. The N-acetylpyrazolines 6b-d showed moderate to high and broad anticancer activities against leukemia (K-562, and MOLT-4), lung (A-549, HOP-92, and NCI-H522), colon (COLO 205, HCT-116, HCT-15, and HT 29), CNS (SF-295), melanoma (SK-MEL-5), ovarian (OVCAR-4), renal (UO-31), prostate (PC-3), and breast (MCF-7, HS 578T, BT-549, T-47D, and MDA-MB-468) cancer cell lines with GI% ranges (69.04-91.21%), (49.24-104.80%), (41.53-89.09%), (62.17-75.38%), (70.31-109.52%), (57.92-77.71%), (70.26-91.37%), (72.34-90.16), and (39.94-97.30%), respectively. Moreover, the 3',4',5'-trimethoxyphenyl N-acetylpyrazoline 6c, and 6d revealed not only potent anticancer activities but also nearly complete lethal effects against lung cancer cell line (HOP-92) for 6c and melanoma cell line (SK-MEL-5) for 6d with GI% values of 104.80, and 109.52%, respectively.

#### 3.2.3. Tubulin polymerization assay:

All the synthesized *N*-acetylpyazolines **6a-d** were tested at 50  $\mu$ M on tubulin polymerization assay, and the emmission used at 420 nm for 1 h. Paclitaxel, a tubulin stabilizing agent (10  $\mu$ M), and CA-4 (10  $\mu$ M), a tubulin destabilizing agent, were used as positive controls for the tubulin polymerization assay. The *N*acetylpyrazolines **6a-d** showed tubulin-stabilizing effects as paclitaxel at 50  $\mu$ M (Figures 4, and 5). The 3',4',5'-trimethoxyphenyl *N*-acetylpyrazoline **6d** was further tested for EC<sub>50</sub> ( $\mu$ M) values determination in

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comarison to paclitaxel, and it was found its  $EC_{50}$  value > 100  $\mu$ M (Figure 6).



Figure 4: Microtubule-stabilizing effects of compounds 6a-d at 50  $\mu$ M vs paclitaxel, and CA-4 at 10  $\mu$ M.



Figure 5: Enhanced tubulin polymermization for compounds 6a-d at 50  $\mu M.$ 



Figure 6: EC<sub>50</sub> graph of compound 6d on tubulin polymerization

#### Table 3

Growth inhibition (GI) % of compounds 6a-d against a panel of cancer cell lines at 10  $\mu M.$ 

					Compound ID				
Subpanel of Cancer Cell Lines	6a	6b	6c	6d		6a	6b	6c	6d
					GI%				
Leukemia	-				Melanoma Cont.	-			
CCRF-CEM	nd <sup>a</sup>	40.11	59.84	49.93	MDA-MB-435	-	37.52	55.43	56.95
HL-60(1B)	nd	62.68	/8.49	65.37	SK-MEL-2	5.38	30.49	/8./4	55.88
K-562	na	72.04	81.36	69.04	SK-MEL-28	-	26.77	57.90	43.24
MOLT-4	nd	69.37	91.21	/9.51	SK-MEL-5	-	/0.31	90.23	109.52
PRMI-8226	5.96	63.43	75.83	63.02	UACC-257	-	31.38	51.22	59.85
SR	nd	60.34	67.93	47.61	UACC-62	36.80	47.96	75.95	65.96
Non-Small Cell Lung Cancer	25.00	40.24	60.10	70.07	Ovarian Cancer	24.44	54.10	57.07	51.02
A549/ATTC	25.08	49.24	68.18	/0.9/	IGROVI	24.44	54.19	57.07	51.03
EKVX	31.83	40.55	67.37	53.72	OVCAR-3	-	29.69	59.03	58.58
HOP-62	_0	25.78	29.77	23.77	OVCAR-4		57.92	77.71	70.62
HOP-92	11.47	82.67	104.80	62.88	OVCAR-5		42.22	40.15	20.16
NCI-H226	-	50.18	72.02	50.06	OVCAR-8	19.41	38.07	53.32	56.51
NCI-H23	-	43.35	52.81	41.95	NCI/ADR-RES	-	31.36	61.56	43.90
NCI-H322M	8.33	33.88	40.21	18.14	SK-OV-3	-	48.05	28.72	15.06
NCI-H460	-	50.97	75.73	58.72	Renal Cancer				
NCI-H522	40.68	57.91	73.16	71.47	786-0	5.47	42.24	66.03	42.11
Colon Cancer					A498	-	40.61	44.91	35.09
COLO 205	<sup>c</sup>	41.53	89.09	58.70	ACHN	10.68	40.80	71.80	57.12
HCC-2998	-	13.98	28.76	16.63	CAKI-1	19.33	46.78	70.69	52.83
HCT-116	22.18	68.50	75.69	64.69	RXF 393	-	26.33	68.62	47.05
HCT-15	30.09	57.51	65.42	72.09	SN 12C	22.36	46.10	58.93	56.84
HT29	23.64	71.90	61.69	59.94	TK-10	8.67	49.49	67.91	55.94
KM12	12.70	42.85	66.11	61.01	UO-31	38.14	70.26	91.37	77.61
SW-620	-	41.46	44.91	38.80	Prostate Cancer				
CNS Cancer					PC-3	18.61	72.34	90.16	76.39
SF-268	17.22	28.04	52.21	52.89	DU-145	-	29.37	59.56	48.71
SF-295	22.94	62.35	75.38	62.17	Breast Cancer				
SF-539	7.46	38.12	48.30	36.23	MCF7	32.39	65.33	82.89	79.86
SNB-19	19.71	39.14	47.82	48.59	MDA-MB- 231/ATTC	15.32	39.00	35.98	30.41
SNB-75	-	20.51	50.68	38.73	HS 578T		41.22	58.98	43.74
U251	9.74	40.10	55.69	53.22	BT-549	nd	55.12	82.62	66.20
Melanoma					T-47D	39.94	39.94	75.17	97.30
MALME-3M		18.16	56.88	51.92	MDA-MB-468	6.14	52.90	65.76	68.93
M14		40.13	63.09	45.78					
Mean GI %	8.87	46.54	64.77	54.63					

a: Not determined.
b: No GI% detected at 10 μM.
c: GI% lower than 5%.

#### 3.3. Molecular docking study:

Molecular dockings simulations were performed to investigate the binding pattern of the newly synthesized compounds 6a-d at the Taxol-binding-site of the microtubule and to explain their promising microtubule-stabilizing activity. In the current study, high-resolution cryo-EM reconstruction of Taxolstabilized microtubule (PDB ID: 5SYF) was utilized to perform the molecular docking simulations [50, 51]. Primarily, self-docking validation was carried out to validate the used docking protocol by docking Taxol at its binding site on the microtubule. The self-docking validation step accurately regenerated the interaction pattern of Taxol in its binding site showing that the used docking setup is appropriate for the intended docking study. This is demonstrated by the small RMSD value between the co-crystalized inhibitor and its docking pose (1.550Å) and by the capability of the attained pose to duplicate the key interactions achieved by the co-crystalized pose with the residues His229, Thr276, and Arg369 (Figure 7 and 8). In its binding site, Taxol interacts through Van der Waals interactions by its 3'-benzoyl moiety with  $\beta$ -tubulin His229. Furthermore, through hydrogen bonding, it interacts by its 3'-oxygen, 2'-OH, and oxetane ring oxygen with His229 nitrogen, Arg369 backbone carbonyl, and Thr276 (in strand S7) backbone NH, respectively (Figure 7 and 8).

Compared to the natural ligand, Nthe acetylpyrazoline derivatives 6a-d showed a predicted docking energy score (S) range of -9.97 to -10.49 kcal/mol (Table 4). The tested compounds 6a-d showed a comparable predicted binding affinity which agrees with their experimental activity. Compounds 6a-d showed a common binding pattern in Taxolbinding-site interacting with the key amino acid Thr276 (in strand S7) (Figures 9-12). The binding pattern involves the fitting of the N-acetylpyrazoline ring in the Taxol's oxetane ring region achieving Hbond interaction with the key amino acid Thr276. On one side directing the hydrophobic (un)substituted benzyloxy phenyl moiety, in the region of Taxol's 3'phenyl moiety, deeply towards the hydrophobic subpacket in the tubulin beta subunit lined by the hydrophobic side chains of the amino acids Val23, Ala233, Phe272, Pro274, Pro360, and Leu371. On the other side directing the (di)(tri)methoxyphenyl moiety towards Arg278 interacting via hydrophobic interaction with Leu217 and Leu219 of the side chains (Figures 9-12).

#### Table 4

Docking scores (*S*) in kcal/mol for the *N*-acetylpyrazolines **6a-d** and Taxol in microtubule binding sites.

Compound ID	Docking score (S) (kcal/mol)			
Paclitaxel	-12.75			
6a	-10.49			
6b	-10.11			
6с	-9.97			
6d	-10.12			



**Figure 7**: 2D interaction diagram showing Taxol docking pose interactions with the key amino acids in Taxol-binding-site on the microtubule (Distance in Å).



**Figure 8**: 2D Superimposition diagram of the co-crystallized (red) and the docking pose (green) of Taxol in Taxol-binding-site on the microtubule with RMSD of 1.550Å.





**Figure 9**: 2D interaction diagram (A) and 3D representation (B) showing interactions of compound **6a** docking pose at the Taxol site on the microtubule. (Distances in Å)



(B) Figure 10: 2D interaction diagram (A) and 3D representation (B) showing interactions of compound **6b** docking pose at the Taxol site on the microtubule. (Distances in Å)





(A)



**Figure 11**: 2D interaction diagram (A) and 3D representation (B) showing interactions of compound **6c** docking pose at the Taxol site on the microtubule. (Distances in Å)

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**(B)** 

Figure 12: 2D interaction diagram (A) and 3D representation (B) showing interactions of compound **6d** docking pose at the Taxol site on the microtubule. (Distances in Å)

# **3.4.** Prediction of physicochemical and ADME properties as well as drug-likeness and medicinal chemistry friendliness of compounds 6a-d:

The promising activity of the *N*-acetylpyrazolines **6ad** encouraged us to further predict their physicochemical and ADME properties as well as their drug-likeness and medicinal chemistry friendliness. As a result, the *SwissADME* web tool provided by the Swiss Institute of Bioinformatics (SIB) was utilized [52-54].

Table 5

SwissADME predicted physicochemical and drug-likeness properties.

As can be seen in table 5, compounds 6a-d exhibited promising physicochemical properties displaying molecular weight range of 460.52 - 504.57 Da, rotatable bond number of 9 or 10, topological polar surface area (TPSA) of 69.59 or 78.52 Å<sup>2</sup> which is less than the cutoff for oral bioavailability (140  $Å^2$ ) [60] and a predicted consensus logP range of 4.00-4.35. Furthermore, the target compounds predicted to be moderately water soluble according to log S model from Ali J. et al., [61]. For further details see supporting materials. Predicted ADME properties showed that the target compounds 6a-d are highly GIT absorbable. Furthermore, compounds 6c-d showed no predicted BBB permeation and so with no predicted CNS side effects, however, 6a-b analogues predicted to be able to pass BBB. All compounds were predicted to be P-gp substrates (For further details see supporting materials).

As for their drug-likeness, the target compounds adhered to Lipinski's rule of five, with at most one violation (**Table 5**). Additionally, they exhibited a favourable Abbott bioavailability score of 0.55 [62]. All the target compounds showed a promising medicinal chemistry friendliness with no pan assay interference compounds (PAINS) alerts [63] in their structures and with reasonable synthetic accessibility ranging 4.22 - 4.53 (1 is easy to synthesize and 10 is difficult) (For further details see supporting materials).

Compound ID	MW	#Rot. bond	#H-bond acc.	#H-bond don.	TPSA	Cons. LogP	#Lipinski's violations
6a	460.52	9	6	0	69.59	4.02	0
6b	478.51	9	7	0	69.59	4.35	0
6с	490.55	10	7	0	78.82	4.00	0
6d	504.57	10	7	0	78.82	4.34	1

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#### 4. Conclusion

New N-acetylpyrazoline derivatives 6a-d were synthesized via the reaction of corresponding chalcones 5a-d and hydrazine hydrate in acidic medium. The synthesized pyrazolines were evaluated in vitro for their anticancer activity against liver (HuH-7, and HepG-2) cancer and (BNL) normal cell lines. The *N*-acetylpyrazolines **6b-d** revealed potent anticancer activity against HuH-7 cell line  $IC_{50} =$ 14.50, 11.00, and 0.30 µM, respectively. Compounds 6a-d were further screened for their anticancer potential against different cancer cell lines at 10 µM, and the 3',4',5'-trimethoxyphenyl N-acetylpyrazoline 6d revealed potent anticancer activities mean GI% (64.54%). Moreover, the N-acetylpyrazolines 6a-d were tested for their effects on tubulin polymerization at 50 µM compared to paclitaxel, and CA-4 as standards, whereas, they showed tubulin stabilizing effects similar to paclitaxel. A molecular docking study was performed to investigate the binding pattern of the Nacetylpyrazolines 6a-d at the Taxol-binding site of microtubules. A predicted ADME properties study showed that compounds **6a-d** are highly GIT absorbable without predicted BBB permeation and so with no predicted CNS side effects.

#### 5. Conflicts of interest

"There are no conflicts to declare".

#### 6. Formatting of funding sources

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